



**UNIVERSIDADE FEDERAL DO RIO GRANDE – FURG
PROGRAMA DE PÓS-GRADUAÇÃO EM AQUICULTURA**



**Efluentes da Carcinocultura e seus Efeitos sobre o Sistema
Estuarino da Lagoa dos Patos**

WILLIAM BAUER

**FURG
RIO GRANDE – RS
2015**

UNIVERSIDADE FEDERAL DE RIO GRANDE
PROGRAMA DE PÓS-GRADUAÇÃO EM AQUICULTURA

**Efluentes da Carcinocultura e seus Efeitos sobre o Sistema
Estuarino da Lagoa dos Patos**

WILLIAM BAUER

Tese apresentada ao programa de Pós-graduação em
Aquicultura da Universidade Federal do Rio Grande –
FURG, como requisito parcial à obtenção do título de
Doutor.

Orientador: Dr. Luis Henrique da Silva Poersch

Rio Grande, RS
Junho/2015

DEDICATÓRIA

A Deus, razão do meu ser

“Conheço teu medo, a tua felicidade e os teus sonhos. Conheço tua estrada
e sei exatamente o teu destino...

E sem que tu tenhas que me pedir, eu entendo o que tu queres. Conheço o
teu sorriso, e sei tudo que está dentro do teu coração. Conheço e te
reconheço em qualquer lugar...

Sei do teu amor, da tua saudade, dos sonhos que movimentam a tua vida e
da esperança que te faz lutar...

Acompanho-te desde sempre! Estou ao teu lado mesmo quando pensas que
te abandonei...

Conheço-te, porque eu te criei.”

AGRADECIMENTOS

Ao CNPq, pela bolsa concedida nos primeiros 18 meses de doutorado e à Capes, pela bolsa concedida no tempo restante e pela bolsa de doutorado-sanduíche;

Ao meu orientador Dr. Luis Poersch, Mineiro, pela sua amizade e toda atenção e apoio ao longo dessa caminhada;

Ao meu orientador de doutorado-sanduíche Dr. Michael Schwarz e a toda sua equipe pela oportunidade de trabalhar e pelo aprendizado adquirido no Virginia Seafood Agricultural Research and Extension Center;

Aos colegas e amigos da EMA pelas conversas e por todo apoio e ajuda nas saídas de campo, juntos mesmo nas madrugadas de inverno;

À Bruna, sempre presente nas coletas e ajudando no processamento das amostras;

Ao Raphael Pinotti do Laboratório de Ecologia de Invertebrados Macrobióticos pela ajuda incansável;

À professora Monica Wallner-Kersanach do Laboratório de Hidroquímica por todas as nossas conversas e ajuda ao longo da extensa caminhada durante as análises de carbono, nitrogênio e metais;

Ao professor Paulo Abreu pela oportunidade de realizar a identificação planctônica e de clorofila *a* no Laboratório de Ecologia de Fitoplâncton e Microorganismo Marinhos;

Ao Alessandro Cardozo, pelo auxílio na identificação do zooplâncton;

Aos meus pais e toda minha família pelo apoio;

À Angélica, minha eterna companheira de todas as horas, pela paciência e ajuda durante todos esses anos;

A todos os meus amigos.

ÍNDICE

RESUMO GERAL	6
GENERAL ABSTRACT	9
INTRODUÇÃO GERAL	11
<i>CRESCIMENTO DA AQUICULTURA</i>	<i>11</i>
<i>EFEITO DO ENRIQUECIMENTO ORGÂNICO SOBRE O SEDIMENTO</i>	<i>13</i>
<i>EFEITOS SOBRE A BIODIVERSIDADE</i>	<i>14</i>
<i>LOCAL DE ESTUDO</i>	<i>16</i>
OBJETIVO GERAL	18
<i>OBJETIVOS ESPECÍFICOS</i>	<i>18</i>
REFERÊNCIAS BIBLIOGRÁFICAS	19
CAPÍTULO 1	26
<i>VARIABILIDADE DO PLÂNCTON E DA QUALIDADE DA ÁGUA EM UM ESTUÁRIO ANTES E APÓS O LANÇAMENTO DE EFLUENTES DA CARCINOCULTURA: IMPACTOS E REGENERAÇÃO</i>	<i>26</i>
CAPÍTULO 2	51
<i>IMPACTO DOS EFLUENTES DA CARCINOCULTURA SOBRE A COMUNIDADE MACROZOOBENTÔNICA DA LAGOA DOS PATOS, BRASIL</i>	<i>51</i>
CAPÍTULO 3	69
<i>IMPACTOS DOS EFLUENTES DA CARCINOCULTURA SOBRE O SEDIMENTO</i>	<i>69</i>
DISCUSSÃO GERAL	88
REFERÊNCIAS BIBLIOGRÁFICAS	95
ANEXO	100

1 **RESUMO GERAL**

2
3 A aquicultura é o setor de produção de alimentos de origem animal que mais cresce no
4 mundo. A carcinocultura, como parte desse setor, tem gerado diversos benefícios em
5 termos sócio-econômicos, mas também tem gerado uma atenção especial aos aspectos
6 ambientais envolvidos com a atividade. A produção em cativeiro do camarão branco do
7 Pacífico *Litopenaeus vannamei* vem se consolidando cada vez mais no entorno do
8 estuário da Lagoa dos Patos, região sul do Brasil. Esse estuário possui importância
9 ecológica, industrial, portuária, agrícola e pesqueira, o que torna necessário a
10 compreensão do potencial impacto poluidor da atividade nessa região. Nesse estudo,
11 parâmetros de qualidade da água, composição e abundância do fitoplâncton,
12 protozooplâncton, mesozooplâncton e macrozoobentos foram avaliados espaço-
13 temporalmente no corpo d'água receptor dos efluentes de uma fazenda produtora de
14 camarões da espécie *Litopenaeus vannamei* localizada as margens do estuário da Lagoa
15 dos Patos. Foram avaliados ainda os teores de carbono orgânico total (COT), nitrogênio
16 total (NT), Cobre (Cu) e Zinco (Zn) do sedimento do viveiro de cultivo, da bacia de
17 sedimentação e do estuário receptor dos efluentes. Foram avaliadas ainda as
18 concentrações de Cu e Zn do tecido dos camarões cultivados ao final do ciclo produtivo.
19 As amostras foram tomadas antes da descarga dos efluentes (AD) e; 1 dia (1 PD), 5 dias
20 (5 PD), 10 dias (10 PD), 20 dias (20 PD), 30 dias (30 PD), 60 dias (60 PD) e 90 dias (90
21 PD) após a descarga dos efluentes. Vale ressaltar que cada estudo contou com um
22 cronograma próprio e assim nem todas as análises citadas acima tiveram coletas em
23 todas as datas mencionadas. Especificamente para a avaliação do macrozoobentos, um
24 segundo ciclo de produção foi acompanhado para realização de amostragens. Os pontos
25 de coleta no estuário distribuíram-se desde a desembocadura do canal de lançamento
26 dos efluentes até uma distância de 250 m do mesmo. Dados de temperatura, oxigênio
27 dissolvido, pH, salinidade, clorofila *a*, nitrogênio amoniacal total (NAT), nitrito, nitrato,
28 fósforo total, sólidos suspensos totais e turbidez da água foram mensurados. Além da
29 temperatura e salinidade que sofrem variações sazonais em ambientes estuarinos, NAT,
30 clorofila *a* e turbidez foram os parâmetros de qualidade de água que sofreram alterações
31 mais marcadas em decorrência do lançamento dos efluentes. No entanto, de maneira
32 geral, essas alterações foram restritas a uma distância de até 20 m do canal de descarga
33 dos efluentes por um período de tempo de no máximo 5 dias. Diatomáceas,
34 cianobactérias, cloroficeas e ciliados foram identificados e quantificados. A comunidade

1 predominante foi a de clorofíceas, seguido das diatomáceas, cianobactérias e ciliados.
2 Houve um aumento na concentração dos diferentes grupos no primeiro dia pós-descarga
3 (1 PD). Entretanto, esse aumento pode estar relacionado ao enriquecimento de
4 nutrientes das águas estuarinas ocasionado por chuvas abundantes, e não propriamente
5 ao despejo dos efluentes. A abundância meso-zooplanctônica foi baixa e representada
6 exclusivamente por copépodos, na sua grande maioria da espécie *Acartia tonsa*. A única
7 exceção foi a amostragem 30 PD que sofreu um acréscimo expressivo, sem no entanto
8 apresentar relação com os parâmetros abióticos avaliados. Apesar de diferenças
9 significativas ($p < 0,05$) terem sido observadas em algumas amostragens, essas não
10 aparentam ter nenhuma relação com o lançamento dos efluentes. Sete grupos
11 macrozoobentônicos foram observados (Polychaeta, Tanaidacea, Isopoda, Gastropoda,
12 Bivalvia, Malacostraca e Ostracoda) ao longo de dois ciclos produtivos (2012 e 2013).
13 A densidade e a riqueza de espécies sofreram pequena variabilidade ao longo dos
14 pontos amostrais do estuário para ambas as etapas. Já ao longo do tempo, em 2012
15 pode-se observar um aumento desses índices no inverno comparativamente ao verão.
16 Ao longo das coletas de 2013, a densidade do macrobentos teve um padrão oposto e a
17 riqueza de espécies sofreu pouca variabilidade. A concentração de COT variou de 0,12
18 a 0,67% e a concentração de NT ficou abaixo do limite de detecção do equipamento
19 ($<0,1$ ppm) em todos os locais e campanhas amostrais. Os teores de Cu e Zn no
20 sedimento foram quantificados através de extração fraca (fração de metais lábeis ou
21 potencialmente biodisponível) e extração semi-forte (fração de metais mais fortemente
22 adsorvida). O Cu na fração lábil variou de 0,12 a 1,27 $\mu\text{g/g}$ e o Zn variou de 0,52 a 3
23 $\mu\text{g/g}$, enquanto que a fração mais fortemente adsorvida de Cu variou de 0,3 a 2,65 $\mu\text{g/g}$
24 e o Zn de 30,44 a 121,4 $\mu\text{g/g}$. Diferenças significativas (ANOVA, $p < 0,05$) foram
25 observadas nos resultados de COT entre alguns pontos amostrais nas campanhas 1PD e
26 10PD, sem no entanto haver relação com o lançamento dos efluentes. A análise de
27 correlação de Pearson não mostrou relação entre o lançamento dos efluentes e aumento
28 nos valores de COT, NT, Cu ou Zn no sedimento, com exceção da relação encontrada
29 entre os valores de COT e Cu e Zn no sedimento no primeiro dia pós-descarga (1 PD).
30 Ao final do ciclo, exemplares de camarões coletados de dois viveiros de engorda
31 revelaram concentrações médias de $6,63 \pm 0,2$ $\mu\text{g/g}$ de Cu e $19,76 \pm 0,2$ $\mu\text{g/g}$ de Zn no
32 viveiro 1, e $7,6 \pm 0,51$ $\mu\text{g/g}$ de Cu e $19,13 \pm 0,32$ $\mu\text{g/g}$ de Zn no viveiro 2. Apesar de
33 alguns parâmetros de qualidade de água terem sofrido variações em decorrência do
34 lançamento dos efluentes, esses efeitos foram agudos e pontuais, restritos a uma

1 distância de até 20m da margem e observados até os 5 dias após o descarte dos
2 efluentes. Além disso, os valores ficaram dentro da faixa exigida pela Resolução
3 ambiental vigente. O fitoplâncton, protozooplâncton, mesozooplâncton e o
4 macrozoobentos sofreram apenas variações decorrentes das oscilações naturais dos
5 parâmetros abióticos que ocorrem em sistemas estuarinos. Por fim, os teores de COT,
6 NT, Cu e Zn no sedimento e Cu e Zn no tecido dos camarões ficaram dentro dos valores
7 máximos estipulados na legislação, mostrando que o lançamento dos efluentes
8 provenientes da produção de camarões no entorno do estuário da Lagoa dos Patos não
9 trouxe efeitos adversos ao meio ambiente.

10

1 GENERAL ABSTRACT

2

3 Aquaculture is the food producing sector of higher growth worldwide. The shrimp
4 farming, as part of this sector, has generated several socio-economic benefits. However,
5 it has also developed particular attention to environmental aspects involved with the
6 activity. The production of Pacific white shrimp *Litopenaeus vannamei* is increasing in
7 areas adjacent to Patos Lagoon estuary, Southern Brazil. This estuary has importance on
8 several areas as ecology, industry, ports, agriculture and fisheries, which makes the
9 understanding of the potential pollution impact of the shrimp farm in this ecosystem
10 extremely necessary. In this study, the water quality, composition and abundance of
11 phytoplankton, protozooplankton, meso-zooplankton and macrozoobenthos were spatio-
12 temporally evaluated in the Patos Lagoon estuary, which received effluents from a
13 commercial shrimp farm. Total organic carbon (TOC), total nitrogen (TN), copper (Cu)
14 and zinc (Zn) content in sediment of pond, sedimentation basin and the estuary were
15 also analyzed. Moreover, concentrations of Cu and Zn in shrimp tissues were measured
16 at the end of the growth out cycle. Samples were taken before the effluents discharge
17 (BD) and; 1 day (1 PD) 5 days (5 PD), 10 days (10 PD), 20 days (20 PD), 30 days (30
18 PD), 60 days (60 PD) and 90 days (90 PD) after the effluents discharge. Only for the
19 macrozoobenthos analysis, samples were collected in two cycle's period. The sampling
20 sites in the estuary were distributed across the effluents discharge channel (EDC), 20m,
21 30m, 100m and 250m from the channel. Temperature, dissolved oxygen, pH, salinity,
22 chlorophyll a, total ammonia nitrogen (TAN), nitrite, nitrate, total phosphorus, total
23 suspended solids and turbidity were measured. Temperature and salinity suffered
24 seasonal influences and TAN, chlorophyll a and turbidity showed the most marked
25 changes because of the effluents discharge. However, these changes were restricted to a
26 distance of 20 m from the EDC for a period of 5 days. Diatoms, cyanobacteria,
27 chlorophyceae and ciliates were identified and quantified. The predominant community
28 was chlorophytes, followed by diatoms, cyanobacteria and ciliates. There was an
29 increase in the abundance of different groups on the 1 PD sampling. However, this may
30 be related to nutrient enrichment of estuarine waters caused by rains, and not necessarily
31 to the effluents discharge. The meso-zooplankton abundance was low and was
32 represented exclusively by copepods, mostly *Acartia tonsa* species. The only exception
33 was the sampling 30 PD, when the abundance was higher, but there was no relationship

1 with the evaluated abiotic parameters. Seven macrozoobenthic groups were observed
2 (Polychaeta, Tanaidacea, Isopoda, Gastropoda, Bivalvia, Malacostraca and Ostracoda)
3 over two cycles period (2012 and 2013). Spatially, the density and species richness
4 suffered little variability over the sampling sites in the estuary for both campaigns.
5 Temporarily, it was observed an increase in these indices in winter compared to summer
6 in 2012. Over the 2013 samples, the macrozoobenthos density had an opposite pattern
7 and species richness was little variable. The TOC concentration ranged from 0.12 to
8 0.67% and the concentration of TN was below the equipment detection limit (<0.1 ppm)
9 in all sampling sites. The contents of Cu and Zn in the sediment were quantified by
10 weak extraction (labile fraction or potentially bioavailable) and semi-strong extraction
11 (more strongly adsorbed fraction). Cu in the labile fraction ranged from 0.12 to 1.27 μg
12 g^{-1} and Zn ranged from 0.52 to 3 $\mu\text{g g}^{-1}$, while in the more strongly adsorbed fraction,
13 Cu ranged from 0.3 to 2.65 $\mu\text{g g}^{-1}$ and Zn from 30.44 to 121.4 $\mu\text{g g}^{-1}$. Significant
14 differences (ANOVA, $p < 0.05$) were observed in the TOC values among sites in
15 samplings 1PD and 10PD, but not related to the effluents discharge. The Pearson
16 correlation analysis showed no relationship between the effluents discharge and increase
17 in TOC, TN, Cu or Zn in the sediment, except for the relation found between the values
18 of TOC, Cu and Zn in the sediment on 1 PD sampling. At the end of the cycle period,
19 shrimps were collected from two ponds and after the tissue analysis it revealed mean
20 concentrations of $6.63 \pm 0.2 \mu\text{g g}^{-1}$ of Cu and $19.76 \pm 0.2 \mu\text{g g}^{-1}$ of Zn in pond 1 and 7.6
21 $\pm 0.51 \mu\text{g g}^{-1}$ of Cu and $19.13 \pm 0.32 \mu\text{g g}^{-1}$ of Zn in pond 2. Although some water
22 quality parameters have suffered variations due to the effluents discharge, these effects
23 were acute and punctual, restricted to a distance of 20m from the EDC and observed
24 until 5 days after the discharge. In addition, all values were within the limits stipulated
25 by standard guidelines and available Brazilian legislation. Phytoplankton,
26 protozooplankton, meso-zooplankton and the macrozoobenthos showed natural
27 fluctuations related to abiotic parameters in estuarine systems. Finally, TOC, TN, Cu
28 and Zn in the sediment and Cu and Zn in the shrimp tissue were within the maximum
29 amounts stipulated by standard guidelines, showing that the effluents discharge from the
30 commercial shrimp farm in Patos Lagoon estuary did not cause negative impacts on the
31 environment.

32

33

1 INTRODUÇÃO GERAL

2 *3 Crescimento da aquicultura*

4 A aquicultura marinha e costeira representam um importante componente na
5 cadeia produtiva de alimentos (Webb et al., 2012). O crescimento populacional e a
6 estabilização das capturas selvagens têm permitido que essa atividade cresça a elevadas
7 taxas, sendo assim considerada de grande potencial para suprir a demanda mundial por
8 proteína de alta qualidade (Brander, 2007). América Latina e Caribe representam
9 atualmente a segunda posição no ranking mundial de crescimento, com taxas médias
10 que alcançaram 10%/ano entre 2000-2012 (FAO, 2014). Ainda segundo a FAO (2014),
11 o Brasil ocupa a décima segunda posição mundial na produção de pescados através da
12 aquicultura, com um montante total de mais de 700 mil Ton/ano. Dentro desse contexto,
13 apenas a produção de crustáceos representa mais de 70 mil Ton/ano, o que demonstra a
14 expressividade dessa atividade no Brasil.

15 Ao longo dos seus 8.500 km de linha de costa, o Brasil apresenta grande
16 potencial para o desenvolvimento da aquacultura marinha e costeira (Barroso et al.,
17 2007). No extremo sul do Brasil, a carcinicultura começou a se instalar no final da
18 década de 1990 com o cultivo de *Farfantepenaeus paulensis* e posteriormente se
19 consolidou com a criação do camarão branco do pacífico *Litopenaeus vannamei*. Com
20 sua importância cada vez mais expressiva frente ao mercado, a carcinocultura tem
21 ganhado foco por parte das autoridades ambientais no que diz respeito ao uso de água e
22 emissão de efluentes ao meio adjacente. Isso porque a aquicultura ainda tem uma
23 participação expressiva na poluição orgânica, eutrofização, lançamento de nutrientes e
24 efluentes ao ecossistema, floração de algas tóxicas, depleção de oxigênio e mudanças na
25 macrofauna bêntica (Naylor et al., 1998; Páez-Osuna, 2001; Anderson et al., 2002;
26 Aubin, 2006; Olsen et al., 2008).

27 Diversos incidentes até então mal compreendidos, causaram o colapso de
28 diversas fazendas de criação de camarões ao redor do mundo. Atualmente os motivos
29 são claros, entre eles a falta de manejo e a super-intensificação sem controle dos
30 sistemas de criação. A intensificação dos sistemas de criação favorece ainda mais o
31 potencial poluidor dessa atividade uma vez que aumenta-se a quantidade de fertilizantes
32 utilizados, o uso de dietas balanceadas ricas em nitrogênio (N) e fósforo (P) e,
33 consequentemente, os produtos decorrentes da excreção dos animais. Esses fatores são

1 apontados como os principais agentes potenciais para a eutrofização de zonas costeiras
2 (Cho et al., 1994; Jackson et al., 2004). A ração é citada como a principal responsável
3 pela deterioração da qualidade da água, uma vez que apenas 15-30% do alimento
4 ofertado é assimilado pelos animais (Barbieri & Ostrensky, 2002). De acordo com Boyd
5 & Tucker (1998), do montante oferecido pelas dietas, cerca de 25-45% do N, 20-30%
6 do P e 10-15% do C é absorvido e assimilado pelos camarões. O restante permanece
7 dentro dos viveiros na forma de fezes ou matéria orgânica em decomposição até ser
8 liberada para o meio ambiente através dos efluentes ao final do ciclo produtivo.

9 Islam (2005) reporta a importância das práticas de manejo na regulação da saída
10 de nutrientes para o meio ambiente. A troca de água dos viveiros ao longo do ciclo
11 produtivo é uma prática comum em sistemas de aquicultura. Essas são realizadas a fim
12 de controlar a concentração de fitoplâncton e de nutrientes nos viveiros, evitando que os
13 compostos nitrogenados e o oxigênio dissolvido alcancem níveis que possam ser
14 prejudiciais aos camarões (Lemonnier & Faninoz, 2006). A carga de nutrientes
15 originados durante um ciclo de criação é variável de acordo com o tempo de cultivo, o
16 sistema adotado e as técnicas de manejo empregadas. Primavera (1997) destaca ainda
17 que o efeito do efluente sobre o corpo d'água receptor depende de fatores como
18 magnitude de descarga, composição do efluente, taxa de diluição e tempo de residência.
19 A capacidade assimilativa de um determinado ambiente para o enriquecimento de
20 nutrientes é finita e a quantidade excessiva de nutrientes pode alterar a composição de
21 espécies, diversidade e dinâmica da comunidade biótica (Kennish, 1992). Além de
22 afetar diretamente a qualidade da água e os sedimentos, os resíduos orgânicos
23 particulados podem também ser utilizados diretamente por animais consumidores no
24 ecossistema receptor, podendo levar a mudanças nas comunidades planctônicas e
25 bentônicas (Roditi et al., 2000).

26 Um sistema que vêm sendo amplamente utilizado no tratamento de águas
27 residuais é a bacia de sedimentação. A utilização dessa tecnologia tem crescido desde os
28 anos 1970 nos mais diversos setores, entre eles a aquicultura. Diversos processos
29 bióticos e abióticos ocorrem nessas bacias, como mineralização microbiana,
30 nitrificação/desnitrificação, absorção de nutrientes pela vegetação, deposição do
31 material particulado, entre outros. Uma vez que os efluentes permaneçam um tempo
32 suficientemente longo na bacia de decantação, torna-se possível a precipitação de todos
33 os sólidos em suspensão, inclusive os mais finos (Boyd, 1990). Isso proporciona a
34 liberação de uma água para o ambiente com melhores parâmetros de qualidade. Lin et

1 al. (2005) reportam remoção de 55-66% dos sólidos suspensos e redução de 91-99% da
2 turbidez dos efluentes de um cultivo intensivo de *L. vannamei* através do uso dessa
3 tecnologia. As propriedades físico-químicas da água são usadas como indicadores
4 ambientais tanto na água dos viveiros como nos efluentes da carcinocultura. Esses
5 parâmetros permitem um melhor entendimento na avaliação dos impactos dos efluentes
6 sobre o meio ambiente receptor (Chua et al., 1989). Um dos grandes desafios da
7 carcinocultura é superar as preocupações ambientais para melhorar a eficiência
8 econômica, através do desenvolvimento e implementação de uma abordagem integrada
9 para a redução de resíduos nitrogenados (Jackson et al., 2003). Assim, o monitoramento
10 dos efluentes e dos parâmetros físico-químicos da água do ambiente receptor torna-se de
11 extrema importância para assegurar a viabilidade de um sistema de cultivo.

12

13 *Efeito do enriquecimento orgânico sobre o sedimento*

14 O sedimento tem um papel crucial na ciclagem de nutrientes. Em ecossistemas
15 costeiros rasos, como é o caso da Lagoa dos Patos, o sedimento pode atuar como a
16 principal fonte de nutrientes para a coluna d'água (Warnken et al., 2002). Os nutrientes
17 e matéria orgânica produzidos nos viveiros de cultivo de camarões estão presentes na
18 forma dissolvida e suspensa na água, e uma importante porção se acumula no sedimento
19 ou volatiliza para a atmosfera (Páez-Osuna et al., 1999). O material orgânico presente
20 no fundo dos viveiros da aquicultura é proveniente da produção primária do
21 fitoplâncton, dos restos de ração e dos produtos de excreção dos animais (Funge-Smith
22 & Briggs, 1998; Steeby et al., 2004). A matéria orgânica por sua vez tende a se
23 acumular no sedimento, mantendo-se mesmo após o final do ciclo de produção. Esse
24 acúmulo pode trazer impactos negativos ao ambiente biológico e químico dos viveiros
25 (Suplee & Cotner, 1996) ou do ambiente receptor dos efluentes, uma vez que essas
26 grandes quantidades de nutrientes tornam o sedimento um local favorável para o
27 desenvolvimento microbiano pela disponibilidade de matéria orgânica.

28 O incremento de matéria orgânica ao ecossistema pode causar eutrofização.
29 Sistemas eutrofizados geralmente revelam presença de carbono orgânico particulado
30 (Pelletier et al., 2011), o que torna esse composto um bom indicador de enriquecimento
31 em sedimentos (Hyland et al., 2005). O Nitrogênio orgânico por sua vez exerce um
32 importante papel como fonte de nutrientes (Fütterer, 2000). Esse nutriente provém
33 principalmente da proteína das rações, por meio da excreta na forma de amônia e por

1 meio das fezes na forma de nitrogênio orgânico (Henry-Silva & Camargo, 2007).
2 Briggs & Funge-Smith (1994) avaliando viveiros de criação de camarão na Tailândia
3 concluíram que 31% do N que entra no sistema é absorvido pelo sedimento.

4 A aquicultura é muitas vezes reportada como uma fonte potencial de metais
5 traço, os quais estão presentes como componentes naturais de rações, impurezas em
6 fertilizantes ou como princípio ativo de pesticidas (Tacon & Forster, 2003). Sedimentos
7 podem aprisionar metais introduzidos no sistema aquático servindo como bons
8 indicadores de poluição e permitindo uma avaliação consistente de contaminação
9 espacial e temporal (Salomons & Förstner, 1984; Buchman, 1989). O cobre (Cu) é um
10 elemento traço presente nas dietas essencial para os camarões sintetizarem hemocianina
11 na sua hemolinfa (Cuzon, 2004) e o zinco (Zn) é um cofator em diversos sistemas de
12 enzimas (Davis et al., 2002). O conteúdo residual de metais pode se acumular no
13 sedimento, servindo assim como indicadores dos efluentes da aquicultura (Chou et al.,
14 2002). A remobilização do sedimento favorece a liberação da fração lábil do metal para
15 a coluna d'água, o que pode causar efeitos tóxicos para os organismos (Wallner-
16 Kersanach et al., 2009). Metais traço que estão adsorvidos ao sedimento podem ser
17 liberados em decorrência de mudanças nas condições físico-químicas, principalmente
18 no pH e no potencial-redox (Cappuyns & Swennwn, 2005).

19 Além dos efeitos deletérios do acúmulo dos metais traço no sedimento, eles são
20 também conhecidos graças a sua potencialidade em se concentrar em órgãos de
21 organismos aquáticos. O excesso desses metais quando inseridos nas dietas além do
22 requerimento nutricional necessário, pode fazer com que o camarão *L. vannamei* possa
23 absorver e acumular no tecido ou dispersar para a água ou sedimento quando não
24 digerido (Wu & Yang, 2011). A análise de metais traço de organismos aquáticos pode
25 prover informações importantes do grau de contaminação ambiental e do potencial
26 impacto do consumo desse alimento (Ip et al., 2005).

27

28 *Efeitos sobre a biodiversidade*

29 *Comunidades planctônicas*

30 Diversos estudos têm acompanhado os fatores que afetam a variabilidade do
31 fitoplâncton nas áreas rasas do estuário da Lagoa dos Patos (Abreu et al., 1995;
32 Bergesch & Odebrecht, 1997; Fujita & Odebrecht, 2007; Abreu et al., 2010). Nesse
33 ambiente, a produtividade fitoplanctônica parece ser limitada pela luz, temperatura da

1 água e disponibilidade de nutrientes (Abreu et al., 1994). Entretanto, a abundância
2 fitoplanctônica também é dependente das condições hidrológicas locais, já que em
3 locais com alta hidrodinâmica a diluição dos nutrientes pode ser mais rápida do que sua
4 produção. Estudos prévios já têm demonstrado que a descarga de efluentes provenientes
5 de cultivo de camarões no estuário da Lagoa dos Patos podem causar mudanças de curto
6 prazo no ambiente receptor (Cardozo e Odebrecht, 2012). O *input* de nutrientes nos
7 viveiros de cultivo provenientes das dietas formuladas e das fertilizações pode
8 contribuir para o lançamento de efluentes ricos em nitrogênio e fósforo (Naylor et al.,
9 1998). Isso pode colaborar para o processo de enriquecimento orgânico do ecossistema,
10 processo conhecido como eutrofização. Esse fato deve ser observado com atenção já
11 que existe uma relação direta entre o aumento de nutrientes no meio ambiente e o
12 aumento da biomassa fitoplantônica em ambientes oligotróficos e pouco eutróficos,
13 como é o caso desse estuário (Hargrave, 1995). Assim, o monitoramento das
14 comunidades fitoplantônicas representam uma importante ferramenta de manejo de
15 forma a reduzir riscos ambientais (Abreu et al., 2010).

16 Da mesma forma, o zooplâncton pode ser utilizado como indicador da qualidade
17 da água em viveiros de criação de camarões e das condições ambientais do ambiente
18 receptor dos efluentes. Esses organismos respondem a baixos níveis de oxigênio
19 dissolvido, altos níveis de nutrientes, contaminantes tóxicos, predação, entre outros
20 (Casé et al., 2008). Assim como na maioria dos estuários temperados, a ocorrência e a
21 abundância de espécies de zooplâncton são determinadas principalmente por variações
22 sazonais de salinidade e de temperatura (Montú, 1980). O protozooplâncton por sua vez
23 pode exercer um papel chave na transferência de carbono da teia alimentar microbiana
24 para níveis tróficos mais elevados (Yang et al., 2012).

25

26 *Comunidade macrozoobentônica*

27 A entrada de nutrientes no meio proveniente dos efluentes da aquacultura pode
28 gerar mudanças físico-químicas significativas no sedimento, o que pode diminuir a
29 diversidade biológica e favorecendo o aparecimento de espécies oportunistas
30 (Johannssen et al., 1994). Esse enriquecimento orgânico pode levar a severas
31 modificações na estrutura das comunidades bênticas, levando a seleção de poucas
32 espécies tolerantes. Quando as taxas de acumulação de matéria orgânica ultrapassam a
33 capacidade de assimilação do sedimento, zonas anaeróbicas podem surgir, aumentando

assim a produção e liberação de compostos tóxicos reduzidos, como amônia, sulfeto de hidrogênio e metano (Alongi et al., 1999; Islam, 2005). Essas condições extremas prejudicam ainda mais a qualidade do sedimento e a estrutura da comunidade bentônica (Islam, 2005), podendo trazer como consequência o desaparecimento da fauna bêntica e a criação de uma zona anóxica (Heilskov & Holmer, 2001). Segundo Belan (1970) o aparecimento de organismos oportunistas pode ser usado como indicativo do enriquecimento do sedimento.

Organismos bentônicos desempenham um papel crucial no suprimento e mineralização da matéria orgânica (Heilskov & Holmer, 2001), o que faz com que o acompanhamento espaço-temporal das comunidades gere dados precisos na avaliação da qualidade ambiental de um ecossistema receptor de nutrientes. Diversos trabalhos têm sido desenvolvidos com vistas a avaliar e mitigar o impacto dos nutrientes liberados pela aquicultura sobre a comunidade bentônica (Bartoli et al., 2001; Soares et al., 2004; Carvalho et al., 2009; Aguardo-Giménez et al., 2011). Em regiões estuarinas, os organismos bentônicos são o principal link entre os produtores primários e os níveis tróficos superiores (Foreman et al., 1995), além de desempenharem um papel crucial no suprimento e mineralização da matéria orgânica (Heilskov & Holmer, 2001). No estuário da Lagoa dos Patos, a composição e abundância do macrozoobentos tem sido alvo de diversos estudos (Bemvenuti et al., 1978; Capitoli et al., 1978; Rosa e Bemvenuti, 2006; Colling et al., 2007), mas o acompanhamento dessa comunidade exposta ao lançamento dos efluentes da carcinocultura são escassos.

A hipótese testada é a de que o lançamento de efluentes proveniente do cultivo de camarões em sistema semi-intensivo possa causar alterações espaço-temporais sobre parâmetros de qualidade de água, o plâncton e nos níveis de carbono, nitrogênio e metais traço no sedimento do ambiente estuarino receptor.

Local de estudo

A Lagoa dos Patos é a maior laguna costeira estrangulada do mundo (Kjerfve, 1986), estende-se por 270 km de linha de costa e localiza-se entre as latitudes 30°S e 32°S (Möller et al. 1996). A hidrodinâmica é regulada principalmente pelas relações entre descarga fluvial e ação dos ventos, e, secundariamente, pelo efeito da maré (Möller et al. 2009; Seeliger, 2010). Sua formação está associada a múltiplas barreiras de areia complexas e pode ser dividida em três unidades biológicas: setor Norte (com a cidade de Porto Alegre e o Rio Guaíba como principais tributários), o setor Médio

1 (pouco populoso e com entrada de água doce do Rio Camaquã) e o setor Sul
2 (caracterizado como a região estuarina) (Seeliger et al., 1997). Possui importância
3 ecológica, industrial, portuária, agrícola e pesqueira, tornando o aporte de nutrientes de
4 origem antrópica a principal causa de eutrofização desse sistema aquático (Reis &
5 D'Incao, 2000; Seeliger, 2010; Niencheski et al., 2014). Este estudo foi realizado nas
6 proximidades de uma fazenda comercial de criação de camarões da espécie *Litopenaeus*
7 *vannamei* localizada as margens do estuário da Lagoa dos Patos, sul do Brasil (Rio
8 Grande do Sul, Brasil - 31°56'04S, 52°00'11W) (Figura 1).

9

10



11 Figura 1: Localização da fazenda de cultivo e dos respectivos pontos de coleta. V1 = Viveiro,
12 V2 = Viveiro 2 e BS = Bacia de Sedimentação.
13

14

15

16

1 **OBJETIVO GERAL**

2 Avaliar espaço-temporalmemente os possíveis impactos dos efluentes de uma
3 fazenda de criação de camarões marinhos sobre a qualidade da água, o plâncton, o
4 macrozoobentos e os sedimentos do ambiente estuarino receptor.

5

6 *Objetivos específicos*

- 7 • Determinar a variabilidade do plâncton e da qualidade da água antes e após a
8 liberação dos efluentes;
- 9 • Identificar a influência do cultivo de camarões sobre a comunidade
10 macrozoobentônica estuarina ao longo de dois ciclos produtivos;
- 11 • Determinar a variabilidade da concentração de carbono orgânico total, nitrogênio
12 total, cobre e zinco no sedimento do viveiro, bacia de sedimentação e do estuário
13 antes e após a liberação dos efluentes; determinar ainda os teores de cobre e zinco
14 nos camarões ao final do ciclo produtivo.

15

1 **REFERÊNCIAS BIBLIOGRÁFICAS**

- 3 ABREU, PC, E GRANÉLI, C ODEBRECHT, D KITZMANN, LA PROENCA, C
4 RESGALLA & E GRANELI. 1994. Effect of fish and mesozooplankton
5 manipulation on the phytoplankton community in the Patos Lagoon estuary,
6 Southern Brazil. *Estuaries* 17: 575–584.
- 7 ABREU, PC, C HARTMANN & C ODEBRECHT. 1995. Nutrient-rich Saltwater and,
8 its Influence on the Phytoplankton of the Patos Lagoon Estuary, Southern
9 Brazil. *Estuar. Coast. Shelf Science*, 40: 219–229.
- 10 ABREU PC, M BERGESCH, LA PROENÇA & C ODEBRECHT. 2010. Short- and
11 long- term chlorophyll a variability in the shallow microtidal Patos Lagoon
12 estuary, Southern Brazil. *Estuar. Coast.,* 33: 554-569.
- 13 AGUADO-GIMÉNEZ, F, MA PIEDECAUSA, C CARRASCO, JM GUTIÉRREZ, V
14 ALIAGA & B GARCÍA-GARCÍA. 2011. Do benthic biofilters contribute to
15 sustainability and restoration of the benthic environment impacted by offshore
16 cage finfish aquaculture? *Mar. Pollut. Bull.*, 62, 1714–1724.
- 17 ALONGI, DM, FT LINDSAY & A TROTT. 1999. Rates and pathways of benthic
18 mineralization in extensive shrimp ponds of the Mekong delta, Vietnam.
19 *Aquaculture*175: 269-292.
- 20 ANDERSON, DM, PM GLIBERT, JM BURKHOLDER. 2002. Harmful algal blooms
21 and eutrophication nutrient sources, composition, and consequences. *Estuaries*
22 25: 704–726.
- 23 AUBIN, J. 2006. Characterization of the environmental impact of a turbot
24 (*Scophthalmus maximus*) recirculating production system using life cycle
25 assessment. *Aquaculture* 261: 1259–1268.
- 26 BARBIERI JÚNIOR, RC & ANT OSTRENSKY. 2002. Camarões Marinho – Engorda.
27 Aprenda Fácil Editora. Viçosa– MG. 370p.
- 28 BARROSO, GF, LHS POERSCH, RO CAVALLI (Orgs.). 2007. Sistemas de cultivos
29 aquícolas na zona costeira do Brasil: recursos, tecnologias, aspectos ambientais
30 e sócio-econômicos. Museu Nacional, Rio de Janeiro. 326p.
- 31 BARTOLI, M, D NIZZOLI, P VIAROLI, E TUROLLA, G CASTALDELLI, EA
32 FANO & R ROSSI. 2001. Impact of *Tapes philippinarum* farming on nutrient

- 1 dynamics and benthic respiration in the Sacca di Goro. *Hydrobiologia*, 455:
2 203–212.
- 3 BELLAN G, 1970. Pollution by sewage in Marseilles. *Mar. Pollut. Bull.*, 1: 59–60.
- 4 BEMVENUTI, CE, RR CAPÍTOLI & NM GIANUCA. 1978. Estudos de ecologia
5 bentônica na região estuarial da Lagoa dos Patos. II. Distribuição quantitativa
6 do macrobentos infralitoral. *Atlântica* 3: 23-32.
- 7 BERGESCH M & C ODEBRECHT. 1997. Análise do fitoplâncton, protozooplâncton e
8 de alguns fatores abióticos no estuário da Lagoa dos Patos. *Atlântica* 19: 31–
9 50.
- 10 BOYD, CE. 1990. Water Quality in Ponds for Aquaculture. Alabama Agricultural
11 Experiment Station, Auburn University, AL.
- 12 BOYD, CE & CS TUCKER. 1998. Pond Aquaculture Water Quality Management.
13 Kluwer Academic Publishers, Dordrecht.
- 14 BRANDER, KM. 2007. Global fish production and climate change. *PNAS* 104, 19709–
15 19714.
- 16 BRIGGS, MRP & SJ FUNGE-SMITH. 1994. A nutrient budget of some intensive
17 marine shrimp ponds in Thailand. *Aquac. Fish. Manag.*, 25:789–811.
- 18 BUCHMAN MF. 1989. A review and summary of trace contaminant data for coastal
19 and estuary Oregon: U.S. Department of Commerce, National Oceanic and
20 Atmospheric Administration, National Ocean Service, NOAA Technical
21 Memorandum NOS OMA 42, 115p.
- 22 CAPÍTOLI, RR, CE BEMVENUTI & NM GIANUCA. 1978. Estudos de ecologia
23 bentônica na região estuarial da Lagoa dos Patos, I. Comunidades bentônicas.
24 *Atlântica* 3: 5-21.
- 25 CAPPUYNS, V & R SWENNWN. 2005 Kinetics of Element Release during Combined
26 Oxidation and pHstat leaching of Anoxic River Sediments. *Appl. Geochem.*,
27 20: 1169-1179.
- 28 CARDOZO, AP & C ODEBRECHT. 2012. Effects of shrimp pond water on
29 phytoplankton: importance of salinity and trophic status of the receiving
30 environment. *Aquac. Res.*, 1-11.
- 31 CARVALHO, S, M FALCÃO, J CÚRDIA, A MOURA, D SERPA, MB GASPAR, MT
32 DINIS, P POUSÃO FERREIRA & LC FONSECA. 2009. Benthic dynamics

- 1 within a land-based semi-intensive aquaculture fish farm: the importance of
2 settlement ponds. *Aquacult. Int.*, 571–587.
- 3 CASÉ, M, EE LEÇA, SN LEITÃO, EE SANT'ANNA, R SCHWAMBORN & ATM
4 JUNIOR. 2008. Plankton community as an indicator of water quality in
5 tropical shrimp culture ponds. *Mar. Pollut. Bull.*, 56: 1343–1352.
- 6 CHO C, J HYNES, K WOOD & YH YOSHIDA. 1994. Development of high nutrient
7 dense, low-pollution diets and prediction of aquaculture wastes using
8 biological approaches. *Aquaculture* 124: 293-305.
- 9 CHOU, CL, K HAYA, LA PAON, L BURRIDGE & JD MOFFATT. 2002.
10 Aquaculture-related trace metals in sediments and lobsters and relevance to
11 environmental monitoring program ratings for nearfield effects. *Mar. Pollut.*
12 *Bull.*, 44: 1259–1268.
- 13 CHUA, TE, JN PAW & FY GUARIN. 1989. The environmental impact of aquaculture
14 and the effects of pollution on coastal aquaculture development in southeast
15 Asia. *Mar. Pollut. Bull.*, 20: 335–343.
- 16 COLLING, LA, CE BEMVENUTI & MS GANDRA. 2007. Seasonal variability on the
17 structure of sublittoral macrozoobenthic association in the Patos Lagoon
18 estuary, southern Brazil. *Iher. Ser. Zool.*, 97: 257-263.
- 19 CUZON, G. 2004. Nutrition of *Litopenaeus vannamei* reared in tanks or in ponds.
20 *Aquaculture* 235: 513–551.
- 21 DAVIS, DA, CR ARNOLD & I MCCALLUM. 2002. Nutritional value of feed peas
22 (*Pisum sativum*) in practical diet formulations for *Litopenaeus vannamei*.
23 *Aquac. Nutr.*, 8: 87–94.
- 24 FAO, 2014. The State of World Fisheries and Aquaculture 2014. Food and Agriculture
25 Organization of the United Nations, Rome. 243p.
- 26 FOREMAN, K, I VALIELA & R SARDA. 1995. Control of benthic marine food webs.
27 *Sci. Mar.*, 59: 119–128.
- 28 FUJITA CCO & C ODEBRECHT. 2007. Short-term variability of chlorophyll a and
29 phytoplankton composition in a shallow area of the Patos Lagoon estuary
30 (Southern Brazil). *Atlântica* 29: 93-107.
- 31 FUNGE-SMITH, SJ & MRP BRIGGS. 1998. Nutrient budgets in intensive shrimp
32 ponds: implications for sustainability. *Aquaculture* 164: 117–133.
- 33 FÜTTERER, DK. 2000. The solid phase of marine sediments, in Schulz, H.D., Zabel,
34 M., (eds.), *Marine Geochemistry*: Berlin, Springer. 1-26 p.

- 1 HARGRAVE, BT. 1995. Impacts of man's activities on aquatic system. In:
2 Fundamental of Aquatic Ecosystems, Ed. Blackwell Science. 245-264 p.
- 3 HEILSKOV AC & M HOLMER. 2001. Effects of benthic fauna on organic matter
4 mineralization in fish-farm sediments: importance of size and abundance. ICES
5 J Mar Sci 58: 427–434.
- 6 HENRY-SILVA, GG & AFM CAMARGO. 2007. Impacto das atividades de
7 aquicultura e sistemas de tratamento de efluentes com macrófitas aquáticas –
8 relato de caso*. B. Inst. Pesca 34: 163 – 173.
- 9 HYLAND J, L BALTHIS, I KARAKASSIS, P MAGNI, J SHINE, O VESTERGAARD
10 & R WARWICK. 2005. Organic carbon of sediments as an indicator of stress
11 in the marine benthos. Mar. Ecol. Prog. Ser., 295:91–103.
- 12 ISLAM, MS. 2005. Nitrogen and phosphorus budget in coastal and marine cage
13 aquaculture and impacts of effluent loading on ecosystem: review and analysis
14 towards model development. Mar Pollut Bull., 50: 48–61.
- 15 IP, CCM, XD LI, G ZHANG, CSC WONG & WL ZHANG. 2005. Heavy metal and Pb
16 isotopic compositions of aquatic organisms in the Pearl River Estuary, South
17 China. Environ. Pollut., 138: 494–504.
- 18 JACKSON, C, N PRESTON, PJ THOMPSON & M BURFORD. 2003. Nitrogen
19 budget and effluent nitrogen components at an intensive shrimp farm.
20 Aquaculture, 218: 397–411.
- 21 JACKSON, C, N PRESTON & PJ THOMPSON. 2004. Intake and discharge nutrient
22 loads at three intensive shrimp farms. Aquac. Res., 35: 1053–1061.
- 23 JOHANNESSEN, P, H BOTNEN & O TVEDTEN. 1994. Macrobenthos: before, during
24 and after a fish farm. Aquac. Fish. Manag., 25: 55-66.
- 25 KENNISH, MJ. 1992. Ecology of Estuaries: Anthropogenic Effects. CRC Press, Boca
26 Raton, FL.
- 27 KJERFVE, B. 1986 Comparative oceanography of coastal lagoons. In Estuarine
28 Variability (Wolfe, D. A., ed.). Academic Press, New York, pp. 63-81.
- 29 LEMONNIER, H & S FANINOZ. 2006. Effects of water exchange rate on effluent and
30 sediment characteristics and on partial nitrogen budget in semi-intensive
31 shrimp ponds in New Caledonia. Aquac. Res., 37: 938–948.
- 32 LIN, Y, S JING, D LEE, Y CHANG, Y CHEN & K SHIH. 2005. Performance of a
33 constructed wetland treating intensive shrimp aquaculture wastewater under
34 high hydraulic loading rate. Environ Pollut., 134: 411–421.

- 1 MÖLLER, OO, JA LORRENZZENTI, JL STECH & MM MATA. 1996. The Patos
2 Lagoon summertime circulation and dynamics. Cont. Shelf Res., 16: 335-351.
- 3 MÖLLER, OO, JP CASTELLO & AC VAZ. 2009. The effect of river discharge and
4 winds on the interannual variability of the pink shrimp *Farfantepenaeus*
5 *paulensis* Production in Patos Lagoon. Est. Coast 32: 787–796.
- 6 MONTÚ, M. 1980. Zooplâncton do estuário da Lagoa dos Patos. I. Estrutura e
7 variações temporais e espaciais da comunidade. Atlântica 4: 53-72.
- 8 NAYLOR, RL, RJ GOLDBURG, H MOONEY, M BEVERIDGE, J CLAY, C FOLKE,
9 N KAUTSKY, J LUBCHENCO, J PRIMAVERA, M WILLIAMS. 1998.
10 Nature's subsidies to shrimp and salmon farming. Science, 282: 883–884.
- 11 NIENCHESKI, LF, WS MOORE & HL WINDOM. 2014. History of human activity in
12 coastal southern Brazil from sediment. Mar. Pollut. Bull., 78: 209–212.
- 13 OLSEN, LM, M HOLMER, Y OLSEN. 2008. Perspectives of nutrient emission from
14 fish aquaculture in coastal waters Literature review with evaluated state of
15 knowledge FHF project no. 542014. The Fishery and Aquaculture Industry
16 Research Fund.
- 17 PÁEZ-OSUNA, F, SR GUERRERO-GALVÁN & AC RUIZ-FERNANDÉZ. 1999.
18 Discharge of Nutrients from Shrimp Farming to Coastal Waters of the Gulf of
19 California. Mar. Pollut. Bull., 38: 585–592.
- 20 PÁEZ-OSUNA, F. 2001. The environmental impact of shrimp aquaculture: causes,
21 effects and mitigating alternatives. Environ. Manage., 28: 131–140.
- 22 PELLETIER, MC, DE CAMPBELL, KT HO, RM BURGESS, CT AUDETTE & NE
23 DETENBECK. 2011. Can sediment total organic carbon and grain size be used
24 to diagnose organic enrichment in estuaries? Environ. Toxicol. Chem., 30:
25 538–547.
- 26 PRIMAVERA, JH. 1997. Fish predation on mangrove-associated penaeids. The role of
27 structures and substrate. J. Exp. Mar. Biol. Ecol., 215: 205–216.
- 28 REIS EG & F D'INCAO. 2000. The present status of artisanal fisheries of extreme
29 Southern Brazil: an effort towards community-based management. Ocean
30 Coast Manage., 43: 585-595.
- 31 RODITI, HA, NS FISHER & SA SANUDO-WILHELMY. 2000. Uptake of dissolved
32 organic carbon and trace elements by zebra mussels. Nature 407: 78–80.

- 1 ROSA, LC & CE BEMVENUTI. 2006. Temporal variability of the estuarine
2 macrofauna of the Patos Lagoon, Brazil. *Revista de Biología Marina y*
3 *Oceanografía* 41: 1–9.
- 4 SALOMONS, W & U FÖRSTNER. 1984. Metals in the Hidrocycle. Berlin Springer
5 Verlag. 340p.
- 6 SEELIGER, U, C ODEBRECHT & JP CASTELLO. 1997. Subtropical convergence
7 environments. The coast and sea in the Southwest- ern Atlantic. Berlin:
8 Springer, 308 pp.
- 9 SEELIGER, U. 2010. Introdução. In: SEELIGER, U & C ODEBRECHT (eds.) O
10 estuário da Lagoa dos Patos: Um século de transformações. FURG, Rio
11 Grande, 11-13.
- 12 SOARES, R, S PEIXOTO, C BEMVENUTI, W WASIELESKY, F D'INCAO, N
13 MURCIA & S SUITA S. 2004. Composition and abundance of invertebrate
14 benthic fauna in Farfantepenaeus paulensis culture pens (Patos Lagoon estuary,
15 Southern Brazil). *Aquaculture* 239: 199–215.
- 16 STEEBY, JAMES A, JOHN A HARGREAVES, CRAIG S TUCKER, AND SUE
17 KINGSBURY. 2004. Accumulation, organic carbon and dry matter
18 concentration of sediment in commercial channel catfish ponds. *Aquacult.*
19 Eng., 30: 115–126.
- 20 SUPLEE, MW & JB COTNER. 1996. Temporal changes in oxygen demand and
21 bacterial sulfate reduction in inland shrimp ponds. *Aquaculture* 145: 141–158.
- 22 TACON, AGJ & IP FORSTER. 2003. Aquafeeds and the environment: policy
23 implications. *Aquaculture* 226: 181–189.
- 24 WALLNER-KERSANACH, M, CEF ANDRADE, MR MILANI & LFH
25 NIENCHESKI. 2009. In situ measurement of trace metals in estuarine waters
26 of the Patos Lagoon using the diffusive gradient in thin film (DGT). *Journal*
27 *Braz. Chem. Soc.* 20: 333-340.
- 28 WARPNKEN, KW, GA GILL, RD LEHMAN, TM DELLA PENNA & MA ALLISON.
29 2002. The effects of shrimp trawling on sediment oxygen demand and the
30 release of trace metals and nutrients from estuarine sediments. *Est. Coast. and*
31 *Shelf Science*, 57: 25-42.
- 32 WEBB, JM, R QUINTÃ, S PAPADIMITRIOU, L NORMAN, M RIGBY, DN
33 THOMAS, AND L LE VAY. 2012. Halophyte filter beds for treatment of
34 saline wastewater from aquaculture. *Water Res.*, 46: 5102-5114.

- 1 WU, XY & YF YANG. 2011. Heavy metal (Pb, Co, Cd, Cr, Cu, Fe, Mn and Zn)
2 concentrations in harvest-size white shrimp *Litopenaeus vannamei* tissues from
3 aquaculture and wild source. J. Food Compos. Anal., 24: 62–65.
- 4 YANG, EJ, JH HYUN, D KIM, J PARK, SH KANG, HC SHIN & S LEE. 2012.
5 Mesoscale distribution of protozooplankton communities and their herbivory in
6 the western Scotia Sea of the Southern Ocean during the austral spring. J. Exp.
7 Mar. Biol. Ecol., 428: 5–15.
- 8

CAPÍTULO 1

VARIABILIDADE DO PLÂNTON E DA QUALIDADE DA ÁGUA EM UM ESTUÁRIO ANTES E APÓS O LANÇAMENTO DE EFLuentes DA CARCINOCULTURA: IMPACTOS E REGENERAÇÃO

Artigo submetido ao periódico *Brazilian Journal of Oceanography* (ISSN: 1982-436X).

1 **PLANKTON AND WATER QUALITY VARIABILITY IN AN ESTUARY**
2 **BEFORE AND AFTER THE SHRIMP FARMING EFFLUENTS: POSSIBLE**
3 **IMPACTS AND REGENERATION**

4

5 **PLANKTON AND WATER QUALITY VARIABILITY SUBJECT TO SHRIMP**
6 **FARM EFFLUENT**

7

8 William Bauer

9 Paulo Cesar Abreu

10 Luís H. Poersch*

11 Universidade Federal de Rio Grande – FURG

12 Instituto de Oceanografia

13 (Av. Itália Km 08, Campus Carreiros, CP 474, 96201-900 Rio Grande, RS, Brasil)

14 *Corresponding author: lpoersch@mikrus.com.br

15

1 ABSTRACT

2 Water quality, chlorophyll a, phytoplankton, proto and mezo-zooplankton abundance
3 were spatiotemporally evaluated in an estuary receiving effluents from a Pacific white
4 shrimp *Litopenaeus vannamei* farm in Patos Lagoon estuary, Southern Brazil. Samples
5 were taken before (BD) and; 1 day (1 PD) 5 days (5 PD), 10 days (10 PD), 20 days (20
6 PD) and 30 days (30 PD) after the effluents discharge. Some water quality parameters
7 were affected by the effluents discharge; however, these changes were restricted to a
8 distance of 20 m from the effluent discharge channel for a period of 5 days. The
9 microbial community was dominated by chlorophyceae, followed by diatoms,
10 cyanobacteria and ciliates. There was an increase in the abundance of different groups
11 on the 1 PD sampling compared to BD. The zooplankton abundance was low in
12 practically all sites, except for 30 PD sampling. The meso-zooplanktonic organisms
13 were represented by copepods, mostly *Acartia tonsa*. Despite some effects on water
14 quality and phytoplankton and protozooplankton abundance until 5 PD sampling, these
15 alterations dissipated in a short period of time. We conclude that the environment
16 quickly assimilated the effluents discharge, and the water quality parameters remained
17 within the limits stipulated by standard guidelines.

18

19 RESUMO

20 Parâmetros de qualidade da água, composição e abundância do fitoplâncton e do proto e
21 meso-zooplâncton foram avaliadas espaço-temporalmemente no estuário receptor de
22 efluentes de uma fazenda produtora de camarão *Litopenaeus vannamei* na Lagoa dos
23 Patos, Brasil. As amostras foram tomadas em sete pontos antes (BD) e; 1 dia (1 PD), 5
24 dias (5 PD), 10 dias (10 PD), 20 dias (20 PD) e 30 dias (30 PD) após a descarga dos
25 efluentes. Alguns dos parâmetros de qualidade de água sofreram alterações devido ao
26 lançamento dos efluentes, restritos a 20 m do canal de descarga dos efluentes e por um
27 período máximo de 5 dias. A comunidade microbiana sofreu variação em 1 PD
28 comparativamente a BD. A abundância zooplânctônica foi baixa na maioria das
29 amostras, com exceção de alguns pontos em 30 PD. Os copépodos foram o único grupo
30 encontrado no meso-zooplâncton, na sua grande maioria a espécie *Acartia tonsa*.
31 Apesar de modificações em alguns parâmetros avaliados nos primeiros dias após o
32 lançamento dos efluentes, essas alterações foram assimiladas em um curto período. Os
33 parâmetros de qualidade de água mantiveram-se dentro dos limites estipulados pela
34 legislação e o ambiente assimilou de maneira rápida as mudanças ocorridas.

35 Descriptors: Aquaculture, Environmental Impact, Plankton, Effluent.

36 Descritores: Aquicultura, Impacto Ambiental, Plâncton, Efluente.

37

38

1 INTRODUCTION

2 The global seafood consumption has been increasing mainly due the world's
3 growing population, fisheries stagnation and consumption habits improvement (FAO,
4 2012). Shrimp is still the largest single commodity in value terms, representing about 15
5 percent of the total value of internationally traded fishery products (FAO, 2014). The
6 Brazilian shrimp farming started in 1970 in the Northeast region, raising different
7 *Penaeus* species (MOLES; BUNGES, 2002). In the Southeast, facilities started to work
8 in the late 1990 with the native shrimp *Farfantepenaeus paulensis*, and later with the
9 Pacific white shrimp *Litopenaeus vannamei* around the Patos Lagoon estuary. The Patos
10 Lagoon is the world's largest chocked lagoon (KJERFVE, 1986) with 270 km of
11 coastline between latitudes 30°S and 32°S (MÖLLER et al., 1996). The importance of
12 this lagoon remains on its ecology, industries, ports, agriculture and fisheries, and the
13 anthropogenic impact by the nutrient input is the major cause of eutrophication of this
14 aquatic system. (REIS; D'INCAO, 2000; SEELIGER, 2010; NIENCHESKI et al.,
15 2014).

16 The shrimp farming, as an anthropic activity, can contribute to the nutrient input
17 in adjacent ecosystems by the release of effluents loaded with nitrogen and phosphorous
18 (NAYLOR et al., 1998). These nutrients come from fertilizes use, shrimps' excretion
19 and unconsumed aquafeeds (CHO et al., 1994; BURFORD, 1997; JACKSON et al.,
20 2004; HERBECK et al., 2013). The organic matter and nutrient accumulation in shrimp
21 ponds lead to large phytoplankton blooms (ALONSO; OSUNA, 2003), increasing the
22 effluents pollution potential and generating anoxia conditions in the receiving water
23 body (BURFORD; WILLIAMS, 2001; JACKSON et al., 2004).

24 The phytoplankton is considered a sensitive biological indicator that responds to
25 the anthropogenic stress (COUTINHO et al., 2012). According to HARGRAVE (1991),
26 there is a direct relationship between increased nutrients in coastal waters and the
27 enlargement of phytoplankton biomass in oligotrophic environments as Patos Lagoon.
28 Several studies have been conducted in the Patos Lagoon estuary shallow areas to
29 evaluate its environmental variability (ABREU et al., 1995; BERGESCH;
30 ODEBRECHT, 1997; FUJITA; ODEBRECHT, 2007; ABREU et al., 2010). ABREU et
31 al. (2010) reported that the most significant variations in chlorophyll *a* in this estuary
32 occur within days to weeks. As stated by OLSEN et al. (2008), the assimilation capacity
33 of the water column is mediated by two main mechanisms: nutrient assimilation and

1 hydrodynamics. The nutrient uptake and assimilation by the phytoplankton transfers
2 energy and materials to higher trophic levels and the hydrodynamics involve the
3 transport and dilution of nutrients and planktonic organisms.

4 Therefore, the nutrients input can increase not only the phytoplankton, but also
5 all food web. Protozooplankton play an essential role in the carbon transfer of the
6 microbial food web to higher trophic levels (YANG et al., 2012). The ciliates are
7 sensitive to environmental alterations, and their community fluctuations can affect the
8 food web and the energy transfer (CHEN et al., 2009). The zooplankton is a group of
9 organisms that can also be used as biological indicators, once they respond to low
10 dissolved oxygen, high nutrient levels and toxic contaminants (CASÉ et al., 2008). In
11 the Patos Lagoon the protozooplankton is comprised of flagellates, dinoflagellates and
12 ciliates (ABREU; ODEBRECHT, 1998). The largest biomass of this group is found
13 during spring, while the smallest occurs during winter (ABREU et al., 1992). The
14 zooplankton species in this lagoon is strictly related to local hydrological conditions
15 (MONTÚ et al., 1998), and the most abundant organisms are copepods of *Acartia*
16 genus, only the *Acartia tonsa* occurring in high densities (> 40.016 organisms/m³)
17 (MONTÚ et al., 1997).

18 Previous studies showed that the shrimp farm effluents discharge can cause short
19 term changes in Patos Lagoon (CARDOZO; ODEBRECHT, 2012). However, no
20 detailed analyzes on spatial-temporal variation of water quality and plankton organisms
21 were available. Thus, this study aim to evaluate the *L. vannamei* farm effluents effects
22 on the water quality, phytoplankton, proto and meso-zooplankton in an estuary in
23 Southern Brazil.

24

25 MATERIAL AND METHODS

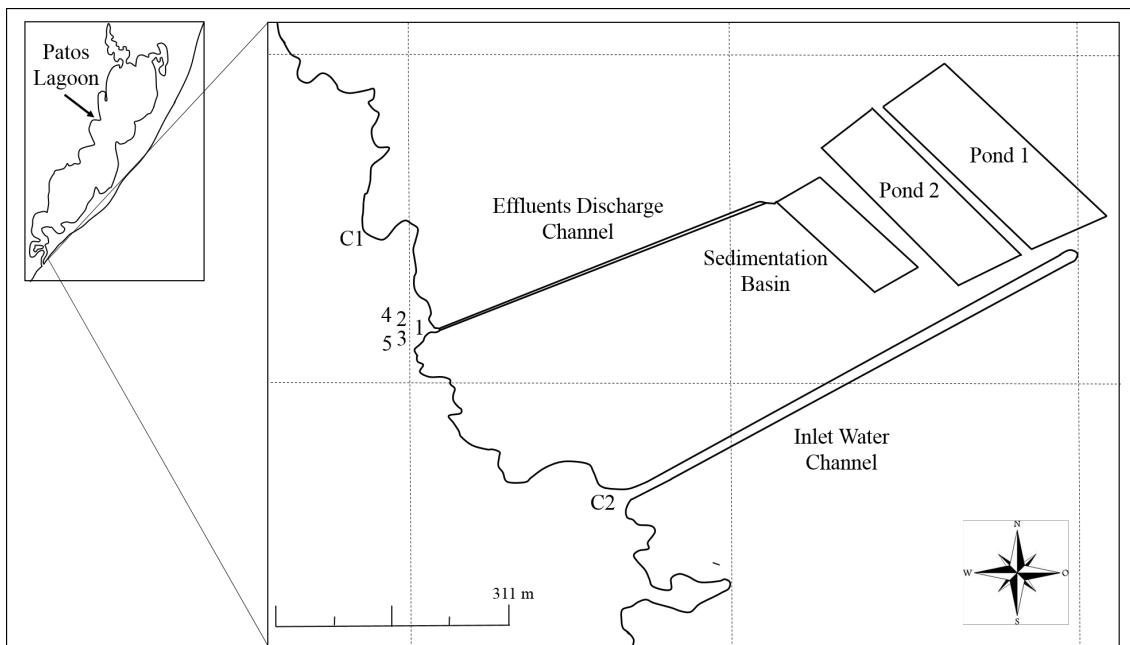
26 *Study site and sampling procedures*

27 This study was conducted in the Patos Lagoon estuary adjacent to a commercial
28 *L. vannamei* shrimp farm (Rio Grande do Sul, Brazil - 31°56'04S, 52°00'11W). The
29 earthen ponds were filled with water from the estuary, and before stocking it was
30 proceeded the fertilization with 270 kg of urea ((NH₂)₂ CO) and 28 Kg of calcium triple
31 superphosphate (Ca (H₂PO₄)₂) to stimulate phytoplanktonic growth. The facility works
32 in a semi-intensive system, and the stocking density was 12 shrimp/m² with a total area
33 of 3.75 ha. Shrimps were fed with commercial pellets (35% of crude protein). The cycle

1 lasted 112 days; there was no effluent discharge during the culture period; water was
2 only added in order to compensate the evaporation and soil infiltration. Both ponds
3 share a 1 ha vegetated sedimentation basin, which was empty until the end of the cycle.

4 The sampling sites were assigned as follows: across the EDC (1); in 20 m from
5 the channel (2 and 3); 30 m from the channel (4 and 5) and two control sites located at
6 100 m and 250 m from channel (C1 and C2, respectively) (Figure 1). In addition to
7 these sites, the ponds and the sedimentation basin were also sampled. This study was
8 conducted from February to April 2012, and samplings were taken before the discharge
9 (BD), 1-day post-discharge (1 PD), 5 days post-discharge (5 PD), 10 days post-
10 discharge (10 PD), 20 days post-discharge (20 PD) and 30 days post-discharge (30 PD).

11



12

13 Figure 1: Sampling sites in the shrimp farming. C1 = Control 1, C2 = Control 2, 1 = Effluents discharge
14 channel, 2 and 3 = 20m from the channel, 4 and 5 = 30m from the channel, P1 = Pond 1 and P2 = Pond 2.
15

16 Water quality

17 Temperature (WTW Oxi 3205), dissolved oxygen (WTW Oxi 3205), pH (YSI
18 60) and salinity (refractometer) were measured *in situ*, with three replicates.
19 Meteorological data were obtained from the Meteorological Station - Federal University
20 of Rio Grande. At each site, three replicates were taken from the water surface for
21 chlorophyll *a*, total ammonia nitrogen (UNESCO, 1983), nitrite (BENDSCHNEIDER;
22 ROBINSON, 1952), nitrate and total phosphorous (AMINOT; CHAUSSEPIED, 1983).

1 At these same sites water was collect from the surface and placed in amber glass
2 flasks containing formaldehyde 4% solution for further evaluation of phytoplankton and
3 protozooplankton identification and density estimative. The aliquots were analyzed in
4 sedimentation chamber (2.1 ml) under inverted microscope following the Utermöhl
5 method (HASLE, 1978). Ciliates and diatoms were counted in all chamber's area under
6 20X magnification. Chlorophyceae and cyanobacteria were counted in random fields
7 under 10X magnification until <30% coefficient of variation was reached.

8 Samples for zooplankton analysis were collected using cylindrical-conical net
9 with 30cm in diameter (150 µm mesh) fitted with a mechanical flow meter attached to
10 the net mouth. The net was hauled at surface covering an average distance of 15 meters.
11 The collected material was transferred to 100 mL glass flasks containing formaldehyde
12 4% solution. Sub-samples were then transferred to Bogorov chambers and analyzed
13 under a stereoscopic microscope (BOLTOVSKOY, 1981). The zooplankton specimens
14 were identified to major groups.

15 For the chlorophyll *a*, 25 mL aliquotes were filtered (Whatman GF/F) and
16 extracted with acetone 90% in the dark and the concentration was estimated measuring
17 the fluorescence in a calibrated fluorometer (Turner TD-700). (WELSCHMEYER,
18 1994).

19 *Statistics*

20 Zooplankton data were compared with the non-parametric Kruskal-Wallis test,
21 with 5% significance (SOKAL; ROHLF, 1995).

22

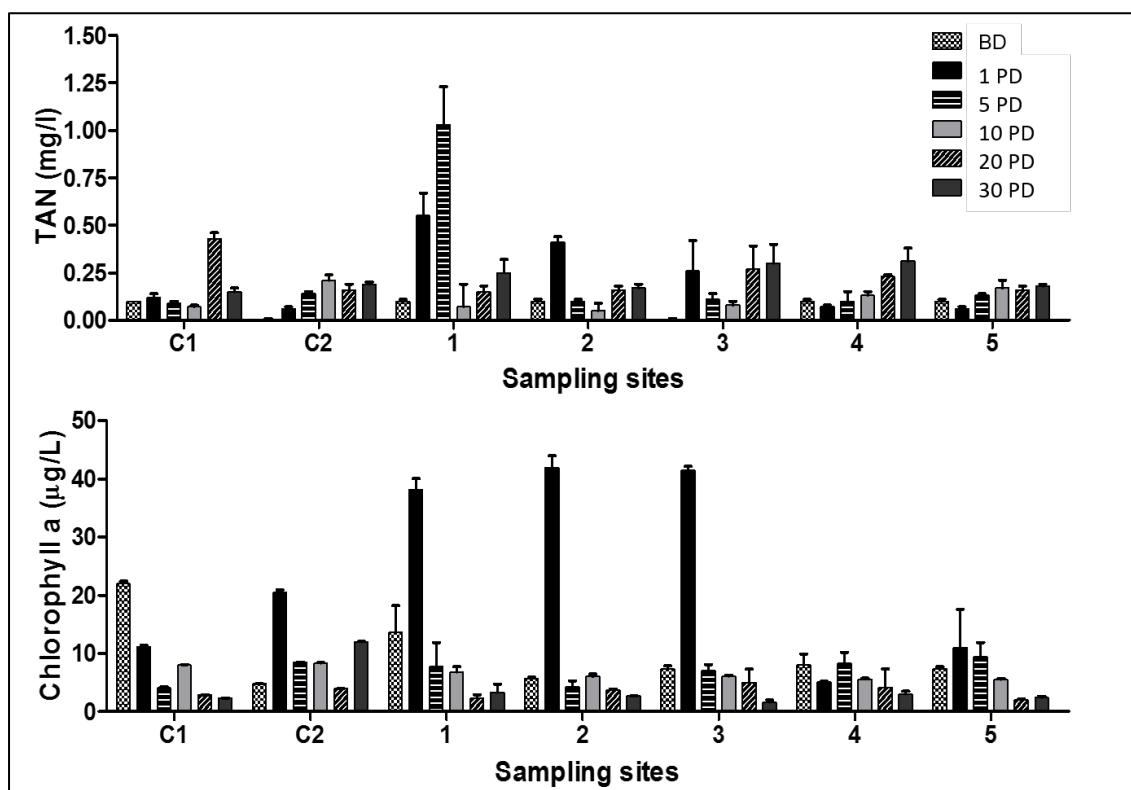
23 RESULTS

24 *Water quality in the ponds and in the receiving estuary*

25 Water quality parameters, as well as the chlorophyll *a* values are shown in
26 Figure 2. The total ammonia nitrogen (TAN) had small variation over the sampling
27 period and sites, showing only two peaks observed in sample 1 PD (0.55 mg / L) and 5
28 PD (1.03 mg / L) in site 1. The nitrite, nitrate and phosphate had null values in all
29 samples, both in the ponds and in the estuary. The only exception was the low nitrite

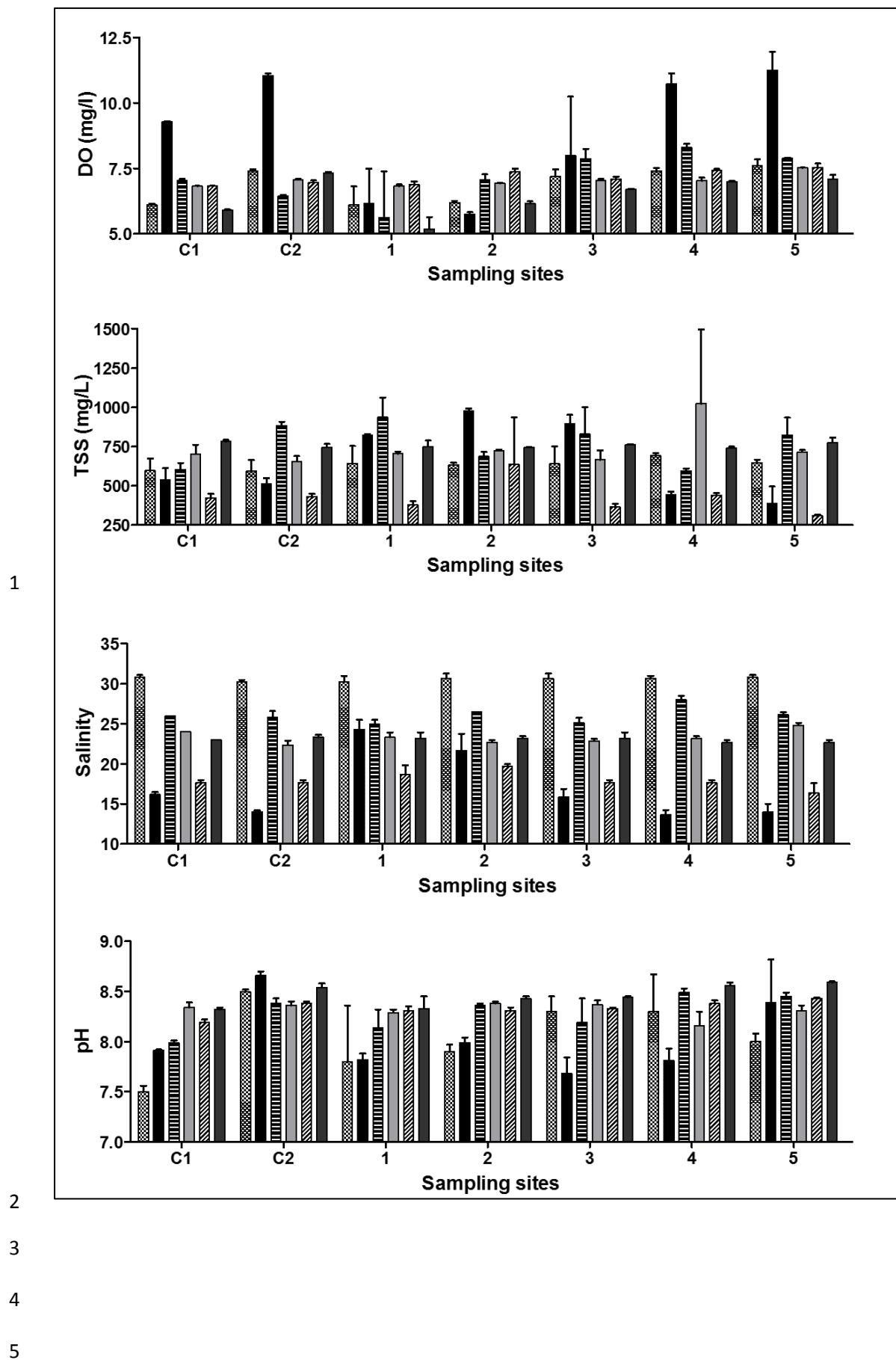
1 concentration observed on the first day after the discharge (1 PD) in sites 1 (0.01 ± 0 mg
 2 / L), 2 (0.01 ± 0.01 mg / L) and in the sedimentation basin (0.02 ± 0.01 mg / L).

3 The chlorophyll a concentration in ponds and sedimentation basin are shown in
 4 Table 1. Over the different sites, it ranged from 1.8 to 20 mg / L, except for the first day
 5 post discharge, where the sites 1, 2 and 3 showed mean values of 38.17 ± 1.89 , $41.98 \pm$
 6 1.97 and 41.47 ± 0.66 mg / L, respectively. The dissolved oxygen ranged from 5.19 to
 7 11.25 mg / L, showing little spatio-temporal variation, except in sampling 1 PD, where
 8 it was observed lower values at the effluent discharge channel. The total suspended
 9 solids (TSS) ranged from 308.3 to 1023.3 mg / L, and concentrations increased in PD 1
 10 and 5 PD with the plume reaching 20 m away from the effluent channel discharge. The
 11 salinity varied from 13.33 to 31. There was an increase in the sites next to the margin as
 12 a result of ponds water (that had higher salinity) discharge. The pH ranged between 7.47
 13 and 8.56, having no apparent relation with the release of effluents. The water turbidity
 14 was 1.93 to 43.83 NTU, and it can be noticed an increase in sites 1, 2 and 3 on 1 PD and
 15 site 1 on 5 PD (Fig. 2). The temperature had only seasonal variation between sampling
 16 stations (20.67°C to 30.8°C).



17

18



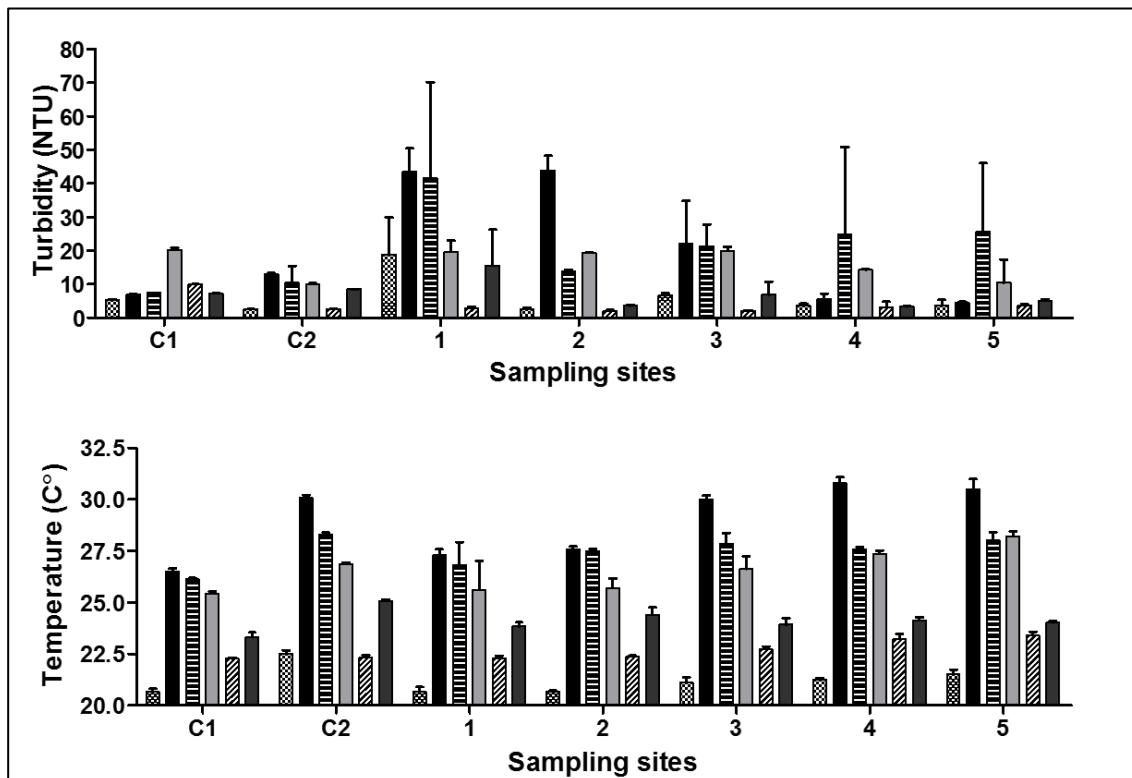


Figure 2. Water quality (mean \pm SD) over sampling period in sites control (C1 and C2), effluents channel discharge (1), 20 m (2 and 3) and 30 m (4 and 5) from the effluents channel discharge. TAN = Total Ammonia Nitrogen; DO = Dissolved Oxygen; TSS = Total Solid Suspended.

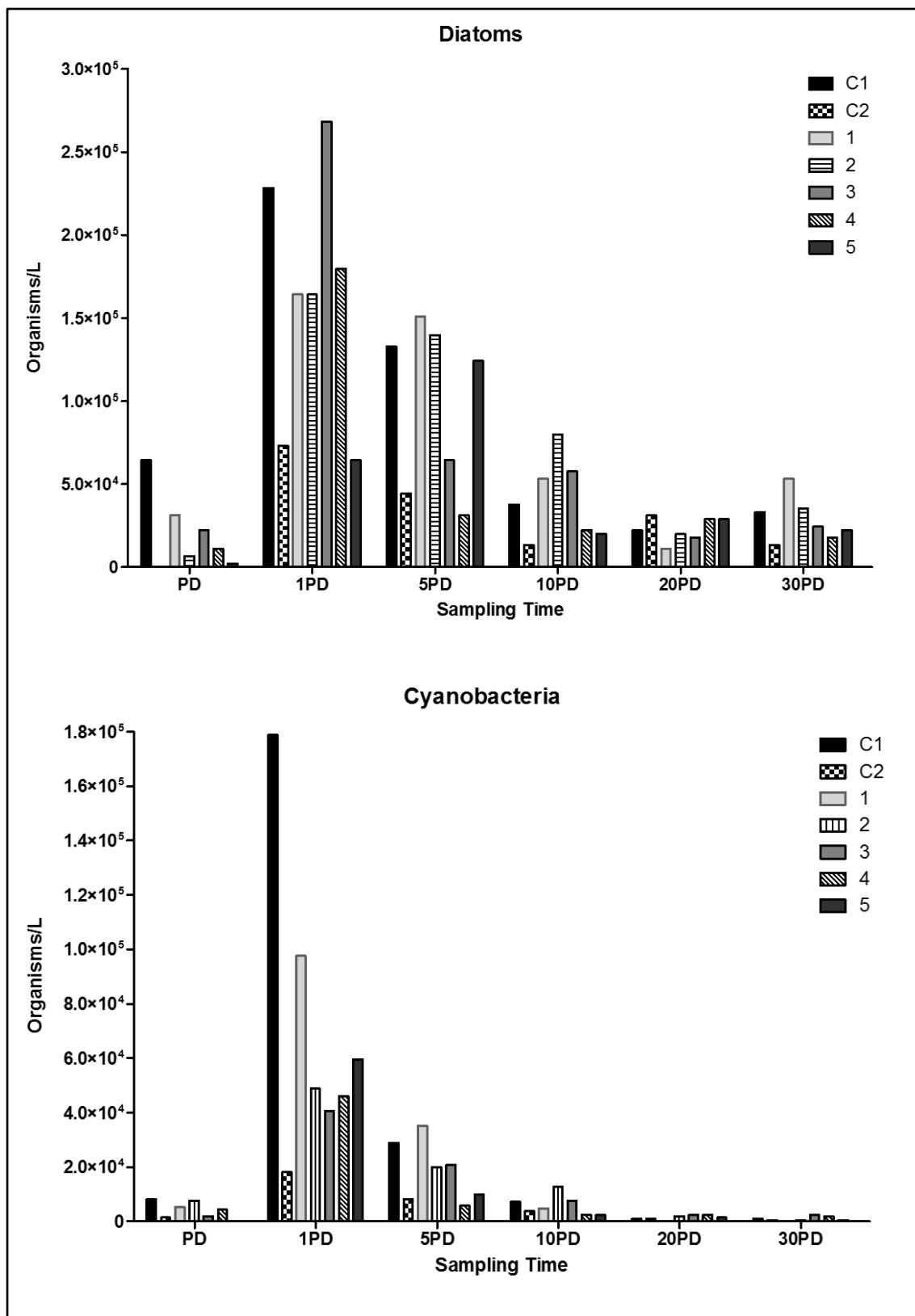
Table 1. Water quality values (mean \pm SD) from ponds 1 and 2 and sedimentation basin in the moment of shrimp harvest.

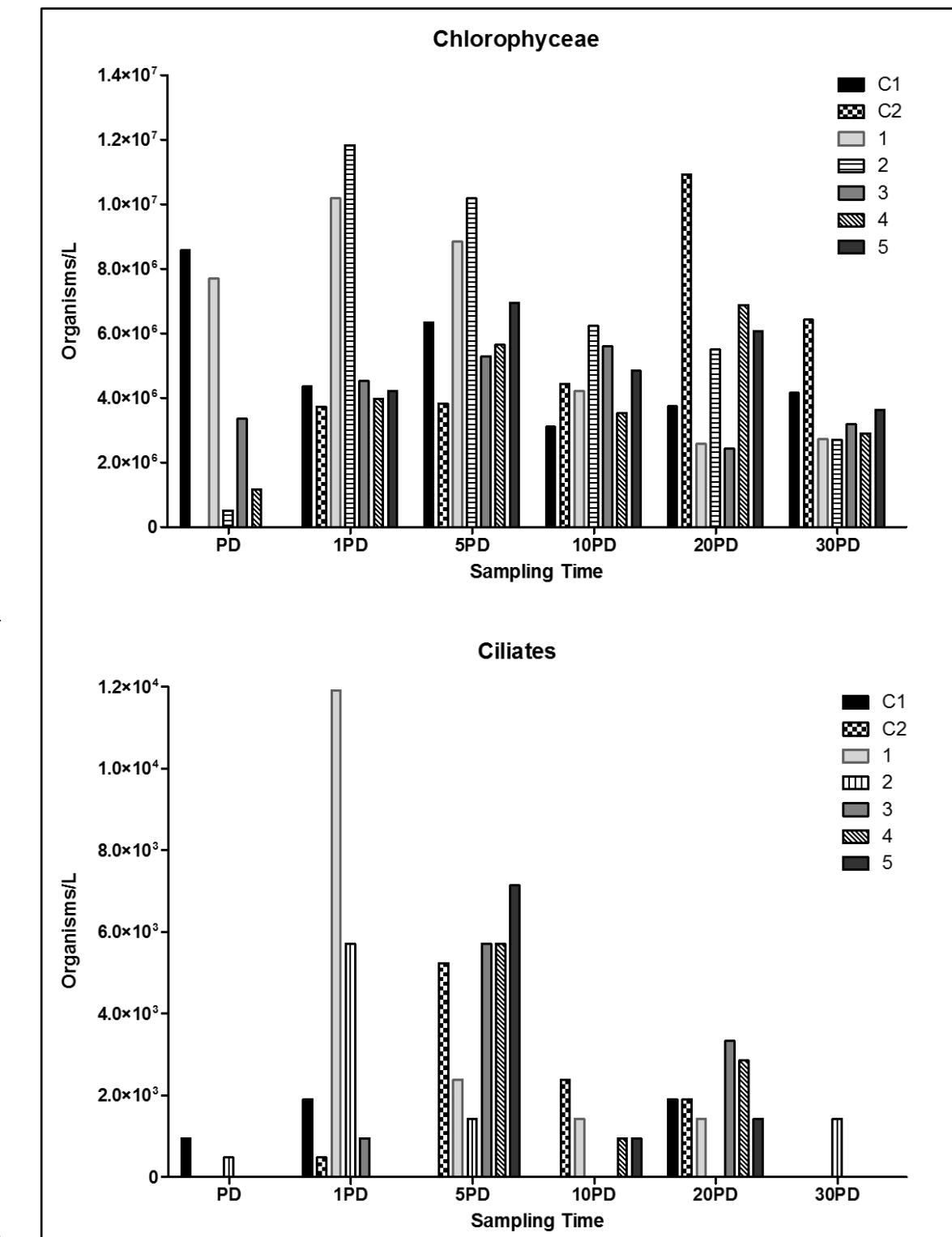
	Pond 1	Pond 2	Sedimentation Basin
Total ammonia nitrogen (mg/L)	0,94 \pm 0,05	0,69 \pm 0,03	1,1 \pm 0,03
Chlorophyll a (μg/L)	32,96 \pm 7,14	36,47 \pm 2,01	40,75 \pm 0,55
Dissolved oxigen(mg/L)	3,73 \pm 0,02	4,50 \pm 0,03	3,34 \pm 0,02
Total solid suspended (mg/L)	580 \pm 62,45	635 \pm 108,28	821,67 \pm 79,11
Salinity	24,17 \pm 0,29	23 \pm 0,00	23 \pm 0,00
pH	7,79 \pm 0,05	7,89 \pm 0,01	7,67 \pm 0,04
Turbidity (NTU)	25,8 \pm 0,17	26,17 \pm 0,06	8,24 \pm 0,01
Temperature (°C)	26,1 \pm 0,10	26,43 \pm 0,12	26,43 \pm 0,21

Phytoplankton and protozooplankton communities

The phytoplankton and protozooplankton communities in the estuary were analyzed through the identification and quantification of the following groups: chlorophyceae, diatoms, cyanobacteria and ciliates. Their spatio-temporal distribution is shown in Figure 3. The predominant group was the chlorophyceae, followed by

1 diatoms, cyanobacteria and ciliates. Diatoms, cyanobacteria and ciliates were more
 2 abundant at all points on 1 PD and there was also an increased concentration of
 3 Chlorophyceae on 1 PD comparing to pre-harvest sampling.

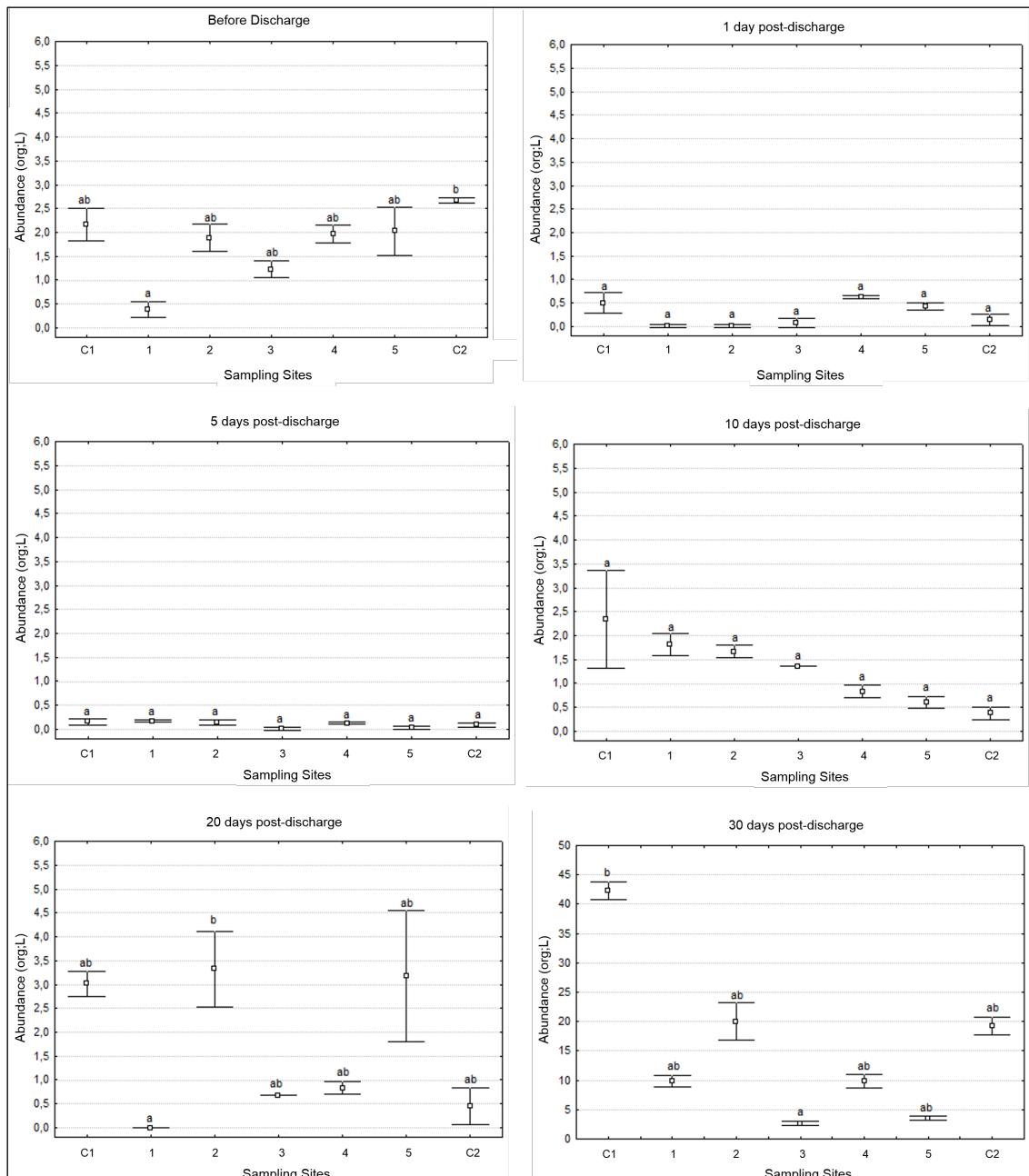




2
3 Figure 3. Phytoplankton and protozooplankton communities spatio-temporal variation in sites
4 control (C1 and C2), in front of effluents discharge channel (1), 20 m (2 and 3) and 30 m (4 and
5) from the effluents discharge channel.

6

1 Zooplankton abundance was low in almost all samples, except in 30PD when
 2 some sites showed peaks (Fig. 4). Copepods were dominants, mainly *Acartia tonsa*
 3 species.



6
 7 Figure 4: Total abundance of meso-zooplankton (org/L) over different sites distributed
 8 spatiotemporally. * Figures in different scales.
 9

10 DISCUSSION

11
 12 The main nutrient sources in shrimp farming come from aquafeeds and organic
 13 and inorganic fertilizers used to increase the *production* of phytoplankton

1 (HARGREAVES, 1998). Only the commercial feeds are responsible for the input of
2 76% of nitrogen and 83.4% of the phosphorus in the system, depending on management
3 strategies (PÁEZ-OSUNA et al., 1997). Hence, the pond water becomes richer in
4 nutrients, suspended solids, plankton and oxygen demand, when compared to coastal
5 receiving water bodies (SCHWARTZ; BOYD, 1994). As there were no water exchange
6 during the cycle period, nutrient absorption and plankton production dynamics were
7 gradually increasing until the end of the grow out period when the feed intake was
8 higher.

9 All ponds water is discharged in a vegetated sedimentation basin prior to reach
10 the estuary water receiving. The sedimentation basin improves effluents water quality
11 through the transport of suspended solids, nutrient cycling, biomass production, nutrient
12 absorption by plants and animals, and distribution of organic matter and oxygen
13 (SHPIGEL et al., 2013). JACKSON et al. (2003) report a reduction of 60% of TSS,
14 23% of the total nitrogen and 35% of total phosphorus in the effluent after the passage
15 in the sedimentation basin. However, even with the basin, part of nutrients and plankton
16 eventually reach the estuary. These compounds may favor an increase in the natural
17 productivity of the water body receptor (TACON; FORSTER, 2003) or may be quickly
18 diluted.

19 Accordingly, the local hydrodynamics and the environment carrying capacity
20 can advantage the low residence time of the exported material. The Patos Lagoon is
21 characterized by its strong hydrological interaction between wind and river discharge
22 (KJERFVE, 1986; MÖLLER; FERNANDES, 2010). MÖLLER et al. (1996) also report
23 that in shallow areas of the Patos Lagoon estuary predominate NE winds, which induce
24 water exchanges and low residence times. The low estuarine salinity observed in 1 PD
25 may be related to the receding water flows caused by wind and / or rain, which may also
26 have diluted the nutrients and organisms from the effluents. The results indicate that
27 water quality parameters had little variation during the trial, except for the temperature.
28 Total ammonia nitrogen, chlorophyll a and turbidity were the most variable parameters.
29 However, these alterations occurred only in the first days after the effluent discharge
30 and they were restricted to 20m away from the EDC, showing that the estuary had a
31 quick recovery.

32 The nutrients load is often used as an indicator of water quality. The sampling
33 sites 1, 2 and 3 exhibited the higher TAN values in 1 PD, whereas nitrite, nitrate and
34 phosphate in the effluent were practically null. The phytoplankton community play an

1 important role in the nitrogen absorption preventing the accumulation to toxic levels in
2 shrimp ponds (BURFORD, 1997; GLIBERT; BURFORD, 1999), and consequently in
3 the effluent. Similar values are reported to the Patos Lagoon estuary (KANTIN;
4 BAUMGARTEN 1982; ABREU et al, 1995) and to areas close to shrimp farms
5 (BRIGGS; FUNGE-SMITH, 1994; BURFORD, 1997; CARDOZO et al, 2011).

6 The concentration of chlorophyll a also showed little variability over the sample
7 period, and the values are close to those reported in the literature for the region
8 (ABREU et al, 2010; CARDOZO et al 2011), despite the higher value in 1 PD sample.
9 The highest concentrations were noted next to the effluents discharge channel, and
10 values were elevated until sample 5 PD. This is probably due the high levels also found
11 in the ponds and in the sedimentation basin, not being specifically an increase of the
12 primary productivity in the environment due to the effluents release.

13 In general, cyanobacteria, diatoms and ciliates showed similar pattern in space
14 and time, having their density increased in 1 PD sample, not necessarily related to the
15 discharge of effluents. In addition to the effects on the water quality, restricted to 20 m
16 away from the effluents discharge channel, freshwater input by rains may have been an
17 important nutrients source for phytoplankton and protozooplankton development.
18 However, no data on water quality between samples 20 PD and 30 PD is available to
19 prove the hypothesis of increased levels of nutrients in the water. ABREU et al. (2010)
20 found no relationship between chlorophyll a levels in Patos Lagoon estuary and abiotic
21 factors, but there was a significant relationship between the mean annual values of
22 chlorophyll a and average rainfall in the estuary.

23 The salinity of Patos Lagoon estuary have a defined pattern: more saline waters
24 when SO winds are predominant and have low river discharge, and less saline waters in
25 ebb from the increase in river discharge (MÖLLER; FERNANDES, 2010).
26 CARDOZO; ODEBRECHT (2012) also reported that major differences in salinity
27 between the pond and the environment result in lower levels of chlorophyll a and
28 primary production, because the immediate inhibiting on microalgae growth. Thus, it
29 may also have inhibited the growth of phytoplankton and protozooplankton released
30 through effluents.

31 The ciliates abundance and diversity has been used as a water quality indicator
32 and ecosystems dynamics (FOISSNER, 1988). Heterotrophic ciliates feed on
33 microorganisms in aquatic ecosystems and they are important in the energy flow
34 (SHERR; SHERR, 1988; DECAMP et al., 2003). These organisms serve as food for

1 fish larvae, playing a key role in the link between the microbial loop and the higher
2 trophic levels (FUKAMI ET AL., 1999). YANG et al. (2011) showed that the
3 community and abundance of protozooplankton usually follows the spatial dynamics
4 and abundance of phytoplankton, which was observed in this study.

5 Nonetheless, there is another hypothesis on phytoplankton and protozooplankton
6 increased abundance on sample 1 PD. The local hydrodynamics may have dispersed the
7 effluent plume, which could indicate that the chosen control sites were not effectively
8 far enough from the EDC. No sample of ponds was collected to analyze the composition
9 and abundance of these organisms, being impossible to determine the main groups
10 exported to the environment by the discharge of effluents. In culture ponds the
11 occurrence of phytoplankton species can be temporary or long-term (ALONSO-
12 RODRÍGUEZ; PÁEZ-OSUNA, 2003). But there is no consent on the dominance of a
13 certain group since variability depends on several factors such as light, water
14 temperature, nutrient availability and predation by meso-zooplankton (BURFORD,
15 1997; FUJITA; ODEBRECHT, 2007; ABREU et al., 2010). Nevertheless, even if the
16 effluent has been responsible for the increase in phytoplankton and protozooplankton
17 abundance in 1 PD, the lack of favorable conditions for their development dispersed
18 these organisms.

19 The zooplankton community was dominated almost exclusively by the copepod
20 *Acartia tonsa*. This is a dominant estuarine species that depends partly on tide transport
21 mechanisms (MONTÚ et al., 1998). This species is significant in the secondary
22 production of estuary food web, and several studies have focused in the Acartiidae
23 family (IRIGOIEN; CASTLE, 1995; ARA, 2001). The zooplankton concentration was
24 low over the sampling sites and showed few oscillations, except for sampling 30 PD.
25 Low zooplankton abundance is reported by CARDOZO et al. (2011) for the same
26 region. Despite the increase in the abundance of cyanobacteria, diatoms and ciliates in
27 sample 1 PD, there was no increase in zooplankton abundance. That could be expected
28 because of the greater availability of food. The occurrence and abundance of
29 zooplankton species in Patos Lagoon is mainly determined by seasonal variations of
30 salinity, temperature, wind direction and intensity, freshwater discharge and food supply
31 (MCLAREN; CORKETT, 1981; MONTÚ et al, 1998; MUXAGATA et al 2012).
32 However, it was not possible to identify the relationship between the increased
33 zooplankton abundance in sample 30 PD and the tested parameters that could affect
34 these organisms. HIRST; BUNKER (2003) report that the chlorophyll a can be used as

1 a good indicator of food availability for copepods. A constant concentration of
2 chlorophyll a shows that herbivory action was not markedly present, different from the
3 results of CARDOZO et al. (2011), which reported an opposite pattern between
4 chlorophyll a and zooplankton density levels. The estuarine water temperature, which
5 seems to influence the reproduction of *A. tonsa* (MONTÚ et al., 1997), showed little
6 variation during the sampling period. Likewise, temporal data on wind strength
7 (INMET) shows that the wind was constant and had low speed during the sample
8 period. The input of saline water into the estuary favors marine species such as *A. tonsa*
9 to dominate the zooplankton community (MONTÚ et al., 1997). Analysis on salinity
10 data between samples 20 PD and 30 PD also revealed no salt wedge from entering the
11 estuary that could be related to this abundance increase. One possible explanation is the
12 decrease in predation action since several species of fish larvae and juveniles depend on
13 Patos Lagoon estuary to feed (VIEIRA et al., 1998). Sampling 30 PD coincided with the
14 transition between summer and autumn, and the estuary temperature has great influence
15 on the ichthyoplankton, where the greatest abundance of fish eggs and larvae occurs in
16 summer (SINQUE; MUELBERT 1998).

17 In addition to the environmental variables favorable to disperse the effluents,
18 other factors may have contributed to the low pollution potential of the effluents.
19 According to ALONSO-RODRÍGUEZ; PÁEZ-OSUNA (2003), feeding management
20 and the use of appropriate technology can limit shrimp farming environmental impact.
21 Despite the changes observed in some water quality parameters, the levels of
22 chlorophyll a and phytoplankton, protozooplankton and meso-zooplankton abundance
23 in the first days after the release of effluents suggest that the carrying capacity of
24 environment was not exceeded.

25 The biotic and abiotic parameters analyzed in this study indicate that the proper
26 management and the favorable environmental conditions were extremely important on
27 generating low potential impact effluent in the shrimp farm. All parameters were in
28 accordance with the environmental standards set out in Brazilian legislation, having
29 minimal environmental impacts on Patos Lagoon estuary.

30 The authors are grateful to Denise Aernoudts (shrimp farm owner), The Ministry of
31 Fisheries and Aquaculture (MPA), Brazilian Council of Research (CNPq) and
32 Coordination for the Improvement of Higher Level or Education Personnel (CAPES).
33 L.H. Poersch and Paulo Cesar Abreu received productivity research fellowship from

- 1 CNPq.
- 2
- 3 REFERENCES
- 4 ABREU, P. C.; BIDDANDA, B.; ODEBRECHT, C. Bacterial dynamics of the Patos
5 Lagoon estuary, Southern Brazil (32° S, 52° W): relationships with
6 phytoplankton production and suspended material. **Estuar. Coast Shelf Sci.**, V.
7 35, p. 621-635, 1992.
- 8 ABREU P. C.; HARTMANN, C.; ODEBRECHT, C. Nutrient-rich Saltwater and, its
9 Influence on the Phytoplankton of the Patos Lagoon Estuary, Southern Brazil.
10 **Estuar. Coast Shelf Sci.**, V. 40, p. 219–229, 1995.
- 11 ABREU, P. C.; ODEBRECHT, C. Bactérias e Protozooplâncton. In: SEELIGER, U.;
12 ODEBRECHT, C.; CASTELLO, J.P. (Eds.). **Os Ecossistemas Costeiro e**
13 **Marinho do Extremo Sul do Brasil**. Rio Grande: Editora Ecoscientia, 1998. p.
14 40-41.
- 15 ABREU, P. C.; BERGESCH, M.; PROENÇA, L. A.; ODEBRECHT, C. Short- and
16 long- term chlorophyll a variability in the shallow microtidal Patos Lagoon
17 estuary, Southern Brazil. **Estuar. Coast.**, V. 33, p. 554-569, 2010.
- 18 ALONSO-RODRÍGUES, R.; PÁEZ-OSUNA, F. Nutrients, phytoplankton and harmful
19 algal blooms in shrimp ponds : a review with special reference to the situation in
20 the Gulf of California. **Aquaculture**, V. 219, p. 317–336, 2003.
- 21 AMINOT, A.; CHAUSSEPIED, M. **Manuel des analyses chimiques en milieu marin**.
22 Brest: Centre National pour L'Exploitation des Oceans, 379 pp, 1983.
- 23 ARA, K. Temporal variability and production of the planktonic copepods in the
24 Cananéia Lagoon estuarine system, São Paulo, Brazil: 2 - *Acartia lilljeborgi*.
25 **Plankton Biol. Ecol.**, V. 48, p. 35–45, 2001.

- 1 BENDSCHNEIDER, K.; ROBINSON, R. J. A new spectrophotometric method for the
2 determination of nitrite in sea water. **J. Mar. Res.**, V. 11, p. 87-96, 1952.
- 3 BERGESCH, M.; ODEBRECHT, C. Análise do fitoplâncton, protozooplâncton e de
4 alguns fatores abióticos no estuário da Lagoa dos Patos. **Atlântica**, V. 19, p. 31–
5 50, 1997.
- 6 BOLTOVSKOY, D. Submuestro. In: _____. **Atlas del zooplancton del Atlántico
7 Sudoccidental y métodos de trabajo con el zooplancton marino**. Mar del
8 Plata: INIDEP, 1981. p. 143-146.
- 9 BRIGGS, M. R. P.; FUNGE-SMITH, S. J. A nutrient budget of some intensive marine
10 shrimp ponds in Thailand. **Aquacult. Fish. Manage.**, V. 25, p. 789–811, 1994.
- 11 BURFORD, M. Phytoplankton dynamics in shrimp ponds. **Aquacult. Res.**, V. 28, p.
12 351–360, 1997.
- 13 BURFORD, M. A.; GLIBERT, P. M. Short-term nitrogen uptake and regeneration in
14 early and late growth phase shrimp ponds. **Aquacult. Res.**, V. 30, p. 215–227,
15 1999.
- 16 BURFORD, M. A.; WILLIAMS, K. C. The fate of nitrogenous waste from shrimp feed.
17 **Aquaculture**, V. 198, p. 79-93, 2001.
- 18 CARDOZO, A. P.; BRITTO, V. O.; ODEBRECHT, C. Temporal variability of
19 plankton and nutrients in shrimp culture ponds vs. adjacent estuarine water.
20 **Panam. J. Aquat. Sci.**, V. 6, p. 28–43, 2011
- 21 CARDOZO, A. P.; ODEBRECHT, C. Effects of shrimp pond water on phytoplankton:
22 importance of salinity and trophic status of the receiving environment.
23 **Aquacult. Res.**, V. 45, p. 1-11, 2012.

- 1 CASÉ, M.; LEÇA, E. E.; LEITÃO, S. N.; SANT'ANNA E. E.; SCHWAMBORN, R.;
2 JUNIOR, A. T. M. Plankton community as an indicator of water quality in
3 tropical shrimp culture ponds. **Mar. Pollut. Bull.**, V. 56, p. 1343–1352, 2008.
- 4 CHEN, Q. H.; TAM, N. F. Y.; SHIN, P. K. S.; CHEUNG, S. G.; XU, R. L. Ciliate
5 communities in a constructed mangrove wetland for wastewater treatment. **Mar.**
6 **Pollut. Bull.**, V. 58, p. 711–719, 2009
- 7 CHO, C.; HYNES, J.; WOOD, K.; YOSHIDA, Y. H. Development of high nutrient
8 dense, low-pollution diets and prediction of aquaculture wastes using biological
9 approaches. **Aquaculture**, V. 124, p. 293-305, 1994.
- 10 COUTINHO, T. M. P.; BRITO, A. C.; PEREIRA, P.; GONÇALVES, A. S.; TERESA,
11 M. A phytoplankton tool for water quality assessment in semi-enclosed coastal
12 lagoons : Open vs closed regimes. **Estuar. Coast. Shelf Sci.**, V. 110, p. 134–
13 146, 2012.
- 14 DECAMP, O.; CODY, J.; CONQUEST, L.; DELANOY, G.; TACON, A. G. J. Effect
15 of salinity on natural community and production of Litopenaeus vannamei
16 (Boone) within experimental zero-water exchange culture systems. **Aquacult.**
17 **Res.**, V. 34, p. 345-355, 2003.
- 18 FOISSNER, W. Taxonomic and nomenclatural revision of Sladeciek's list of ciliates
19 (Protozoa: Ciliophora) as indicators of water quality. **Hydrobiologia**, V. 166, p.
20 1-64, 1988.
- 21 FAO. **The State of World Fisheries and Aquaculture**. Roma: Food and Agriculture
22 Organization of United Nations, 2012. 230 pp.
- 23 FAO. **The State of World Fisheries and Aquaculture**. Roma: Food and Agriculture
24 Organization of United Nations, 2014. 243 pp.
- 25 FUJITA, C. C. O.; ODEBRECHT, C. Short-term variability of chlorophyll a and
26 phytoplankton composition in a shallow area of the Patos Lagoon estuary
27 (Southern Brazil). **Atlântica**, V. 29, p. 93-107, 2007.

- 1 FUKAMI, K.; WATANABE, A.; FUJITA, S.; YAMAOKA, K.; NISHIJIMA, T.
- 2 Predation on naked protozoan microzooplankton by fish larvae. **Mar. Ecol.-**
- 3 **Prog. Ser.**, V. 185, p. 285-291, 1999.
- 4 HARGRAVE, B. T. Impacts of man's activities on aquatic system. In: BARNES, R. S.
- 5 K.; MANN, K. H. (Eds.). **Fundamental of Aquatic Ecosystems**. Oxford:
- 6 Blackwell Science, 1991. cap. 13, p. 245-264.
- 7 HARGREAVES, J. A. Nitrogen biogeochemistry of aquaculture ponds. **Aquaculture**,
- 8 V. 166, p. 181–212, 1998.
- 9 HASLE, G. R. The inverted microscope method. In: SOURNIA, A. (Ed.).
- 10 **Phytoplankton manual**. Paris: UNESCO, 1978. p. 88-96.
- 11 HERBECK, L. S.; UNGER, D.; WU, Y.; JENNERJAHN, T. C. Effluent, nutrient and
- 12 organic matter export from shrimp and fish ponds causing eutrophication in
- 13 coastal and back-reef waters of NE Hainan, tropical China. **Cont. Shelf Res.**, V.
- 14 57, p. 92–104, 2013.
- 15 HIRST, A. G.; BUNKER, A. J. Growth of marine planktonic copepods: global rates and
- 16 patterns in relation to chlorophyll a, temperature, and body weight. **Limnol.**
- 17 **Oceanogr.**, V. 48, p. 1988–2010, 2003.
- 18 INMET. Instituto Nacional de Meteorologia. Fonte: <http://www.inmet.gov.br/portal/>
- 19 IRIGOIEN, X.; CASTEL, J. Feeding rates and productivity of the copepod *Acartia*
- 20 *bifilosa* in a highly turbid estuary; the Gironde (SW France). **Hydrobiologia**, V.
- 21 322, p. 115–125, 1995.
- 22 JACKSON, C.; PRESTON, N.; BURFORD, M. A.; THOMPSON, P. J. Managing the
- 23 development of sustainable shrimp farming in Australia: the role of
- 24 sedimentation ponds in treatment of farm discharge water. **Aquaculture**, V. 226,
- 25 p. 23–34, 2003.

- 1 JACKSON, C.; PRESTON, N.; THOMPSON, P. J. Intake and discharge nutrient loads
2 at three intensive shrimp farms. **Aquacult. Res.**, V. 35, p. 1053–1061, 2004.
- 3 KANTIN, R.; BAUMGARTEN, M. G. Z. Observações hidrográficas no estuário da
4 Lagoa dos Patos: distribuição e flutuações dos sais nutriente. **Atlântica**, V. 5, p.
5 76-92, 1982.
- 6 KJERFVE, B. Comparative oceanography of coastal lagoons. In: WOLFE, D. A. (ed.)
7 **Estuarine Variability**. New York: Academic Press, 1986. p. 63-81.
- 8 MCLAREN, I. A.; CORKETT, C. J. Temperature-dependent growth and production by
9 a marine copepod. **Can. J. Fish. Aquat. Sci.**, V. 38, p. 77–83, 1981.
- 10 MOLLES, P.; BUNGE, J. Shrimp farming in Brazil: An Industry Overview. Report
11 prepared under the World Bank, NACA, WWF and FAO Consortium Program
12 on Shrimp Farming and the Environment. Work in Progress for Public
13 Discussion. Published by the Consortium, 2002, 26p.
- 14 MÖLLER, O. O.; LORRENZZENTI, J. A.; STECH, J. L.; MATA, M. M.. The Patos
15 Lagoon summertime circulation and dynamics. **Cont. Shelf Res.**, V. 16, p. 335-
16 351, 1996.
- 17 MÖLLER, O.; FERNANDES, E. Hidrologia e hidrodinâmica. In: SEELIGER, U.;
18 ODEBRECHT, C. (Eds.). **O estuário da Lagoa dos Patos: Um século de**
19 **transformações**. Rio Grande: FURG, 2010. p. 17-30.
- 20 MONTÚ, M.; DUARTE, A. K.; GLOEDEN, E I. M. Zooplankton. In.: SEELIGER, U.;
21 ODEBRECHT, C.; CASTELLO, J. P. (Eds.). **Subtropical Convergence**
22 **Environments: The Coast and Sea in the Southwestern Atlantic**. Berlin:
23 Springer-Verlag, 1997. p. 40-43.

- 1 MONTÚ, M.; DUARTE, A. K.; GLOEDEN, E I. M. Zooplâncton. In: SEELIGER, U.;
2 ODEBRECHT, C.; CASTELLO, J.P. (Eds.). *Os Ecossistemas Costeiro e*
3 *Marinho do Extremo Sul do Brasil*. Rio Grande: Ecoscientia, 1998. p. 43-46.
- 4 MUXAGATA, E.; AMARAL, W. J. A.; BARBOSA, C. N. Acartia tonsa production in
5 the Patos Lagoon estuary, Brazil. **ICES J. Mar. Sci.**, V. 69, p. 475-482, 2012.
- 6 NAYLOR, R. L.; GOLDBURG, R. J.; MOONEY, H.; BEVERIDGE, M.; CLAY, J.;
7 FOLKE, C.; KAUTSKY, N.; LUBCHENCO, J.; PRIMAVERA, J.;
8 WILLIAMS, M. Nature's subsidies to shrimp and salmon farming. **Science**, V.
9 282, p. 883–884, 1998.
- 10 NIENCHESKI, L. F.; MOORE, W. S.; WINDOM, H. L. History of human activity in
11 coastal southern Brazil from sediment. **Mar. Pollut. Bull.**, V. 78, p. 209–212,
12 2014.
- 13 OLSEN, L. M.; HOLMER, M.; OLSEN, Y. **Perspectives of nutrient emission from**
14 **fish aquaculture in coastal waters: Literature review with evaluated state of**
15 **knowledge**. FHF project no. 542014. Norge: The Fishery and Aquaculture
16 Industry Research Fund, 2008. 87p.
- 17 PÁEZ-OSUNA, F.; GUERRERO-GALVÁN, S. R.; RUIZ-FERNÁNDEZ, A. C.;
18 ESPINOZA-ANGULO, R. Fluxes and mass balances of nutrients in a semi-
19 intensive shrimp farm in northwestern Mexico. **Mar. Pollut. Bull.**, V. 34, p.
20 290– 297, 1997.
- 21 REIS, E. G.; D'INCAO, F. The present status of artisanal fisheries of extreme Southern
22 Brazil: an effort towards community-based management. **Ocean Coast.**
23 **Manage.**, V. 43, p. 585-595, 2000.
- 24 SCHWARTZ, M. F.; BOYD, C. E. Channel catfish pond effluents. **Prog. Fish Cult.**, V.
25 56, p. 273–281, 1994.

- 1 SEELIGER, U. Introdução. In: SEELIGER, U.; ODEBRECHT, C. (eds.). **O estuário**
2 **da Lagoa dos Patos: Um século de transformações.** Rio Grande: FURG, 2010.
3 p. 11-13.
- 4 SHERR, E.; SHERR, B. Role of microbes in pelagic food webs: a revised concept.
5 **Limnol. Oceanogr.**, V. 33, p. 1225-1227, 1988.
- 6 SHPIGEL, M.; BEN-EZRA, D.; SHAULI, L.; SAGI, M.; VENTURA, Y., SAMOCHA,
7 T.; LEE, J. J. Constructed wetland with Salicornia as a biofilter for mariculture
8 effluents. **Aquaculture**, V. 412-413, p. 52–63, 2013.
- 9 SINQUE, C.; MUELBERT, J. H. Ictioplâncton. In: SEELIGER, U.; ODEBRECHT, C.;
10 CASTELLO, J. P. (Eds.). **Os Ecossistemas Costeiro e Marinho do Extremo Sul**
11 **do Brasil.** Rio Grande: Ecoscientia, 1998. p. 56-60.
- 12 SOKAL, R. R.; ROHLF, F. J. **Biometry: The Principles and Practice of Statistics in**
13 **Biological Research.** 3 ed. New York:W. H. Freeman and Co., 1995. 887p.
- 14 TACON, A. G. J.; FORSTER, I. P. Aquafeeds and the environment: policy
15 implications. **Aquaculture**, V. 226, p. 181–189, 2003.
- 16 UNESCO. **Manual and Guides - Chemical methods for use in marine**
17 **environmental monitoring.** France: Intergovernmental Oceanographic
18 Commission, 1983. 56p.
- 19 VIEIRA, J. P.; CASTELLO, J. P.; PEREIRA, L. E. Ictiofauna. In: SEELIGER, U.;
20 ODEBRECHT, C.; CASTELLO, J. P. (Eds.). **Os Ecossistemas Costeiro e**
21 **Marinho do Extremo Sul do Brasil.** Rio Grande: Ecoscientia, 1998. p. 60-68.
- 22 WELSCHMEYER, N. A. Fluorometric analysis of chlorophyll a in the presence of
23 chlorophyll b and pheopigments. **Limnol. Oceanogr.**, V. 39, p. 1985-1992,
24 1994.

1 YANG, E. J.; HYUN, J. H.; KIM, D.; PARK, J.; KANG, S. H.; SHIN, H. C.; LEE, S.
2 Mesoscale distribution of protozooplankton communities and their herbivory in
3 the western Scotia Sea of the Southern Ocean during the austral spring. **J. Exp.**
4 **Mar. Biol. Ecol.**, V. 428, p. 5–15, 2012.
5

1 **CAPÍTULO 2**
2

3 **IMPACTO DOS EFLUENTES DA CARCINOCULTURA SOBRE A**
4 **COMUNIDADE MACROZOOBENTÔNICA DA LAGOA DOS PATOS, BRASIL**

5
6 Artigo submetido ao periódico Anais da Academia Brasileira de Ciências (ISSN: 1678-
7 2690).

8

1 IMPACT OF SHRIMP FARM EFFLUENT ON MACROZOOBENTHOS
2 COMMUNITY IN PATOS LAGOON ESTUARY, SOUTHERN BRAZIL

3
4 **William Bauer**
5 **Bruna Machiavelli**
6 **Luis Henrique da Silva Poersch***
7

8 *Estação Marinha de Aquacultura - Instituto de Oceanografia/Universidade Federal de*
9 *Rio Grande (Rua do Hotel, 02. Bairro Cassino. 96210-030. Rio Grande, RS, Brasil)*

10
11
12 *Keywords:* Macrozoobenthos, Effluents, Sediment, *Litopenaeus vannamei*.

13
14 SHRIMP FARM IMPACT ON MACROZOOBENTHOS IN BRAZIL

15
16 Seção: Ciências Agrárias (Aquicultura).

17
18 *Corresponding author. Address: Av. Itália Km 08, Campus Carreiros, CP 474, 96201-
19 900 Rio Grande, RS, Brasil. Phone/Fax: (+55) 53 32361685. E-mail:
20 lpoersch@mikrus.com.br

1 ABSTRACT

2

3 The Pacific white shrimp *Litopenaeus vannamei* production in earth ponds has been
4 increasing in the surrounding of the Patos Lagoon estuary, Southern Brazil. This study
5 evaluated the spatio-temporal effects of shrimp farming effluent on macrozoobenthos
6 community over two cycles period (2012 and 2013). The samples were collected before
7 and after the effluents discharge in the estuary receiving and the sampling sites were
8 assigned as follows: across the effluents discharge channel (EDC), 20 m, 30 m, 100 m
9 and 250 m from the EDC. Collected sample were sieved through a 500 μ m mesh size,
10 sorted, identified to lowest possible taxonomic level and counted. Seven groups were
11 recorded (Polychaeta, Tanaidacea, Isopoda, Gastropoda, Bivalvia, Malacostraca and
12 Ostracoda) along the sampling campaigns. Spatially, the density and species richness
13 had little variability over the sampling sites in both campaigns. Temporally, during
14 2012 campaign, these indices increased over time in winter compared to summer, unlike
15 2013 campaigns, where the density decreased in colder months and species richness
16 suffer little variability. The results show that the effluents discharge not influenced the
17 macrozoobenthos community and the statistical differences recorded ($p < 0.05$) were
18 probably caused by natural fluctuations of environmental parameters, natural in
19 estuarine ecosystems.

20

1 INTRODUCTION

2

3 The artisanal fishery of *Farfantepenaeus paulensis* shrimp is a significant
4 activity in the Patos Lagoon estuary with socio-economic importance. The production is
5 extremely variable and depends on environmental conditions (Castello and Möller
6 1978; D'Incao and Reis 2002). The aquaculture of Pacific white shrimp *Litopenaeus*
7 *vannamei* in this estuary emerged as an alternative to meet the market demand of this
8 seafood. However, as a growing agribusiness its environmental sustainability should be
9 conducted carefully. Several studies evaluated the potential environmental impacts of
10 aquaculture around the world (Carroll et al. 2003; Crawford et al. 2003; Forchino et al.
11 2011), and the aquatic environmental degradation by effluents discharge is a main
12 concern.

13 The aquaculture effluents are usually rich in nutrients and organic solids
14 (Burford et al. 2003) from fertilizers, phytoplankton, unconsumed feed and animal
15 excretion (Jackson et al. 2004). This particulate organic matter input acts as a source of
16 organic carbon for a diverse community of filtering and decomposers organisms
17 (Albertelli et al. 1999). These organisms are susceptible to environmental fluctuations
18 and disorders that impact them in different time scales (Gray and Christie 1983). In
19 some cases, the disturbance caused by the effluents can cause physical and chemical
20 changes in the sediment. Thus, it can reduce the biological diversity and favor
21 opportunistic species (Johannssen et al. 1994), which affects the richness and the
22 density of the macrozoobenthic community (Heip 1995). Anaerobic zones may occur
23 and toxic compounds such as ammonia, hydrogen sulfide and methane are released
24 when the organic accumulation rates exceed the carrying capacity of the substrate
25 (Alongi et al. 1999). These extreme conditions lead to the destruction of the
26 environmental quality and community structure (Islam 2005), conducting to the
27 impoverishment / disappearance of fauna (Heilskov and Holmer 2001).

28 Several studies have focused on the evaluation and mitigation of the impacts of
29 aquaculture effluents on benthic communities (Bartoli et al. 2001; Carvalho et al. 2009;
30 Aguado-Giménez et al. 2011). These communities are used as a sensitive tool to detect
31 environmental effects (Carroll et al. 2003), and they can be useful to monitor the
32 impacts of aquaculture effluents. Benthic organisms are the main link between primary
33 producers and higher trophic levels in estuarine regions (Foreman et al. 1995).

1 Moreover, they play a crucial role in the supply and re-mineralization of organic matter
2 (Heilskov and Holmer 2001).

3 The composition and abundance of macrozoobenthos were examined in several
4 studies in Patos Lagoon estuary (Bemvenuti et al. 1978; Capitol et al. 1978; Rosa and
5 Bemvenuti 2006; Colling et al. 2007). However, studies analyzing the impact of shrimp
6 farming effluents on estuarine macrozoobenthos are scarce. This study aims to evaluate
7 the macrozoobenthos subjected to shrimp farm effluents in the Patos Lagoon estuary,
8 analyzing the spatio-temporal variations of the density and species richness.

9

10 **Material and Methods**

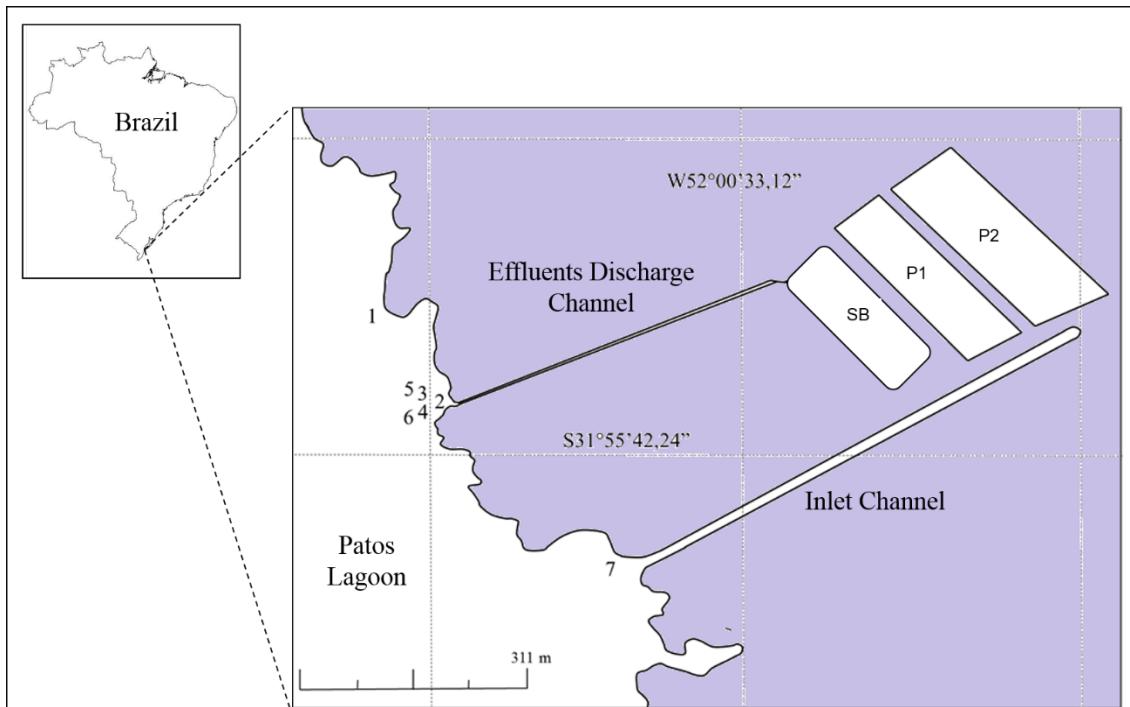
11

12 *Study area and shrimp farm characteristics*

13

14 This study was conducted in a commercial *L. vannamei* farm located in the Patos
15 Lagoon estuary, Southern Brazil (Rio Grande do Sul - 31°55'S, 52°00'W - Fig. 1). The
16 ponds with 3.75 ha area was stocked with 12 shrimps/m² (semi-intensive system).
17 Because of the low temperatures during winter, this farm works on a 1-cycle per year,
18 and after each cycle the ponds are drained and dried. During the cycle period there was
19 no effluents discharge, and water was added in the ponds only to compensate the
20 evaporation and infiltration. Before the discharge, the effluents were transferred to a
21 vegetated sedimentation basin in order to decrease the load of nitrogen, phosphorous
22 and suspended solids.

23



1
2 Figure 1: Sampling sites in the shrimp farming. 1 = Control 1, 7 = Control 2, 2 = Effluents discharge
3 channel, 3 and 4 = 20m from the channel, 5 and 6 = 30m from the channel, P1 = Pond 1, P2 = Pond 2 e
4 SB = Sedimentation basin.

5 *Sampling plan*

6 Sampling was conducted based on BACI (Before-After-Control-Impact) design
7 with some modifications (Underwood 1994). Samples were taken between summer and
8 winter during 2 cycles periods (2012 and 2013), in a total of six campaign per year.
9 Seven sites were previously chosen to water quality analysis and macrozoobenthos
10 sampling (Fig. 1): two control sites, located in 100 m and 250 m from the EDC (1 and
11 7); site 2, in front of the EDC; sites 3 and 4, located 20 m from the EDC; and sites 5 and
12 6, 30 m away from the EDC. Between February and July 2012 the sampling was
13 conducted as follows: prior to the release of the effluents (BD) and 5, 10, 30, 60 and 90
14 days after the effluents discharge (5 PD, 10 PD, 30 PD, 60 PD e 90 PD, respectively).
15 Due to an emergency shrimp harvest caused by bacterial outbreak in 2013, no BD
16 sample from this year is available. Thus, between February and May 2013 the samples
17 were performed immediately after the discharge (1 PD) and as the previous year: 5 PD,
18 10 PD, 30 PD, 60 PD and 90 PD.

20 *Water quality parameters and macrozoobenthos*

1 Temperature (WTW Oxi 3205), dissolved oxygen (WTW Oxi 3205), salinity
2 (refractometer) and pH (YSI 60) were measured *in situ* (n=3) in all sampling sites. At
3 each site, three macrozoobenthos sub-samples were taken using a corer PVC (0,008 m²,
4 20cm depth). Sample were sieved *in situ* through a 500µm mesh size, fixed in 4%
5 buffered formaldehyde to which Rose Bengal stain added. In laboratory, samples were
6 sorted and identified at lowest possible taxonomic level and counted. Samples were also
7 taken in surface layer (interface substrate/water) to analyze sediment characteristics.
8 The granulometry composition were obtained through sieving (> 0,063 mm) and
9 pipetting analysis (< 0,063 mm) according Suguio (1973).

10

11 *Statistical analysis*

12

13 The data were tested for normal distribution before choosing parametric or non-
14 parametric statistical methods. To assess significant spatial and temporal differences in
15 density and richness species of monitored points, Analysis of Variance (ANOVA 1
16 way) and post-hoc Tukey tests were performed ($P < 0.05$). When the assumptions for
17 normality and equal variance were not observed, Kruskal-Wallis followed by post-hoc
18 Dunn was used to determine significant differences.

19

20 RESULTS

21 The water column depth during the sampling period ranged from 10 to 100 cm,
22 but it has not found a relationship between the season and the highest or lowest level of
23 water column. Particle size analysis revealed predominantly sandy substrates (43-51%)
24 with low percentages of silt / clay, with small spatial variations. Temporal variations in
25 temperature (due to seasonal variations) and salinity (caused by flood flows / ebb and /
26 or rain) were observed (Table 1). The dissolved oxygen and pH values remained high
27 throughout the sample period and showed little variability.

28

29

30

31

32

1 Table 1: Abiotic parameters (mean±SD) in different sites through the sampling period (2012
 2 and 2013) in Patos Lagoon estuary before and after shrimp farm effluents discharge. *n.a. = not
 3 available.

		Temperature (°C)	Dissolved Oxygen (mg/L)	Salinity	pH
2012	BD	29,07±1,77	6,92±0,75	30,59±0,49	8,00±0,43
	5PD	26,51±1,11	7,03±1,13	26,00±1,09	8,26±0,21
	10PD	22,58±0,41	7,02±0,22	23,37±0,88	8,31±0,07
	30PD	21,26±0,71	6,44±0,82	23,00±0,51	8,44±0,12
	60PD	15,68±0,50	7,18±0,82	20,06±0,17	7,88±0,04
	90PD	11,87±0,74	8,65±0,77	15,34±0,32	7,64±0,13
2013	1PD	27,38±0,91	6,21±1,12	23,37±0,50	7,52±0,36
	5PD	24,5±0,23	8,11±0,63	30,00±1,11	7,99±0,10
	10PD	21,31±0,31	8,89±0,13	14,53±1,04	7,98±0,14
	30PD	22,83±0,95	8,63±0,41	30,53±0,63	8,08±0,27
	60PD	16,21±0,1	9,55±0,09	13,50±0,44	7,49±0,24
	90PD	*n.a.	n.a.	7,31±1,77	7,61±0,07

4
 5 Three major taxonomic groups were recorded along the sampling campaigns:
 6 Malacostraca, Polychaeta, phylum Mollusca and Ostracoda. The most diverse class was
 7 Malacostraca (6 taxa), followed by Polychaeta (5 spp.), phylum Mollusca (two species
 8 of bivalves and a Gastropoda), and Ostracoda (Table 2).

9 Table 2: Mean density (ind. m⁻²) of macrozoobenthos identified in 2012 and 2013.

Taxa	Density (ind. m ⁻²)	
	2012	2013
Polychaeta (Class)		
<i>Laeonereis acuta</i>	1.905	965
<i>Heteromastus similis</i>	1.674	1.094
<i>Alitta succinea</i>	1.259	375
<i>Parapriionospio pinnata</i>	176	0
<i>Nephtys fluviatilis</i>	172	74
Malacostraca (Class)		
<i>Kalliapseudes schubartii</i>	87	290
<i>Ampithoe</i> sp.	77	1
<i>Farfantepenaeus paulensis</i>	1	0
<i>Cassidinidea fluminensis</i>	0	1
<i>Sphaeromopsis mourei</i>	1	55
<i>Callinectes sapidus</i>	7	1
Mollusca (Phylum)		
<i>Heleobia australis</i>	7	41
<i>Tagelus plebeius</i>	3	0
<i>Erodona mactroides</i>	2	0
Ostracoda	47	0

1 In 2012 sampling, eight taxa accounted for > 99% of the total recorded density,
2 including the polychaete *Laeonereis acuta* (35.1%), *Heteromastus similis* (30.9%),
3 *Alitta succinea* (23.2 %), *Parapriionospio pinnata* (3.2%), *Nephtys fluviatilis* (3.1%),
4 crustaceans *Kalliapseudes schubartii* (1.6%), *Ampithoe* sp. (1.4%) and Ostracoda
5 (0.9%). Spatially, species richness (Kruskal-Wallis) and the density of
6 macrozoobenthos (ANOVA) showed no significant differences over the sampling sites
7 ($P > 0.05$); however, the total density and species richness over time were significantly
8 affected (Kruskal-Wallis, $P < 0.05$). It can be observed a pattern of increase in both
9 attributes during winter when compared to summer (Fig. 2A, 2C).

10 On the 2013 production cycle, seven species accounted for > 99% of the total
11 recorded density. A number of 2.848 individuals were sampled - approximately 3 times
12 smaller than the previous year: *H. similis* (37.8%), *L. acuta* (33.3%), *A. succinea*
13 (12.9%), *K. schubartii* (10%), *N. fluviatilis* (2.6%), *Sphaeromopsis mourei* (1.9%) and
14 *Heleobia australis* (1.4%). The density of organisms differed significantly ($p < 0.05$)
15 between sites in 5 PD, 30 PD, 60 PD and 90 PD, but this occurred in a random way
16 without relationship with effluents discharge. Over time, the only significant difference
17 found occurred between 1 PD and 90 PD samples ($P < 0.05$) (Fig. 2B). As in 2012,
18 macrozoobenthic diversity did not show significant spatial differences ($P > 0.05$), only
19 temporal ($P < 0.05$) (Fig. 2D).

20

21

22

23

24

25

26

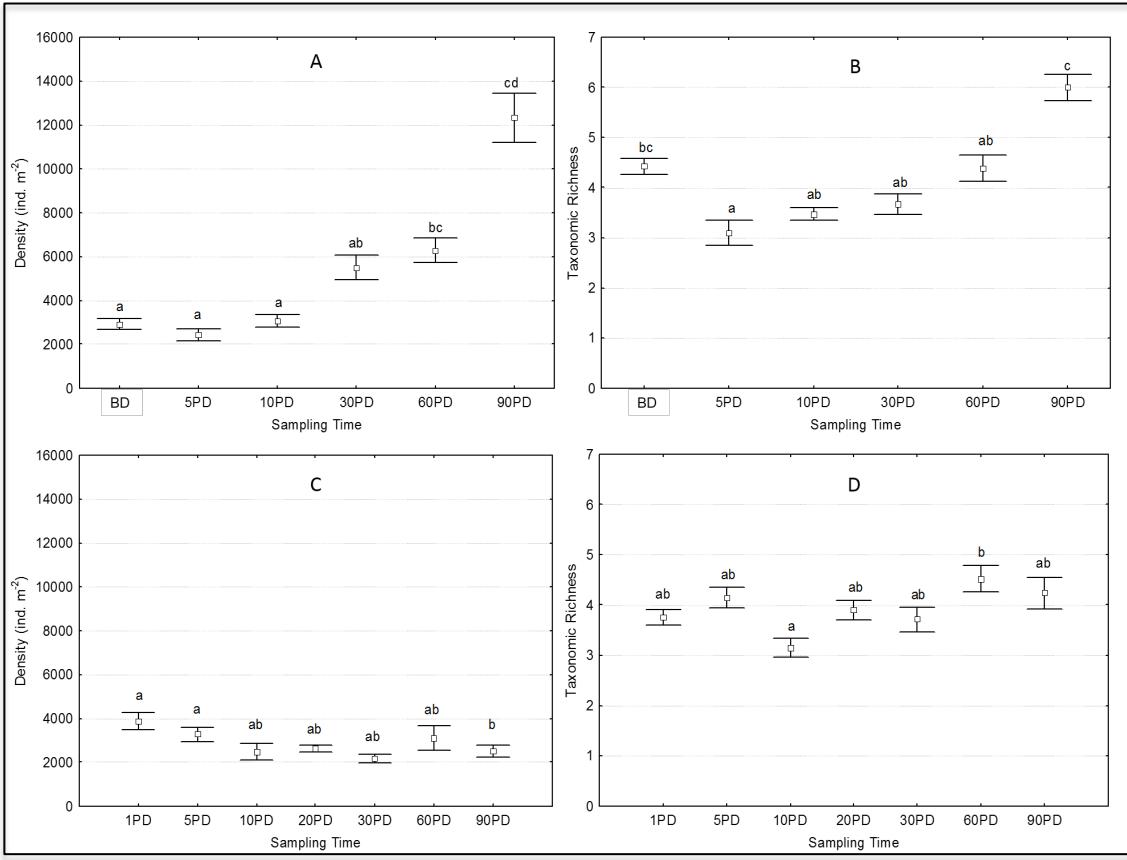


Figure 2: Macrozoobenthos parameters (mean value \pm SE) over sampling campaigns: Density (ind. m^{-2}) and Taxonomic richness in 2012 (A and B) and 2013 (C and D). Different letters denote significant ($p < 0,05$) differences. BD = Before discharge; 1PD = 1 day post-discharge; 5PD = 5 days post-discharge; 10PD = 10 days post-discharge; 20PD = 20 days post-discharge; 30PD = 30 days post-discharge; 60PD = 60 days post-discharge and 90PD = 90 days post-discharge.

DISCUSSION

The Patos Lagoon is characterized by high temperatures in summer and low rainfall rates, which favor the income of seawater into the estuary, increasing the salinity (Rosa and Bemvenuti 2006). These authors report that in winter the opposite situation occurs, which reduces the salinity. In this study, temperature and salinity showed visible seasonal patterns over the sample period. The macrozoobenthos in this lagoon has their spatial and temporal distribution affected by many factors, including water quality, substrate characteristics, and biological interactions (Bemvenuti and Colling 2010). However, environmental parameters such as temperature and salinity may have a significant influence on this community that inhabits subtropical and temperate regions with marked seasonal fluctuations (Gray and Elliott 2009). The macrozoobenthos community did not show a spatial variability in both sampling

1 campaigns (2012 and 2013), since the differences were punctual and not related to the
2 effluents release due to their random nature.

3 The 2012 sampling began in the summer and finished in winter. In 2013, the
4 sample campaign began in the summer and ended in the autumn. Temporally, the
5 macrozoobenthos variability was quite marked during 2012 and less evidenced in 2013.
6 In 2012 cycle period, the density and species richness was lower in summer when
7 compared to winter, and an opposite situation occurred in 2013. A pattern usually
8 observed for macrozoobenthos in Patos Lagoon estuary is the higher densities in
9 summer than in the winter (Bemvenuti 1987; Rosa and Bemvenuti 2006) due to an
10 increase in reproductive activities of many benthic species, despite the predation
11 pressure from higher trophic levels (Bemvenuti 1998). The presence/absence of
12 predators is an important factor to be considered, since many macrozoobenthic species
13 are important food items to organisms such as the blue crab *Callinectes sapidus*, the
14 corvina *Micropogonias furnieri* and the shrimp *Farfantepenaeus paulensis* (Bemvenuti
15 1987, 1997). In fact, benthic animals are directly and indirectly involved in most
16 physical and chemical processes that occur in estuaries (Reish 1980), which makes it
17 difficult to predict what exactly causes this change in macrozoobenthos pattern.

18 Water pollution from intensive prawn farms negatively affects adjacent
19 ecosystems (Ellison 2008). A reduction of biological diversity is the first consequence
20 of coastal degradation by nutrients enrichment (Mouillot et al. 2005) and accordingly
21 benthonic organisms, particularly polychaetes have been used as stress indicator in
22 environmental impact studies (Pocklington et al. 1994). Generally, environments with
23 high accumulation of organic matter (dissolved and/or particulate) possess substrates
24 containing acids and hypoxic/anoxic sediment, which promotes the anaerobic
25 decomposition increasing the production/release of toxic compounds. Thence, these
26 compounds can also change the benthic communities structure (Jackson et al. 2004). In
27 reduced environments is observed a decrease in benthic diversity (Lorenzen et al. 1987),
28 a process that favors the settlement and/or re-colonization by opportunistic species due
29 to the lack of competitors (Rosa and Bemvenuti 2006). In fact, the organic enrichment
30 can cause sudden changes in community structure, reducing macrozoobenthic
31 community assembly to some tolerant species (Pearson and Rosenberg 1978). Benthic
32 macroinvertebrates can be used as indicators of the changes and variability in estuarine
33 ecosystems since they have relatively long life-spans, the different species exhibit

1 tolerances to stress, play an important role in cycling nutrients and materials and they
2 are fundamental providing links to higher levels (Dauvin 2007).

3 It was observed that in both sample campaigns there was a predominance of
4 polychaetes, which play an important role in benthic communities acting recycling and
5 reworking the nutrients, linking the trophic web (Hutchings 1998). They have been
6 widely used as enrichment indicators (Ansari et al. 1986) because organic enrichment
7 can decrease their richness and favor opportunistic species (Tomassetti and Porrello
8 2005). Dominance of polychaetes in terms of density and species composition is also
9 reported by Murugesan et al. (2009) in a study evaluating the impact of shrimp farming
10 in India. Among the five species found in this class, *L. acuta*, *H. similis* and *A. succinea*
11 were the most abundant. *A. succinea* and *H. similis* are commonly recorded in the
12 estuarine-lagoon complex of temperate and subtropical regions of the Atlantic
13 Southwest coast (Pagliosa and Barbosa 2006). These species behavior allow the
14 maintenance of high densities through escape mechanisms, such as burial capacity, high
15 mobility and reproductive strategies that ensure a rapid recolonization of the
16 environment. However, according Bemvenuti et al. (1997) the abundance of these
17 species tends to decrease considerably after environmental disturbances events.

18 The polychaete *H. similis* is a subsurface deposit-feeder that has no clear
19 seasonal pattern and suffers low predation effects (Rosa and Bemvenuti 2006). Its burial
20 ability is considered the main strategies of tolerance to disturbances in the substrate and
21 escape predation, since many epifaunal predators act in the surface layers of the
22 sediment (Bemvenuti, 1988). *L. acuta* is a deposit-feeder species with high abundance
23 and biomass that inhabits mixoaline waters (Pagliosa and Barbosa 2006) from the
24 Northeast Brazil to Southern Argentina (Omena and Amaral 2001). This species is
25 found in regions of muddy substrate with high organic matter content (Pagliosa and
26 Barbosa 2006) and it feeds on this content in the substrate (Olivier 1995), increasing the
27 degradation rate (Heilskov and Holmer 2001). The tanaidacea *K. schubartii* was
28 recorded in significant densities and with little temporal variability, particularly over
29 2013. This species occurs in high densities in sandy-muddy substrates, showing
30 preference for sediment containing higher percentages of silt and clay (Capitoli et al.
31 1978) since it digs U-shape tubes with 15 cm depth (Rosa-Filho and Bemvenuti 1998).
32 Several studies indicate a significant density for the species in this estuary ($> 20,000$
33 ind. m^{-2}), and the marked seasonal influence. Higher salinity and high water
34 temperatures affect the *K. schubartii* recruitment during summer and early autumn ,

1 which suggested a continuous reproduction of the species (Fonseca and D'Incao 2006).
2 As long as the population is not subject to salinities close to zero for extended periods -
3 which may cause gaps in their recruitment - the success of *K. schubartii* recruitment is
4 also determined by the size of the population prior to the reproductive process,
5 indicating a density-dependence for the species (Colling et al. 2007).

6 Generally, a decreasing trend in macrozoobenthic density was observed in 2013
7 when compared to 2012 samplings. Nevertheless, it cannot be observed relation
8 between abiotic parameters over the two years of sampling and the result obtained.
9 Some species have undergone major variations in density along the sampling
10 campaigns, as *P. pinnata*, *Amphitoe sp* and Ostracoda, that almost disappear in 2013
11 and *K. schubartii* and *S. mourei* that increased the density in the same year. In this way,
12 many factors can affect its variability. These fluctuations are influenced by the
13 conditions which favor growth and development of pelagic larvae (Kastoro et al. 1989).
14 Senales et al. (2007) report that climatic events (*El Niño*) and the effects of competition
15 and predation by other species may have caused fail in *P. pinnata* recruiting in Chile.

16 In this study the shrimp farm effluent not contributed to a reduced species
17 richness, which could indicate that there was no accumulation of organic matter enough
18 to cause a restructuring of macrozoobenthic community, different of the results reported
19 by Canary et al. (2009) and Rodríguez-Gallego et al. (2008) that found changes in
20 macrobenthic structure in environments next to shrimp farms. In general, no significant
21 differences over the sampling sites show an absence of disturbances in macrozoobenthic
22 community by the effluents release, since there was no clear distribution change of
23 macrofaunal assembly. This study showed that the effects of effluent discharge on some
24 attributes of macrozoobenthic community (density and richness) were less significant
25 than the natural environmental variation, since the estuarine invertebrate communities in
26 temperate regions have significant seasonal variations.

27

28 Acknowledgments

29 The authors are grateful to Denise Aernoudts (shrimp farm owner), The Ministry of
30 Fisheries and Aquaculture (MPA), Brazilian Council of Research (CNPq) and
31 Coordination for the Improvement of Higher Level or Education Personnel (CAPES).
32 L.H. Poersch received productivity research fellowship from CNPq.

33

1 RESUMO

2

3 A produção em cativeiro do camarão branco do Pacífico *Litopenaeus vannamei* vem se
4 consolidando cada vez mais no entorno do estuário da Lagoa dos Patos, região sul do
5 Brasil. Nesse estudo, foram avaliados os efeitos espaço-temporais dos efluentes de uma
6 fazenda de cultivo de camarões sobre o macrozoobentos ($> 500\mu\text{m}$) ao longo de dois
7 ciclos produtivos (2012 e 2013). As amostragens ocorreram em sete diferentes pontos
8 do ambiente receptor, sendo dois pontos controle e cinco pontos distribuídos em frente a
9 desembocadura do canal de lançamento dos efluentes. As coletas ocorreram em
10 diferentes escalas temporais. Sete grupos foram observados (Polychaeta, Tanaidacea,
11 Isopoda, Gastropoda, Bivalvia, Malacostraca e Ostracoda) ao longo das duas campanhas
12 amostrais. Espacialmente, a densidade e a riqueza de espécies tiveram uma
13 variabilidade pouco expressiva em ambas as campanhas amostrais. Já ao longo do
14 tempo, na campanha de 2012 observou-se um aumento desses índices no inverno
15 comparativamente ao verão, oposto do que foi constatado em 2013. Os resultados
16 sugerem que o macrobentos não sofreu influência do lançamento dos efluentes e que as
17 diferenças estatísticas ($P < 0,05$) observadas foram decorrentes das oscilações naturais
18 dos parâmetros abióticos que ocorrem em sistemas estuarinos.

19

20 *Palavras-chave:* Densidade, Riqueza de espécies, Sedimento, *Litopenaeus vannamei*.

21

22 REFERENCES

23

- 24 Aguado-giménez F, Piedecausa MA, Carrasco C, Gutiérrez JM, Aliaga V and García-
25 garcía B. 2011. Do benthic biofilters contribute to sustainability and restoration of
26 the benthic environment impacted by offshore cage finfish aquaculture? Mar Pollut
27 Bull 62: 1714–1724.
- 28 Albertelli G, Covazzi-Harrigue A, Danovaro R, Fabiano M, Fraschetti S and Pusceddu
29 A. 1999. Differential responses of bacteria, meiofauna and macrofauna in a shelf
30 area (Ligurian Sea, NW Mediterranean): role of food availability. J Sea Res 42:
31 11–26.

- 1 Alongi DM, Lindsay FT and Trott A. 1999. Rates and pathways of benthic
2 mineralization in extensive shrimp ponds of the Mekong delta, Vietnam.
3 Aquaculture 175: 269-292.
- 4 Ansari ZA, Ingole BS and Parulekar AH. 1986. Effect of high organic enrichment of
5 benthic polychaete population in an estuary. Mar Pollut Bull 17: 361- 365.
- 6 Bartoli M, Nizzoli D, Viaroli P, Turolla E, Castaldelli G, Fano EA and Rossi R. 2001.
7 Impact of *Tapes philippinarum* farming on nutrient dynamics and benthic
8 respiration in the Sacca di Goro. Hydrobiologia 455: 203–212.
- 9 Bemvenuti CE, Capitoli RR and Gianuca NM. 1978. Estudos de ecologia bentônica na
10 região estuarial da Lagoa dos Patos. II. Distribuição quantitativa do macrobentos
11 infralitoral. Atlântica 3: 23-32.
- 12 Bemvenuti CE. 1987. Predation effects on a benthic community in estuarine soft
13 sediments. Atlantica 9: 33– 63.
- 14 Bemvenuti CE. 1988. Impacto da predação sobre *Heteromastus similis* Southern, 1921
15 e *Nephtys fluviatilis* Monro, 1937 (Annelida, Polychaeta), em fundos moles
16 estuarinos. Atlântica 10: 85-102.
- 17 Bemvenuti CE. 1997. Benthic Invertebrates. In: Seeliger U, Odebrecht C and Castello
18 JP. (Eds.), Subtropical convergence environments. The coast and sea in the
19 southwestern Atlantic. Berlin: Springer-Verlag, Berlin, Germany, p. 43-46.
- 20 Bemvenuti CE. 1998. In: Seeliger U, Odebrecht C and Castello JP. (Eds.), *Os*
21 *Ecossistemas Costeiro e Marinho do Extremo Sul do Brasil*. Rio Grande:
22 Ecoscientia, Rio Grande, Brazil, p. 46-51.
- 23 Bemvenuti CE and Colling LA. 2010. As comunidades de macroinvertebrados
24 bentônicos. In.: Seeliger U and Odebrecht C. (Eds), O Estuário da Lagoa dos
25 Patos. Um Século de Transformações. 1^a Edição. Universidade Federal do Rio
26 Grande (FURG). p. 101-114.
- 27 Burford MA, Costanzo SD, Dennison WC, Jackson CJ, Jones AB, Mckinnon AD,
28 Preston NP and Trott LA. 2003. A synthesis of dominant ecological processes in
29 intensive shrimp ponds and adjacent coastal environments in NE Australia. Mar
30 Pollut Bull 46: 1456-1469.
- 31 Canary AC, Poersch L and Wasielesky W. 2009. Impacto dos efluentes de cultivo semi-
32 intensivo de camarão sobre a fauna bentônica no sul do Brasil. Acta Scientiarum
33 31: 345-353.

- 1 Capitoli RR, Bemvenuti CE and Gianuca NM. 1978. Estudos de ecologia bentônica na
2 região estuarial da Lagoa dos Patos, I. Comunidades bentônicas. Atlântica 3: 5-
3 21.
- 4 Carroll ML, Cochrane S, Fieler R, Velvin R and White P. 2003. Organic enrichment of
5 sediments from salmon farming in Norway : environmental factors, management
6 practices, and monitoring techniques. Comp Gen Pharmacol 226: 165 – 180.
- 7 Carvalho S, Falcão M, Cúrdia J, Moura A, Serpa D, Gaspar MB, Dinis MT, Pousão
8 Ferreira P and Fonseca LC. 2009. Benthic dynamics within a land-based semi-
9 intensive aquaculture fish farm: the importance of settlement ponds. Aquacult Int
10 17: 571–587.
- 11 Castello JP and Moller OO. 1978. On the relationship between rainfall and shrimp
12 production in the Estuary of the Patos Lagoon (Rio Grande do Sul, Brazil).
13 Atlântica 3: 67– 74.
- 14 Colling LA, Bemvenuti CE and Gandra MS. 2007. Seasonal variability on the structure
15 of sublittoral macrozoobenthic association in the Patos Lagoon estuary, southern
16 Brazil. Iher Ser Zool 97: 257-263.
- 17 Crawford CM, Macleod CKA and Mitchell IM. 2003. Effects of shellfish farming on
18 the benthic environment. Sites Aquaculture 224: 117 – 140.
- 19 Dauvin JC. 2007. Paradox of estuarine water quality: benthic indicators and indices,
20 consensus or debate for the future. Mar Pol Bull 55: 271-281.
- 21 D'Incao F and Reis EG. 2002. Community-based management and technical advice in
22 Patos Lagoon estuary (Brazil). Ocean Coast Manag 45: 531–539.
- 23 Ellison AM. 2008. Managing mangroves with benthic biodiversity in mind: Moving
24 beyond roving banditry. J Sea Res 59: 2–15.
- 25 Fonseca DN and D'Incao F. 2006. Mortality of *Kalliapseudes schubartii* in unvegetated
26 soft bottoms of the estuarine region of the Lagoa dos Patos. Braz Arch Biol
27 Technol 49: 257-261.
- 28 Forchino A, Borja A, Brambilla F, Rodríguez JG, Muxika I, Terova G and Saroglia M.
29 2011. Evaluating the influence of off-shore cage aquaculture on the benthic
30 ecosystem in Alghero Bay (Sardinia, Italy) using AMBI and M-AMBI. Ecol Indic
31 11(5): 1112–1122.
- 32 Foreman K, Valiela I and Sarda R., 1995. Control of benthic marine food webs. Sci Mar
33 59: 119–128.

- 1 Gray JS and Christie H. 1983. Predicting long-term changes in marine benthic
2 communities. Mar Ecol Prog Ser 13: 87-94.
- 3 Gray JS and Elliott M. 2009. Ecology of Marine Sediments. From Science to
4 Management. New York: Oxford University Press, 2nd ed., 225p.
- 5 Heilskov AC and Holmer M. 2001. Effects of benthic fauna on organic matter
6 mineralization in fish-farm sediments: importance of size and abundance. ICES J
7 Mar Sci 58: 427–434.
- 8 Heip C. 1995. Eutrophication and zoobenthos dynamics. Ophelia 41: 113 –136.
- 9 Hutchings P. 1998. Biodiversity and functioning of polychaetes in benthic sediments.
10 Biodivers Conserv 7: 1133–1145.
- 11 Islam MS. 2005. Nitrogen and phosphorus budget in coastal and marine cage
12 aquaculture and impacts of effluent loading on ecosystem: review and analysis
13 towards model development. Mar Pollut Bull 50(1): 48–61.
- 14 Jackson C, Preston N and Thompson PJ. 2004. Intake and discharge nutrient loads at
15 three intensive shrimp farms. Aquacult Res 35: 1053-1061.
- 16 Johannssen P, Botnen P Aand Tvedten O. 1994. Macrofauna: before, during and after
17 a fish farm. Aquac Res 25: 55-66.
- 18 Kastoro W, Aswandy I, Al Hakim I, De Wilde PAWJ and Everaarts JM. 1989. Soft-
19 bottom benthic community in the estuarine waters of East Java. Neth J Sea Res
20 23(4): 463–472.
- 21 Lorenzen S, Prein M and Valentin C. 1987. Mass aggregations of the free-living marine
22 nematode *Pontonema vulgare* Oncholaimi- dae in organically polluted fjords. Mar
23 Ecol Prog Ser 37: 27-34.
- 24 Mouillot D, Laune J, Tomasini JA, Aliaume C, Brehmer P, Dutrieux E and Do Chi T.
25 2005. Assessment of coastal lagoon quality with taxonomic diversity indices of
26 fish, zoobenthos and macrophyte communities. Hydrobiologia 550: 121–130.
- 27 Murugesan P, Ajithkumar TT, Khan SA and Balasubramanian T. 2009. Use of benthic
28 biodiversity for assessing the impact of shrimp farming on environment. J Environ
29 Biol 30: 865–870.
- 30 Olivier M, Desrosiers G, Caron A, Retiere C and Caillou A. 1995. Behavioral responses
31 of the polychaetes *Nereis diversicolor* (O.F. Mueller) and *Nereis virens* (Sars) to
32 food stimuli: use of particulate organic matter (algae and halophytes). Can J Zool
33 73: 2307-2317.

- 1 Omena PO and Amaral ACZ. 2001. Morphometric study of the nereididae *Laeonereis*
2 *acuta* (Annelida: Polychaeta). J Mar Biol Assoc UK 81: 423–426.
- 3 Pagliosa PR and Barbosa FAR. 2006. Assessing the environment–benthic fauna
4 coupling in protected and urban areas of southern Brazil. Biol Conserv 129: 408–
5 417.
- 6 Pearson TH and Rosenberg R. 1978. Macrobenthic succession in relation to organic
7 enrichment and pollution of the marine environment. Oceanogr Mar Biol A Rev
8 16: 229–311.
- 9 Pocklington P, Scott DB and Schafer CT. 1994. Polychate response to different
10 aquaculture activities. Memoires du Museum National d'Histoire Naturelle, 162:
11 511-520.
- 12 Reish DJ. 1980. Use of polychaetous annelids as test organism for marine bioassay
13 experiments. In: Buikema AL Jr, Cairns, J.C Jr (eds). Aquatic Invertebrate
14 Bioassay). ASTM 715, Philadelphia, 140-145p.
- 15 Rodríguez-Gallego L, Meerhoff E, Poersch L, Aubriot L, Fagetti C, Vitancurt J and
16 Conde D. 2008. Establishing limits to aquaculture in a protected coastal lagoon :
17 Impact of *Farfantepenaeus paulensis* pens on water quality, sediment and benthic
18 biota. Aquaculture 277: 30–38.
- 19 Rosa-Filho JS and Bemvenuti CE. 1998. Caracterización de las comunidades
20 macrobentónicas de fondos blandos en regiones estuarinas de Rio Grande do Sul
21 (Brasil). Thalassas 14: 43-56.
- 22 Rosa LC and Bemvenuti CE. 2006. Temporal variability of the estuarine macrofauna of
23 the Patos Lagoon, Brazil. Rev Biol Mar Oceanogr 41: 1–9.
- 24 Sellanes J, Quiroga E, Neira C and Gutiérrez D. 2007. Changes of macrobenthos
25 composition under different ENSO cycle conditions on the continental shelf off
26 central Chile. Cont Shelf Res 27: 1002–1016.
- 27 Suguio K. 1973. Introdução à sedimentologia. São Paulo: E. Blucher, 317 p.
- 28 Tomassetti P and Porrello S. 2005. Polychaetes as indicators of marine fish farm
29 organic enrichment. Aquac Int 13: 109-128.
- 30 Underwood AJ. 1994. On beyond BACI: sampling designs that might reliably detect
31 environmental disturbances. Ecol Appl 4: 3–15.
- 32

1 **CAPÍTULO 3**

2

3 **IMPACTOS DOS EFLUENTES DA CARCINOCULTURA SOBRE O**
4 **SEDIMENTO**

5

6 Artigo submetido e aceito para publicação no periódico *Food and Nutrition Sciences*
7 (ISSN: 09676120, 1573143X).

8

1 Impact of Shrimp Farming Effluent on Sediment
2
3 Impact of Shrimp Farming Effluent: TOC, TN, Cu and Zn Levels in the Sediment

4
5
6
7
8 **William Bauer¹, Monica Wallner Kersanach², Wilson Wasielesky¹ and Luis**
9 **Henrique da Silva Poersch^{1*}**

10
11
12 ¹Marine Station of Aquaculture, Institute of Oceanography, Federal University of Rio
13 Grande, Brazil
14 Rua do Hotel, 02, CEP 96.210-030, Rio Grande, Brazil

15 ²Hidrochemical Laboratory, Institute of Oceanography, Federal University of Rio
16 Grande, Brazil
17 Avenida Itália, km 8, CEP 96.201-900, Rio Grande, Brazil

18 *Corresponding author: lpoersch@mikrus.com.br
19 Tel.: 55(53)3236-8132

1 Abstract

2 The unconsumed food in shrimp farm systems is responsible for increasing the organic
3 compounds in sediment and it is a potential source of trace metals. The aim of this study
4 was to evaluate the total organic carbon (TOC), total nitrogen (TN), copper (Cu) and
5 zinc (Zn) concentrations in sediments from control sites, shrimp pond, sedimentation
6 basin and receiving ecosystem effluents of a semi-intensive *Litopenaeus vannamei* farm
7 in southern Brazil. The sediment samples were taken before the effluent discharge (BD),
8 1-day post-discharge (1PD), 10 days post-discharge (10PD) and 30 days post-discharge
9 (30PD). Shrimps were also collected to analyze Cu and Zn concentrations in the tissue.
10 The TOC concentration ranged from 0.12 to 0.67% and the TN concentration was
11 <0.07% in all samples. Labile Cu and Zn concentrations ranged from 0.12 to 1.27 $\mu\text{g g}^{-1}$
12 and 0.51 to 3.07 $\mu\text{g g}^{-1}$, respectively, while the more strongly adsorbed fraction Cu
13 ranged from 0.3 to 2.65 $\mu\text{g g}^{-1}$ and Zn 30.44 to 121.4 $\mu\text{g g}^{-1}$. Some significant differences
14 ($p<0.05$) were observed among the sites, but not related to the effluent discharge.
15 Pearson correlation analysis showed no relationship between the effluent discharge and
16 increase in TOC, TN, Cu or Zn values in the sediment, except in 1PD. The Cu and Zn
17 concentrations in shrimps' tissue were $6.63\pm0.2\mu\text{g g}^{-1}$ and $19.76\pm0.2\mu\text{g g}^{-1}$ in pond 1,
18 and $7.6\pm0.51\mu\text{g g}^{-1}$ and $19.13\pm0.32\mu\text{g g}^{-1}$ in pond 2. All parameters were within the safe
19 levels stipulated in the Brazilian legislation, showing that this shrimp production did not
20 bring adverse effects to the environment.

21

22 Keywords: copper, *Litopenaeus vannamei*, total nitrogen, total organic carbon, zinc.

23

24

25

26

27

28

29

30

31

32

33

1 Abbreviations

2

3 TOC – Total organic carbon

4 TN – Total nitrogen

5 Cu – Cooper

6 Zn – Zinc

7 BD – Before discharge

8 1PD – 1-day post-discharge

9 10PD – 10 days post-discharge

10 30PD – 30 days post-discharge

11

1 Introduction

2

3 The aquaculture in coastal areas is always inquired about the possible
4 environmental impacts of the activity (Biao et al. 2004). The uncontrolled growth of
5 shrimp farming has led to negative environmental impacts in many countries (Páez-
6 Osuna 2001a; Páez-Osuna 2001b), which increased the concern on the activity
7 expansion. Culture systems become more intensive as new technology packages are
8 available; intensive systems usually have greater concentrations of nutrients and organic
9 matter in the culture environment (Lemonnier and Faninoz 2006), thus increase the need
10 of water exchanges. Therefore, the effluents from these systems are loaded with
11 nitrogen, carbon, phosphorus and suspended solids (Paez-Osuna et al. 1997; Biao et al.
12 2004).

13 According to Boyd & Teicher-Coding (1995), less than fifty percent of carbon
14 and nitrogen from food is converted into shrimps biomass. Briggs & Funge-Smith
15 (1998) state that approximately 18-27% of nitrogen and 6-11% of carbon from the feeds
16 is assimilated by shrimps, and the remainder is available for plankton, volatilizes or
17 stays entrapped in the sediment. The organic and inorganic dissolved nutrients have an
18 indirect ecological impact by increasing primary phytoplankton production, which
19 decease and increase the organic matter sedimentation (Olsen et al. 2008). Thus, the
20 organic matter in aquaculture ponds bottom derives from phytoplankton, unconsumed
21 feed and shrimps excretion (Funge-Smith and Briggs 1998; Steeby et al. 2004). This
22 accumulation can cause biological and chemical impacts to pond environment (Suplee
23 and Cotner 1996) and to receiving waters environment, since such large amounts of
24 nutrients promote microbial growth by the availability of organic matter. This increase
25 of organic matter can cause eutrophication of the ecosystem. Eutrophic systems usually
26 have particulate organic carbon (Pelletier et al. 2011), which is a good indicator of
27 enriched sediment (Hyland et al. 2005). Organic nitrogen also plays an important role as
28 a source of nutrients (Fütterer 2000), and thus is an important element to be evaluated.

29 Sediment can trap metals in the aquatic system serving as a good pollution
30 indicator, allowing a consistent evaluation of spatial and temporal contamination
31 (Solomons & Förstner 1984; Buchman 1989). Aquaculture is often reported as a
32 potential source of trace metals, which are present as natural components in feed,
33 fertilizer or as impurities of pesticides (Tacon & Forster 2003). The copper (Cu) is a
34 trace element present in shrimp a diets essential to synthesize hemocyanin in the

hemolymph (Cuzon 2004) and zinc (Zn) is a cofactor in many enzyme systems (Davis et al. 2002). The quantification of these trace metals has already been evaluated in salmon farming, serving as indicators of aquaculture wastewater (Chou et al. 2002). The sediment, when revolved, releases the labile metal fraction to the water column, which can have toxic effects on organisms (Wallner-Kersanach et al. 2009) by inducing changes in physicochemical conditions, especially changes in pH and redox-potential (Cappuyns & Swennwn 2005). It is necessary to consider that shrimp farming effluents can contribute to trace metals to the adjacent coastal environment, depending mainly on the amount of feed used during the production cycle (Lacerda et al. 2006).

Trace metals can be also quantified in aquatic organisms' tissues, since the excess of trace metals in unbalanced diets for shrimp *L. vannamei* promotes the accumulation of these compounds in their tissues (Yang and Wu 2011) and the analysis of trace metals in aquatic organisms can provide important information on the degree of environmental contamination and the potential impact of this food consumption (Ip et al. 2005).

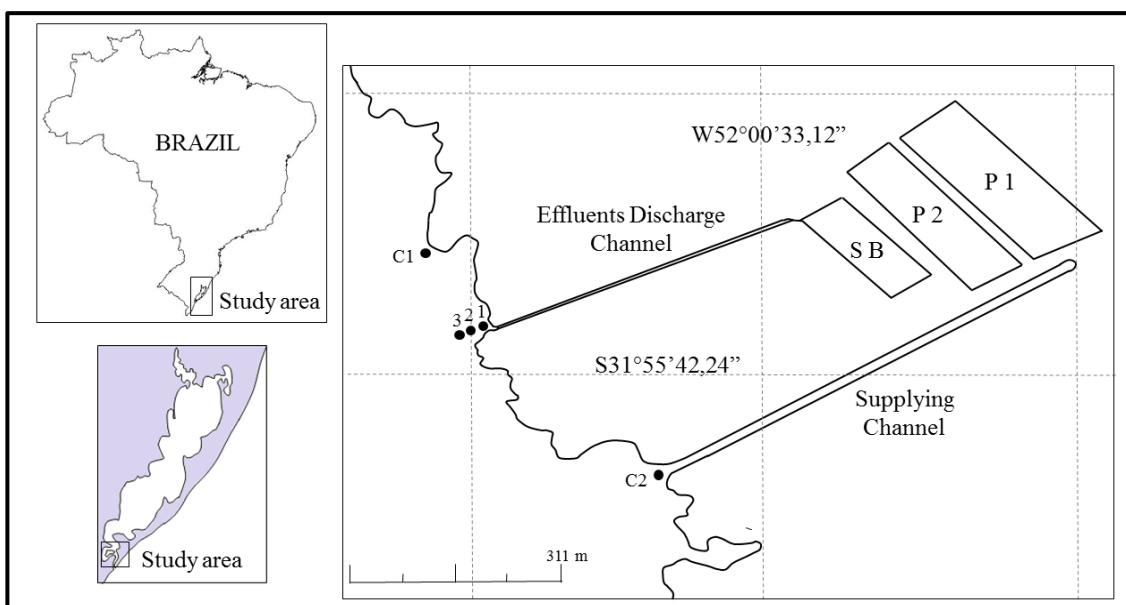
Literature concerning the TOC, TN and trace metals in the shrimp farms sediments are scarce. Therefore, this study evaluated the total organic carbon (TOC), total nitrogen (TN), Cu and Zn in the pond sediment, sedimentation basin and the receiving waters. Additionally, we determined the concentrations of both metals in the tissue of farmed *L. vannamei*.

Materials and Methods

Study area and sampling

This study was conducted in a *L. vannamei* shrimp farm located in the coastal area of Patos Lagoon estuary (Rio Grande do Sul, Brazil - 31°56'04S, 52°00'11W). The area does not suffer anthropogenic effect and the shrimp farm have one cycle per year, where after each cycle the ponds are drained and dried. The culture period began in December 2011 and ended in May 2012 when the shrimps were harvested. Outflowing waters are driven into a sedimentation basin before flowing into the receiving body water. The stocking density was 12/m² shrimp in two ponds, with a total area of 3.75 ha. The facility works in a semi-intensive farming system and there was no effluent discharge during the culture period; water was only added in order to compensate the evaporation and soil infiltration.

1 Sediment samples were collected between February and May 2012. For the TOC
2 and TN analysis, seven sampling sites were chosen: two control sites (C1 and C2) in the
3 estuary, shrimp pond (P1), sedimentation basin (SB) and three sites across the effluents
4 discharge channel (in front of the channel - 1, 20 meters from the channel - 2 and 30 m
5 from the channel - 3) (Figure 1). For the analysis of Cu and Zn in sediment the sampling
6 sites were the control site (C1), shrimp pond (P1), sedimentation basin (SB) and a site in
7 front of the effluent outflow channel (1) (Figure 1). Temporal samplings at these sites
8 for analysis were taken according to effluents discharge, as follows: sample before the
9 discharge (BD), 1-day post-discharge (1PD), 10 days post-discharge (10PD) and 30
10 days post-discharge (30PD).



11
12 Figure 1: Sampling sites in the shrimp farming. C1 = Control 1, C2 = Control 2, 1 = Effluents
13 discharge channel, 2 = 20m from the channel, 3 = 30m from the channel, P1 = Pond 1, P2 =
14 Pond 2 e SB = Sedimentation basin.
15

16 The glassware used in this study was cleaned and stored according to
17 methodology described by Baumgarten et al (2010). At each sampling site, three
18 sediment samples were collected with a 5-cm-diameter PVC pipe. Sub-samples were
19 separated with plastic spatula and stored in plastic bags for Cu and Zn determination.
20 With a stainless steel spatula, sediment samples of surface layer were collected for TOC
21 and TN analysis from each site and stored in 50 ml glass vials. In the same sites
22 sediment samples were also collected to determine the proportions of sand, silt and clay
23 by sieving (> 0.062 mm diameter) and pipetting (<0.062 mm diameter) as described by

1 Suguiio (1973). At the end of the culture period, shrimp specimens were collected from
2 two shrimp ponds (P1 and P2) for Cu and Zn tissue analysis.

3

4 *Analysis of Cu and Zn in the sediment and shrimps muscle tissues*

5

6 Two methods of extraction of Cu and Zn of sediment were performed: low
7 extraction, which provides the labile fraction (potentially bioavailable) and semi-strong
8 extraction (more strongly adsorbed). All the material used for trace metals analysis was
9 previously washed with solution of 20% (v / v) nitric acid. The semi-strong extraction
10 values were compared to Brazilian legislation about these metals.

11 For the Cu and Zn labile extraction ($n = 2$), 1 g of sediment samples was
12 previously macerated and dried in an oven at 60 °C and then re-weighed. The samples
13 were digested in 0.1M hydrochloric acid (v / v) Suprapuro® (Merck, Germany) for 24
14 hours under continuous stirring, in a solid / liquid ratio of 1:20 (g / ml). The leaching
15 products rest for 5 hours to sedimentation, and then they were filtered through Whatman
16 # 44 filter paper in a slow rate at ambient pressure. The extracts obtained were
17 transferred to 25ml volumetric flasks and filled with 0.1M HCl solution (Li et al. 2009,
18 modified).

19 The semi-strong extraction analysis of Cu and Zn ($n = 2$) was conducted by
20 weighing 1g of dry sediment and digesting it with concentrated nitric acid and hydrogen
21 peroxide (Suprapuro®, Merck, Germany) at 4:1 volume proportion. The digestion
22 process occurred in a microwave (CEM Model X-Press Model Mars) for 45 minutes at
23 180 °C and 1600W. Subsequently, the samples were filled with 50 mL of Milli-Q®
24 water.

25 Trace metals determination in shrimps' tissues was carried out with pools of 30
26 specimens from each shrimp pond and the analysis was performed in triplicate. In the
27 procedure, 5 mL of concentrated nitric acid Suprapuro® (Merck, Germany) was added
28 in 0.5g of dry muscle previously weighed, followed by microwave digestion as in the
29 sediment samples. The samples were then filled with 25 mL of Milli-Q® water. The Cu
30 and Zn analysis in sediment and shrimp tissue was performed by Inductively Coupled
31 Plasma Optical Emission Spectrometry (2100DV Model, ICP-OES).

32

33 *Statistical Analysis*

34

1 All data were submitted to the analysis of variance (ANOVA) followed by a
2 Tukey test (mean comparison). Possible relations between the TOC, TN, copper and
3 zinc values (for both extraction methods) was determined using the Pearson correlation
4 analysis. The significance level used for all tests was 5% (Sokal and Rohlf 1995).

5

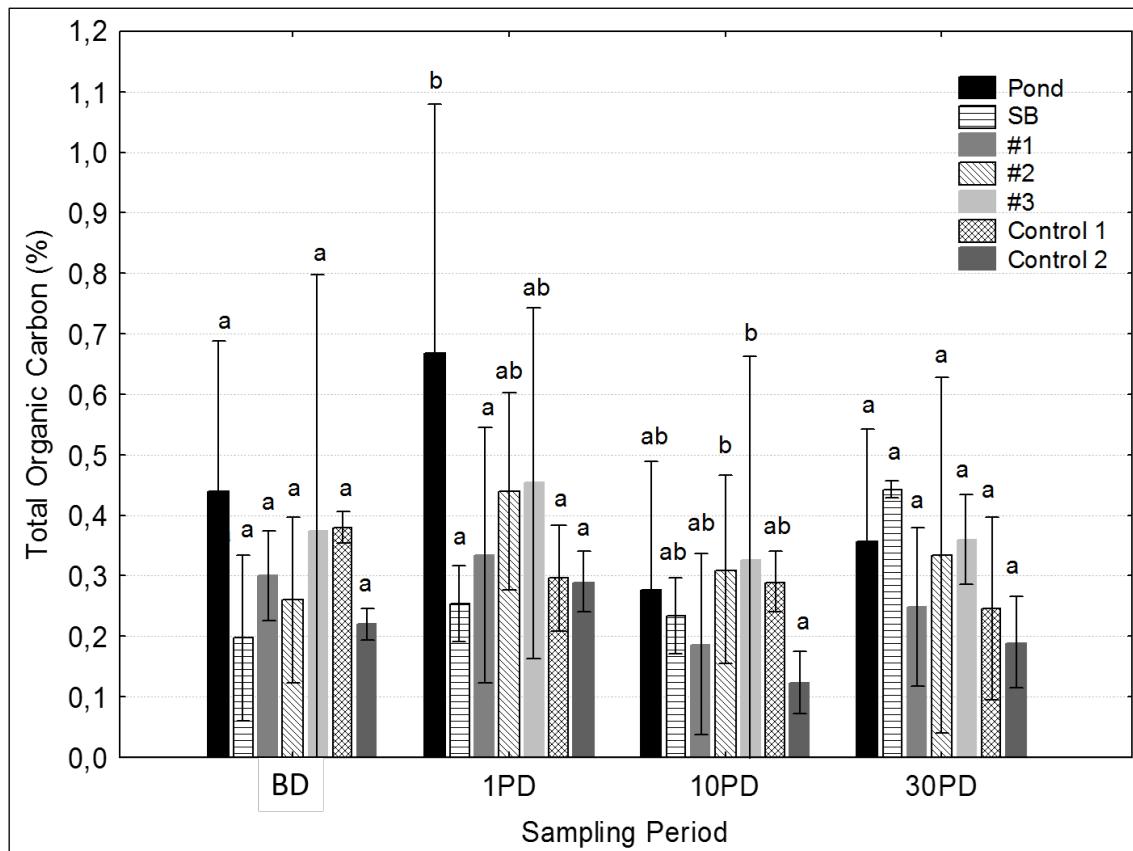
6 Results

7

8 The sediment was mainly composed by fine sand (2.5 Phi) in all sampling sites
9 (43.64 to 48.28%). The percentage of TOC showed variations among sampling sites
10 (Figure 1). In the BD and 30PD samplings, no significant differences were observed (P
11 < 0.05). The shrimp pond in BD and 1PD had the highest mean values of TOC. The
12 shrimp pond in 1PD differed significantly ($P < 0.05$) from the sedimentation basin and
13 the sites 1 (in front of the effluents discharge channel), Control 1 and Control 2. In the
14 10PD, the C2 site differed significantly ($P < 0.05$) from 2 and 3 sites. TN concentrations
15 were below the detection limit of the equipment (< 0.07 %) for all sampling sites
16 analyzed.

17

18

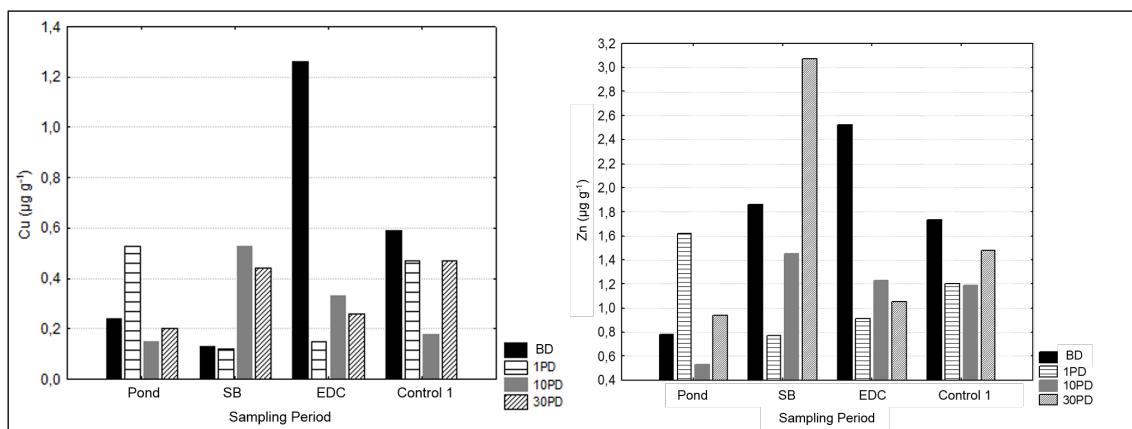


19
20 Figure 1: Spatio-temporal TOC percent variations in sediment (mean \pm SD, n = 3). Different
21 letters denote significantly differences ($P < 0.05$) through the different periods of effluents

1 discharge: sample pre-discharge (BD), 1-day post-discharge (1PD), 10 days post-discharge
2 (10PD) and 30 days post-discharge (30PD). SB = Sedimentation Basin
3

4 The mean concentration of Cu and Zn in the labile fraction ranged from 0.12 to
5 $1.27\mu\text{g g}^{-1}$ and 0.52 to $3.07\mu\text{g g}^{-1}$, respectively (Figure 2). The Pearson's correlation
6 between TOC concentration and the metals indicated a significant and moderate
7 correlation ($P < 0.05$; $r = 0.68$) with the labile fraction of Cu and significant and strong
8 correlation ($P < 0.05$; $r = 0.90$) with the labile fraction of Zn in the sediment sample DP.
9 The highest value of Cu ($1.27 \mu\text{g / g}$) was found at the effluent outflow channel in BD
10 sampling. It is noteworthy that no effluent was released at this date. The sedimentation
11 basin showed the highest Zn value ($3.07 \mu\text{g g}^{-1}$) in 30PD.

12



13
14 Figure 2: Mean concentration (n=2) of Cu e Zn ($\mu\text{g/g}$) in sediment through weak acid extraction
15 (bioavailable) in different sampling sites. SB = Sedimentation Basin; EDC = Effluents
16 Discharge Channel. *Figure in different scales.
17

18 The semi-strong extraction of Cu and Zn in the sediment showed variations
19 between 0.3 to $2.65\mu\text{g g}^{-1}$ for Cu and 30.44 to $121.4\mu\text{g g}^{-1}$ for Zn (Figure 3). Unlike the
20 labile fraction, when Pearson's correlation was applied only Cu indicated a strong
21 correlation ($P < 0.05$; $r = 0.70$) with TOC in 1PD.

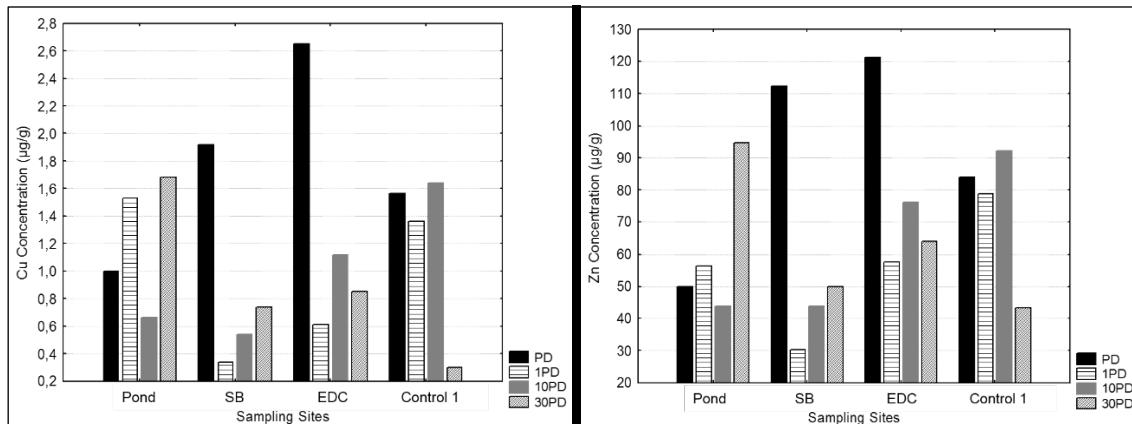


Figure 3: Mean concentration (n=2) of Cu e Zn ($\mu\text{g/g}$) in sediment through semi-strong extraction in different sampling sites. SB = Sedimentation Basin; EDC = Effluents Discharge Channel. *Figure in different scales.

The mean concentration of Cu and Zn in the tissue of the shrimp were $6.63 \pm 0.2\mu\text{g g}^{-1}$ of Cu and $19.76 \pm 0.2\mu\text{g g}^{-1}$ of Zn in pond 1 and $7.6 \pm 0.51\mu\text{g g}^{-1}$ of Cu and $19.13 \pm 0.32\mu\text{g g}^{-1}$ of Zn in pond 2.

Discussion

The TOC in this study was low and had little variation (0.12 to 0.67%). Several factors may have contributed to this result, and the low stocking density of shrimp during the culture period is an important fact, as mentioned in another study for Vinatea et al (2006). Organic carbon is originated from the dead phytoplankton, organic fertilizer, unconsumed feed and animal faeces with soil particles (Olsen et al. 2008). The low stocking densities of shrimp prevent that large nutrient inputs are inserted through the feed, and the appropriate management of ponds' water quality prevents algal blooms. Burford & Williams (2001) report that phytoplankton blooms have deleterious effects on water quality and sediment.

The grain size of the sediment is another determining factor in organic material content of estuarine sediments (Pelletier et al. 2011). The predominantly sandy and fine content found in this study, in addition with constant artificial water aeration during the culture period, may have improve the decomposition of organic matter, avoiding the carbon accumulation in sediment. Sutherland et al (2007) found a strong correlation between the porosity of the sediment and organic content, where sandy sediments were characterized by low organic content. In BD and 30PD sampling, no significant differences were observed of TOC between the sampling sites, unlike occurred in 1PD

1 and 10PD. However, these differences did not appear to be related to the effluent
2 discharge effects.

3 The shrimp pond presented the highest percentage of TOC in BD and 1PD
4 sampling, and these values decreased after shrimp harvest (10PD and 30PD sampling).
5 This is due to the pond drainage and sun-drying, which increases the soil aeration and
6 accelerate the organic matter decomposition (Ayub et al. 1993; Boyd 1995). Boyd et al
7 (2010) evaluated 233 commercial aquaculture ponds and found TOC concentrations in
8 sediment ranging from 1.08 to 7.01%. The authors report that there was not a clear
9 relationship between TOC concentration and the species or stocking density. Another
10 study evaluating ponds sediment in six *L. vannamei* shrimp farms in southern Brazil
11 identified average concentrations of TOC ranging between 0.9 and 2.15% (Vinatea et al.
12 2006). Smith (1996) also reported organic carbon values between 0.96% and 2.49% in a
13 *Penaeus monodon* farm. In Mirim Lagoon, a lake connected to Patos Lagoon estuary,
14 showed TOC concentrations in sediment ranging from 0.21 to 2.4% (Santos et al. 2003).

15 The low concentrations of TN found in the sediment (< 0.07 %) may indicate
16 that there was no excess of bacterial activity to decompose organic matter. Studies have
17 shown that the microbial community (Burford and Williams 2001), phytoplankton
18 (Hargreaves 1998; Burford and Glibert 1999) and the constant aeration throughout the
19 culture period play an important role in the removal of nitrogenous compounds in
20 culture ponds (Sanares et al. 1986). However, several studies have shown that the
21 effluent from shrimp farming can contribute to an increase of nitrogen in the sediment.
22 Anh et al (2010) reported 0.07% to 0.17% of NOT in the sediment of *P. monodon*
23 culture ponds; Lemonnier and Faninoz (2006) found higher levels (0.15 to 0.22%) of
24 TN in semi-intensive shrimp culture ponds in New Caledonia. Teichert-Codding and
25 Boyd (1995) also reported that nitrogen may be converted into gaseous form and lost to
26 the atmosphere. The volatilization of ammonia in semi-intensive shrimp farming in
27 Mexico was estimated at 27.4% (Páez-Osuna et al. 1997).

28 This study showed low values of labile (potentially bioavailable) Cu (0.12 –
29 1.27 $\mu\text{g g}^{-1}$) and Zn (0.51 - 3 $\mu\text{g g}^{-1}$) concentrations in sediment. Labile Cu and Zn
30 concentration increased at 1PD and were significantly ($P < 0.05$) correlated the higher
31 levels of organic carbon found in the sediment. The Pearson's correlation analysis
32 demonstrated higher levels of organic carbon when Cu and Zn were increased in the
33 labile extraction on 1PD. This may be related to the availability of non-consumed feed
34 and faeces shrimps, since this correlation was not found at 10 and 30 days post-

1 discharge. Organic matter has a strong affinity for Cu and Zn (Lin and Chen 1998), and
2 Frías-Espericueta et al (2006) found a positive correlation between organic matter level
3 in the sediment of shrimp farm and the Cu, Zn and Ni concentration. However, the
4 content of C and N was low despite the positive correlation, which explain the low
5 levels of Cu and Zn also found in this study. Even though the high concentration of Cu
6 and Zn have been found in the effluent outflow channel in BD, the sampling occurred
7 prior to effluent discharge. This was probably due to poor water circulation in effluent
8 discharge channel, once the concentrations of both elements decreased after culture
9 period.

10 The content of labile Cu and Zn concentrations in sediment of the present study
11 were low when compared to other studies (Table 1). An trace metal assessment in
12 sediment of several sites of Patos Lagoon estuary showed mean values of 4.5 – 19.2 μg
13 g^{-1} of Cu and 18.5 – 62.5 $\mu\text{g g}^{-1}$ of Zn, both in the labile fraction <0.63 μm (silt and
14 clay), in an area with low anthropogenic impact (Costa et al., unpublished data). The
15 dominance of fine sand and the short period of use of the ponds promote low
16 environmental disturbance, different of the estuarine area of the Patos Lagoon.

17 In the same way, when comparing the semi-strong acid extraction of Cu and Zn
18 (0.3-2.65 $\mu\text{g g}^{-1}$ from 30.44 to 121.4 $\mu\text{g g}^{-1}$, respectively), the values were in general
19 lower than in other studies, except for the Zn concentration (121.4 $\mu\text{g g}^{-1}$) found in the
20 effluent discharge channel (EDC) on pre-discharge sample (PD). Russel et al (2011)
21 evaluated the concentrations of this metals in the sediment next to marine fish farms in
22 Scotland, and found mean values of 35.8 and 89.1 $\mu\text{g g}^{-1}$ of Cu and Zn, respectively. A
23 study in a *Penaeus monodon* farm reports concentrations up to 45 $\mu\text{g g}^{-1}$ of Cu and up to
24 85 $\mu\text{g g}^{-1}$ of Zn in ponds sediment and concentrations below 2 $\mu\text{g g}^{-1}$ of Cu and 64 $\mu\text{g g}^{-1}$
25 of Zn in effluent sediment (Smith 1996). Moreover, another study evaluated the amount
26 of Cu and Zn in a shrimp farm and it was found a negative imbalance between the
27 quantities added via feed and fertilizer and the amounts found in soil, suggesting that
28 these metals may have been immobilized by conversion to organic form (Ritvo et al.
29 1998). Brazilian law does not have specific regulations for these trace metals
30 concentrations in aquaculture facilities, but there are established guidelines for the
31 management of dredged sediment in brackish waters, reports as safe values to the biota
32 34 $\mu\text{g g}^{-1}$ of Cu and 150 $\mu\text{g g}^{-1}$ of Zn (CONAMA 454/2012). Guidelines on soil quality is
33 also available, reporting safe values of 60 $\mu\text{g g}^{-1}$ of Cu and 300 $\mu\text{g g}^{-1}$ of Zn in soil

1 (CONAMA 420/2009). This shows that the results obtained in this work are in
2 accordance to the guidelines cited above.

3

4 Table 1: Trace metal concentrations ($\mu\text{g g}^{-1}$ dry weight, mean values) in the sediment and shrimp tissue of
5 this study area compared with different locations.

Locations	Fraction	Extraction	Cu	Zn	References
Patos Lagoon, Brazil ^a	Total	HCl	0.12-1.27	0.52-3.07	Present study
Patos Lagoon, Brazil ^d	<63 μm	HCl	4.5-19.2	18.5-62.5	Unpublished data
New Brunswick, Canada ^a	<63 μm	HNO ₃	21.2	71.5	Chou et al. 2002
Western Isles, Scotland ^a	<63 μm	HNO ₃	35.8	89.1	Russel et al. 2011
Cultured shrimp ^b	Pond 1	HNO ₃	6.63	19.76	Present study
	Pond 2	HNO ₃	7.6	19.13	
Arabian Sea, Pakistan ^c		HNO ₃	4.55	7.11	Jaffar et al. 1993
Pearl River estuary ^c		n.a.*	1.28	2.60	Wei et al. 2002
Zhanjiang Harbour, China ^b		HNO ₃	24.26	171.56	Wu and Yang 2011
Patos Lagoon, Brazil ^c		HNO ₃	0.45	3.77	Pinto et al. 2013

6 ^a adjacent to aquaculture farms; ^b shrimp farm cultured; ^c wild shrimp tissue; ^d Patos Lagoon without
7 pollutant source; * n.a. - not available.

8

9 The analysis of Cu and Zn concentrations in the muscle on a dry weight basis
10 resulted in low mean values of Cu ($7.1\mu\text{g g}^{-1}$) and Zn ($19.4 \mu\text{g g}^{-1}$). Table 1 shows Cu
11 and Zn values found in the present study compared to other studies. Trace metals in the
12 tissues present in the pink shrimp *Farfantepenaeus paulensis* from fisheries in Patos
13 Lagoon estuary were quantified and the values found were $12.9\mu\text{g g}^{-1}$ of Cu and $32.3\mu\text{g}$
14 g^{-1} of Zn (Pinto et al. 2013). Another study assessing the concentrations of Cu and Zn in
15 *L. vannamei* tissue from intensive shrimp farm in China resulted in mean levels of
16 $24.26\mu\text{g g}^{-1}$ of Cu and $171.56\mu\text{g g}^{-1}$ of Zn (Wu & Yang 2011). Some decapod
17 crustaceans are able to regulate bodily concentration of trace metals such as Cu and Zn,
18 while allowing the necessary requirements for their biochemical necessities and
19 detoxifying the excess (Rainbow 1988; Rainbow et al. 1999). In general, the values in
20 this study were low probably because the area of shrimp farming is an area of the
21 estuary without contribution of urban and industrial effluents. Other factors to consider
22 are the grain size of sediment in the ponds, type of feed and management used. The
23 concentrations found in this study are in accordance with national regulation.

24 These results suggest that the semi-intensive shrimp farm of *L. vannamei* was
25 performed with appropriate management which resulted in low values of TOC, TN, Cu
26 and Zn exported to the estuary sediment and low Cu and Zn concentrations in shrimps'
27 tissues. Therefore, it can be concluded that this shrimp *L. vannamei* culture period had
28 excellent handling conditions, and did not generate a significant organic or trace metals
29 load to sediment and shrimp.

1

2 Acknowledgments

3 The authors are grateful to The Ministry of Fisheries and Aquaculture (MPA), Brazilian
4 Council of Research (CNPq) and Coordination for the Improvement of Higher Level or
5 Education Personnel (CAPES). L.H. Poersch and W. Wasielesky received productivity
6 research fellowship from CNPq.

7

8 References

9

- 10 Anh PT, Kroeze C, Bush SR, Mol APJ (2010) Water pollution by intensive brackish
11 shrimp farming in south-east Vietnam: Causes and options for control. Agr
12 Water Manage 97:872–882.
- 13 Ayub M, Boyd CE, Teichert-Coddington D (1993) Effects of urea application, aeration,
14 and drying on total carbon concentrations in pond bottom soils. Prog Fish Cult
15 55:210–213.
- 16 Baumgarten MGZ, Wallner-Kersanach M, Niencheski LFH (2010) Manual de Análises
17 em Oceanografia Química. Rio Grande, Brasil
- 18 Biao X, Zhuhong D, Xiaorong W (2004) Impact of the intensive shrimp farming on the
19 water quality of the adjacent coastal creeks from Eastern China. Mar Pollut Bull
20 48:543–553.
- 21 Boyd CE, (1995a) Bottom Soils, Sediment and Pond Aquaculture. Chapman and Hall,
22 New York.
- 23 Boyd CE, Teichert-Codding D (1995b) Dry Matter, Ash, and Elemental Composition of
24 Pond-Cultured *Penaeus vannamei* and *P. stylirostris*. World Aquac Soc 26:88–
25 92.
- 26 Boyd CE, Wood CW, Chaney PL, Queiroz JF (2010) Role of aquaculture pond
27 sediments in sequestration of annual global carbon emissions. Environ Pollut
28 158:2537–2540.
- 29 Brasil. Resolução CONAMA – Conselho Nacional do Meio Ambiente. Resolução nº
30 420/2009. Dispõe sobre critérios e valores orientadores de qualidade do solo
31 quanto à presença de substâncias químicas e estabelece diretrizes para o
32 gerenciamento ambiental de áreas contaminadas por essas substâncias em

- 1 decorrência de atividades antrópicas. Diário Oficial da República Federativa do
2 Brasil, Brasília, DF 31 de dezembro de 2009.
- 3 Brasil. Resolução CONAMA – Conselho Nacional do Meio Ambiente. Resolução nº
4 454/2012. Estabelece as diretrizes gerais e os procedimentos referenciais para o
5 gerenciamento do material a ser dragado em águas sob jurisdição nacional.
6 Brasília, DF, 01 nov. 2012.
- 7 Burford MA, Glibert PM (1999) Short-term nitrogen uptake and regeneration in early
8 and late growth phase shrimp ponds. Aquac Res 30:215–227.
- 9 Burford MA, Williams KC (2001) The fate of nitrogenous waste from shrimp feeding.
10 Aquaculture 198:79-93.
- 11 Buchman MF (1989) A review and summary of trace contaminant data for coastal and
12 estuary Oregon: U.S. Department of Commerce, National Oceanic and
13 Atmospheric Administration, National Ocean Service, NOAA Technical
14 Memorandum NOS OMA 42, p 115.
- 15 Cappuyns V, Swennwn R (2005) Kinetics of element release during combined
16 oxidation and pHstat leaching of anoxic river sediments. App Geochem
17 20:1169-1179.
- 18 Chou CL, Haya K, Paon LA, Burridge L, Moffatt JD (2002) Aquaculture-related trace
19 metals in sediments and lobsters and relevance to environmental monitoring
20 program ratings for nearfield effects. Mar Pollut Bull 44:1259–1268.
- 21 Costa LDF, Wallner-Kersanach M (2013) Assessment of the labile fractions of copper
22 and zinc in marinas and port areas in Southern Brazil. Environ Monit Assess
23 185:6767–6781.
- 24 Cuzon G (2004) Nutrition of *Litopenaeus vannamei* reared in tanks or in ponds.
25 Aquaculture 235:513–551.
- 26 Davis DA, Arnold CR, McCallum I (2002) Nutritional value of feed peas (*Pisum*
27 *sativum*) in practical diet formulations for *Litopenaeus vannamei*. Aquac Nutr
28 8:87–94.
- 29 Frías-Espericueta MG, Osuna-López JI, Voltolina D, Correa-González EM, Armenta-
30 Monje MJ, López-López G, Izaguirre-Fierro G (2006) Metals in shrimp farm
31 sediments, Sinaloa, Northwest Mexico. Bull Environ Contam Toxicol 77:912–917.
- 32 Funge-Smith SJ, Briggs MRP (1998) Nutrient budgets in intensive shrimp ponds:
33 implications for sustainability. Aquaculture 164:117–133.

- 1 Fütterer DK (2000) The solid phase of marine sediments. In: Schulz HD, Zabel M (eds)
2 Marine Geochemistry. Berlin, Springer, p 26.
- 3 Jaffar M, Ashraf M, Wharf W (1993) Heavy metal concentrations in fish, shrimp,
4 seaweed, sediment and water from the Arabian Sea, Pakistan. Mar Pollut Bull
5 26:644-647.
- 6 Hargreaves JA (1998) Nitrogen biogeochemistry of aquaculture ponds. Aquaculture
7 166:181–212.
- 8 Hyland J, Balthis L, Karakassis I, Magni P, Shine J, Vestergaard O, Warwick R (2005)
9 Organic carbon of sediments as an indicator of stress in the marine benthos. Mar
10 Ecol Prog Ser 295:91–103.
- 11 Ip CCM, Li XD, Zhang G, Wong CSC, Zhang WL (2005) Heavy metal and Pb isotopic
12 compositions of aquatic organisms in the Pearl River Estuary, South China.
13 Environ Pollut 138:494–504.
- 14 Lacerda LD, Santos JÁ, Madrid RM (2006) Copper emission factors from intensive
15 shrimp aquaculture. Mar Pollut Bull 52:1823-1826.
- 16 Lemonnier H, Faninoz S (2006) Effect of water exchange on effluent and sediment
17 characteristics and on partial nitrogen budget in semi-intensive shrimp ponds in
18 New Caledonia. Aquac Res 37:938–948.
- 19 Li LY, Hall K, Yuan Y, Mattu G, McCallum D, Chen M (2009) Mobility and
20 bioavailability of trace metals in the water-sediment system of the highly
21 Urbanized Brunette Watershed. Water Air Soil Pollut 197:249-266.
- 22 Lin JG, Chen SY (1998) The relationship between adsorption of heavy metals and
23 organic matter in river sediments. Environ Int 24:345-352.
- 24 Mitra A, Mandal T, Bhattacharya DP (1999) Concentrations of heavy metals in *Penaeus*
25 spp. of brackish water wetland ecosystem of West Bengal, India. Indian J
26 Environ Ecoplan 2:97-106.
- 27 Olsen LM, Holmer M, Olsen Y (2008) Perspectives of nutrient emission from fish
28 aquaculture in coastal waters Literature review with evaluated state of
29 knowledge. The Fishery and Aquaculture Industry Research Fund, p 87.
- 30 Páez-Osuna F, Guerrero-Galvan SR, Ruiz-Fernandez AC, Espinoza-Angulo R (1997)
31 Fluxes and mass balances of nutrients in a semi-intensive shrimp farm in north-
32 western Mexico. Mar Pollut Bull 34:290-297.
- 33 Páez-Osuna F (2001a) The environmental impact of shrimp aquaculture: causes, effects,
34 and mitigating alternatives. Environ Manage 28:131–140.

- 1 Páez-Osuna F (2001b) The environmental impact of shrimp aquaculture: a global
2 perspective. Environ Pollut 112:229– 231.
- 3 Pelletier MC, Campbell DE, Ho KT, Burgess RM, Audette CT, Detenbeck NE (2011)
4 Can sediment total organic carbon and grain size be used to diagnose organic
5 enrichment in estuaries? Environ Toxicol Chem 30:538–547.
- 6 Pinto AMTP, Hirdes IM, Sanches-Filho CJ (2013) Determinação de metais pesados nos
7 camarões (*Farfantepenaeus paulensis*) consumidos na cidade de Pelotas-RS.
8 Ecotoxicol. Environ. Contam 8:129-134.
- 9 Rainbow PS (1988) The significance of trace metal concentrations in decapods. Symp
10 Zool Soc Lond 59:291-313.
- 11 Rainbow PS, Amiard-Triquet C, Amiard JC, Smith BD, Best SL, Nassiri Y, Langston
12 WJ (1999) Trace metal uptake rates in crustaceans (amphipods and crabs) from
13 coastal sites in NW Europe differentially enriched with trace metals. Mar Ecol
14 Prog Ser 183:189-203.
- 15 Ritvo G, Dixon JB, Lawrence AL, Samocha TM, Neill WH, Speed ME (1998)
16 Accumulation of Chemical Elements in Texas Shrimp Pond Soils. J World
17 Aquac Soc 29:422–431.
- 18 Russell M, Robinson CD, Walsham P, Webster L, Moffat CF (2011) Persistent organic
19 pollutants and trace metals in sediments close to Scottish marine fish farms.
20 Aquaculture 319:262-271.
- 21 Salomons W, Förstner U (1984) Metals in the hidrocycle. Berlin Springer Verlag.
- 22 Santos IR, Baisch P, Lima GTNP (2003) Metais pesados em sedimentos superficiais da
23 Lagoa Mirim, fronteira Brasil-Uruguai. Geochimica brasiliensis 17:37-47.
- 24 Sanares RC, Katase SA, Fast AW, Carpenter KE (1986) Water quality dynamics in
25 brackishwater shrimp ponds with artificial aeration and circulation. In: Maclean
26 JL, Dizon LB, Hosillos LV (eds). The First Asian Fisheries Forum. Asian
27 Fisheries Society, Manila, Philippines, p 83–86.
- 28 Smith PT (1996) Physical and chemical characteristics of sediments from prawn farms
29 and mangrove habitats on the Clarence River, Australia. Aquaculture 146:47–83.
- 30 Sokal RR, Rohlf FJ (1995) Biometry: The Principles and Practice of Statistics in
31 Biological Research, 3rd Edition. W. H. Freeman and Co., New York. p 887.
- 32 Steeby JA, Hargreaves JA, Tucker CS, Kingsbury S (2004) Accumulation, organic
33 carbon and dry matter concentration of sediment in commercial channel catfish
34 ponds. Aquac Eng 30:115–126.

- 1 Suguio K (1973) Introdução a sedimentologia. São Paulo. Ed. Edgard Blucher. EDUSP,
2 p 317.
- 3 Suplee MW, Cotner JB (1996) Temporal changes in oxygen demand and bacterial
4 sulfate reduction in inland shrimp ponds. Aquaculture 145:141–158.
- 5 Sutherland TF, Petersen SA, Levings, CD, Martin, AJ (2007) Distinguishing between
6 natural and aquaculture-derived sediment concentrations of heavy metals in the
7 Broughton Archipelago, British Columbia. Mar Pollut Bull 54:1451–1460.
- 8 Tacon AGJ, Forster IP (2003) Aquafeeds and the environment: policy implications.
9 Aquaculture 226:181–189.
- 10 Vinatea L, Malpartida J, Andreatta ER (2006) Caracterização do ph, carbono orgânico e
11 potencial redox de solos de viveiros de cultivo semi-intensivo do camarão
12 marinho *Litopenaeus vannamei*. B. Inst. Pesca 32:25-30.
- 13 Zimmermann CF, Keefe CW, Basche J (1997) Determination of carbon and nitrogen in
14 sediments and particulates of estuarine/coastal waters using elemental analysis.
15 U.S. Environmental Protection Agency, Method 440.0, p.9.
- 16 Wallner-Kersanach M, Andrade CFF, Milani MR, Niencheski LFH (2009) In situ
17 measurement of trace metals in estuarine waters of the Patos Lagoon using the
18 diffusive gradient in thin film (DGT). J Braz Chem Soc 20:333-340.
- 19 Wei TL, Yang WL, Lai ZN, Zhang Q and Liu M (2002) Residues of Heavy Metals in
20 Economic Aquatic Animal Muscles in Pearl River Estuary, South China. J Fish
21 Sci China 9:172–176.
- 22 Wu XY, Yang YF (2011) Heavy metal (Pb, Co, Cd, Cr, Cu, Fe, Mn and Zn)
23 concentrations in harvest-size white shrimp *Litopenaeus vannamei* tissues from
24 aquaculture and wild source. J Food Comp Anal 24:62–65.
- 25

1 **DISCUSSÃO GERAL**

2
3 Dentre os parâmetros abióticos avaliados, espacialmente o lançamento dos
4 efluentes causou um efeito agudo e pontual. Nitrogênio amoniacal total (NAT), clorofila
5 *a* e turbidez da água foram os parâmetros que sofreram alterações mais evidentes após a
6 descarga dos efluentes. Concentrações semelhantes as encontradas no presente estudo
7 de compostos nitrogenados e fosfatados (Kantin & Baumgarten 1982; Briggs & Funge-
8 Smith 1994; Abreu et al. 1995; Burford 1997; Cardozo et al. 2011) e clorofila *a* (Abreu
9 et al. 2010; Cardozo et al. 2011) são reportadas para o estuário da Lagoa dos Patos e
10 para proximidades de cultivos de camarões. Apesar das alterações observadas nos
11 parâmetros de qualidade de água, essas ocorreram apenas nos primeiros dias após a
12 descarga dos efluentes (até a amostragem 5 PD) e ficaram restritas até os primeiros 20m
13 de distância do canal de lançamento dos efluentes. Ao longo do tempo amostral, os
14 parâmetros abióticos também se mostraram pouco variáveis, com exceção da
15 temperatura e da salinidade, fatores mais susceptíveis a sofrerem variações em
16 ambientes estuarinos.

17 A hidrodinâmica local e a capacidade suporte do ambiente podem ter sido de
18 suma importância para garantir o baixo tempo de residência do material exportado dos
19 viveiros. A Lagoa dos Patos é caracterizada pela hidrologia forçada pelas relações entre
20 vento e descarga fluvial (Kjerfve 1986; Möller & Fernandes 2010), favorecendo as
21 trocas de água e o baixo tempo de residência induzidos pelos ventos predominantes de
22 NE (Möller et al. 1996). A baixa salinidade observada na amostragem 1 PD pode ser um
23 indicativo de fluxo de vazante derivado do aumento da descarga fluvial (Möller &
24 Fernandes 2010), favorecendo a rápida dispersão do material.

25 A comunidade fitoplânctonica e protozooplânctonica apresentaram um padrão
26 semelhante espaço-temporalmente, onde foi observado um acréscimo na abundância
27 desses organismos na amostragem 1 PD. A única exceção foi a classe das clorofíceas,
28 que se mantiveram em densidades estáveis ao longo de todo o período amostral, sem
29 apresentar oscilações marcadas. Esse aumento ocorrido no primeiro dia pós-descarga
30 pode estar relacionado ao *input* de nutrientes ocasionado pelas chuvas, e não
31 necessariamente a um incremento da produtividade primária ou um possível
32 espalhamento da pluma dos efluentes por todo o local amostral.

33 Abreu et al. (2010) encontraram uma relação significativa entre os valores
34 médios anuais de clorofila *a* e a quantidade de chuva na região estuarina, mostrando que

1 esse fator tem grande influência sobre essa comunidade. A abundância e diversidade de
2 ciliados tem sido utilizada como indicador da qualidade de água e dinâmica de
3 ecossistemas (Foissner 1988), uma vez que esses organismos exercem um importante
4 papel no fluxo de energia (Sherr & Sherr 1988; Decamp et al. 2003). Esses organismos
5 servem de alimento para larvas de peixes, exercendo uma função essencial no link entre
6 o *microbial loop* e os níveis tróficos superiores (Fukami et al. 1999). Yang et al. (2012)
7 demonstraram que a comunidade e o tamanho da estrutura do protozooplâncton
8 geralmente segue a dinâmica espacial e o tamanho da estrutura do fitoplâncton, fato que
9 pode ser observado no presente estudo.

10 Com exceção da amostragem 30 PD, a concentração zooplanctônica foi baixa e
11 sofreu poucas oscilações espaço-temporalmemente, com algumas poucas diferenças
12 estatísticas ($p < 0,05$) observadas entre os pontos amostrais. Baixas concentrações
13 zooplanctônicas são reportadas na literatura para a região também por Cardozo et al.
14 (2011). A ocorrência e abundância de espécies zooplanctônicas no estuário da Lagoa
15 dos Patos é determinada principalmente por variações sazonais de salinidade,
16 temperatura, direção e intensidade do vento, descarga de água doce e oferta de
17 alimentos (McLaren & Corkett 1981; Montú et al. 1998; Muxagata et al. 2012). Apesar
18 de a clorofila *a* poder ser usada como indicador de oferta de alimento para copépodos
19 (Hirst & Bunker 2003), as concentrações constantes de cloroficeas e clorofila *a*
20 demonstram que a ação de herbivoria não foi marcante. Dentre os parâmetros abióticos
21 avaliados não foi possível encontrar uma relação para o aumento de densidade ocorrido
22 na amostragem 30 PD.

23 Além das variáveis ambientais favoráveis a dispersão do material exportado
24 pelos efluentes, o manejo alimentar e o uso de tecnologia adequados podem limitar o
25 impacto ambiental da carcinocultura (Alonso-Rodríguez & Páez-Osuna 2003). Em áreas
26 rasas com baixa capacidade de diluição, os resíduos provenientes da atividade poderão
27 sedimentar próximo ao local de descarga dos efluentes, maximizando o potencial
28 poluidor dos efluentes. Em sistemas de cultivo extensivos as descargas de efluentes são
29 escassas, diferente do que acontece em sistemas semi-intensivos, onde quantidades mais
30 significativas são descarregadas (Alonso-Rodríguez & Páez-Osuna 2003). As dietas
31 artificiais e os fertilizantes orgânicos e inorgânicos são as principais fontes de *input* de
32 nutrientes em sistemas de criação de camarões, o que causa reflexo direto na
33 composição dos efluentes. Do total de N e P que entram em um sistema semi-intensivo
34 de cultivo de camarões, 76% do N e 83,4% do P são provenientes da ração (Páez-Osuna

1 et al. 1997). Desse total, apenas 25-30% do N e P aplicados na forma de rações e
2 fertilizantes são retirados na forma de camarões ao final do ciclo (Boyd & Tucker
3 1998). Dessa forma, geralmente altas concentrações de sólidos suspensos orgânicos,
4 carbono, nitrogênio e fósforo proveniente do alimento não consumido e dos produtos de
5 excreção estão presentes nos viveiros de cultivo e consequentemente nos efluentes
6 (Burford & Williams 2001), favorecendo um aumento da produtividade natural do
7 corpo d'água receptor (Tacon & Forster 2003).

8 O percentual de carbono orgânico total (COT) variou entre 0,12 e 0,67% e o
9 percentual de nitrogênio orgânico total (NT) manteve-se < 0,07%. O carbono orgânico é
10 proveniente do fitoplâncton morto, dos fertilizantes orgânicos, da ração não consumida
11 e das fezes dos animais misturados com as partículas do solo (Olsen et al. 2008). As
12 baixas densidades evitam que grandes *inputs* de nutrientes sejam inseridos através da
13 ração, e o manejo adequado da qualidade da água dos viveiros impedem *blooms* algais.

14 Os valores de COT encontrados no presente estudo estão abaixo dos reportados
15 na literatura para viveiros de criação de camarões (Smith 1996; Vinatea et al. 2006;
16 Boyd et al. 2010) e para o estuário da Lagoa dos Patos (Santos et al. 2003). As baixas
17 concentrações de NT podem ser um indicativo de que não ocorreu excesso de matéria
18 orgânica para ser decomposta no meio. Estudos têm demonstrado que a comunidade
19 microbiana (Burford & Williams 2001), o fitoplâncton (Hargreaves 1998; Burford &
20 Glibert 1999) e o uso contínuo de aeradores de pás ao longo do ciclo produtivo (Sanares
21 et al., 1986) desempenham um importante papel na remoção de compostos nitrogenados
22 em viveiros de cultivo. No entanto, outros estudos têm demonstrado que o efluente do
23 cultivo de camarões pode aportar quantidades significativas de nitrogênio no sedimento
24 (Lemonnier & Faninoz 2006; Anh et al. 2010). A granulometria do sedimento é outro
25 fator determinante no conteúdo orgânico de sedimentos estuarinos (Pelletier et al.
26 2011). O teor predominantemente arenoso e fino encontrado no presente estudo
27 juntamente com a aeração constante da água durante o ciclo de produção, podem ter
28 auxiliado na decomposição da matéria orgânica evitando assim o acúmulo de carbono
29 no sedimento. Sutherland et al. (2007) encontraram uma forte correlação entre a
30 porosidade do sedimento e o conteúdo orgânico, onde sedimentos arenosos foram
31 caracterizados por apresentarem baixo conteúdo orgânico.

32 A bacia de sedimentação presente na fazenda, é uma tecnologia que age na
33 melhora da qualidade da água dos efluentes no que diz respeito ao transporte dos
34 sólidos suspensos totais, ciclagem de nutrientes, produção de biomassa, absorção de

1 nutrientes pelas plantas e animais e distribuição da matéria orgânica e oxigênio (Shpigel
2 et al., 2013). Vários processos bióticos e abióticos regulam a remoção de poluentes
3 nesses locais, como mineralização microbiana, nitrificação-desnitrificação, absorção por
4 macrófitas, precipitação química, sedimentação e adsorção pelo substrato (Lin et al.
5 2005). Jackson et al. (2003) reportam uma redução da carga dos efluentes de viveiros de
6 cultivo de camarões de 60% do SST, 23% do nitrogênio total e 35% do fósforo total. A
7 correta escala da bacia de sedimentação pode auxiliar na retenção de fósforo e amônia
8 dos viveiros reduzindo os possíveis efeitos sobre a comunidade planctônica (Cardozo &
9 Odebrecht. 2012). Nesse sentido, a bacia pode ter agido de forma positiva no controle
10 do potencial poluidor dos efluentes.

11 A macrofauna bentônica apresentou pequena variabilidade espacial em ambas as
12 campanhas amostrais (2012 e 2013). Já ao longo do tempo, essa variabilidade foi
13 bastante marcada ao longo das coletas de 2012 e menos variável nas coletas de 2013.
14 Em 2012, maior densidade e maior riqueza de espécies foram observadas nos meses
15 mais frios comparativamente aos meses mais quentes, situação oposta à que ocorreu
16 durante a campanha amostral de 2013. Bemvenuti (1987) e Rosa & Bemvenuti (2006)
17 reportam aumento da densidade do macrozoobentos no verão quando comparado ao
18 inverno, assim como ocorreu em 2013. Isso porque em períodos de altas temperaturas
19 ocorre aumento da atividade reprodutiva e consequente controle populacional através de
20 predadores infaunais e baixas taxas de recrutamento e consequente baixas taxas de
21 predação nos meses mais frios (Bemvenuti, 1998). Por outro lado, mudanças nos
22 padrões de temperatura e salinização do estuário pode causar o insucesso dos
23 recrutamentos. Existe uma forte influência entre padrões de abundância do macrobentos
24 e flutuações de salinidade em ambientes estuarinos (Holland et al., 1987). Ambientes
25 poluídos com acúmulo de matéria orgânica reduzem a diversidade bentônica (Lorenzen
26 et al., 1987), favorecendo a recolonização por organismos oportunistas após tais eventos
27 de perturbação (Rosa & Bemvenuti, 2006). Esse acúmulo pode tornar os sedimentos
28 ácidos e pobres em oxigênio, facilitando as vias de decomposição anaeróbicas e
29 aumentando a produção e liberação de compostos tóxicos reduzidos, o que pode alterar
30 por fim a estrutura da comunidade bentônica (Jackson et al., 2004).

31 Em fato, o enriquecimento orgânico pode causar mudanças bruscas na estrutura
32 da comunidade reduzindo a assembléia a algumas poucas espécies tolerantes,
33 diminuindo assim a densidade e a riqueza de espécies (Pearson & Rosenberg, 1978). O
34 *input* orgânico no sedimento leva a mudanças nos parâmetros físicos e químicos

1 (Schaanning 1994) o que pode gerar efeitos diretos e indiretos na comunidade faunal
2 (Pearson & Rosemberg 1978). Os macroinvertebrados bentônicos podem ser usados
3 como indicadores das modificações e variabilidade em ecossistemas estuarinos uma vez
4 que são relativamente sedentários, tem vida longa, as diferentes espécies exibem
5 tolerâncias ao stress, tem um importante papel na ciclagem de nutrientes e materiais e
6 no link de transferência de energia para níveis tróficos superiores (Dauvin, 2007). Os
7 resultados desse estudo demonstram que a riqueza de espécies sofreu pouca variação, o
8 que corrobora com os baixos níveis orgânicos encontrados no sedimento.

9 Observou-se durante as campanhas amostrais (2012 e 2013) que a densidade do
10 macrozoobentos foi predominantemente constituída por poliquetas. Esses organismos
11 representam um importante papel no funcionamento das comunidades bentônicas
12 (Hutchings, 1998) e são bastante utilizados como indicadores de enriquecimento
13 orgânico (Ansari et al., 1986). As espécies mais abundantes foram *Alitta succinea*,
14 *Heteromastus similis* e *Laeonereis acuta*. O hábito dessas espécies permite a
15 manutenção de densidades elevadas através de mecanismos de escape a predação, como
16 capacidade de enterramento, mobilidade e estratégias reprodutivas que garantem uma
17 rápida recolonização do ambiente após perturbações. Esses organismos não sofreram
18 redução na densidade após o lançamento dos efluentes. Segundo Bemvenuti et al.
19 (1997), a abundância dessas espécies tende a diminuir consideravelmente após eventos
20 de distúrbios ambientais.

21 O conteúdo de Cu ($0.12 - 1.27 \mu\text{g g}^{-1}$) e Zn ($0.51 - 3 \mu\text{g g}^{-1}$) lábeis encontrados no
22 presente estudo foram baixos quando comparados a outros trabalhos. Um estudo
23 realizado em diversos locais do estuário da Lagoa dos Patos indicou valores médios de
24 $4,5 - 19,2 \mu\text{g g}^{-1}$ de Cu e $18,5 - 62,5 \mu\text{g g}^{-1}$ de Zn, ambos na fração lábil no sedimento
25 de fração $<0,63 \mu\text{m}$ (silte e argila), em uma área com baixo impacto antrópico. (Costa et
26 al., dados não publicados). No que diz respeito à fração de Cu e Zn mais adsorvida ao
27 sedimento ($0,3 - 2.65 \mu\text{g g}^{-1}$ e $30,44 - 121,4 \mu\text{g g}^{-1}$, respectivamente), Russel et al. (2011)
28 avaliando as concentrações de Cu e Zn no sedimento próximo de fazendas marinhas de
29 peixes na Escócia, encontraram valores médios de $35,8$ e $89,1 \mu\text{g g}^{-1}$, respectivamente.
30 Já Smith (1996) avaliando cultivos de *Penaeus monodon* reporta concentrações de até
31 $45 \mu\text{g g}^{-1}$ de Cu e até $85 \mu\text{g g}^{-1}$ de Zn nos sedimentos de viveiros e concentrações abaixo
32 de $2 \mu\text{g g}^{-1}$ de Cu e $64 \mu\text{g g}^{-1}$ de Zn no sedimento dos efluentes. Ritvo et al. (1998)
33 avaliando a quantidade de Cu e Zn em um sistema de cultivo de camarões observaram
34 um desequilíbrio negativo entre as quantidades que foram adicionados através da ração

1 e dos fertilizantes e as quantidades encontradas no solo, sugerindo que esses elementos
2 podem ter sido imobilizados pela conversão para forma orgânica.

3 A legislação brasileira não possui regulamento específico para as concentrações
4 máximas permitidas desses metais traço em instalações aquícolas. A Resolução
5 CONAMA 454/2012, que estabelece as diretrizes para o gerenciamento de material a
6 ser dragado em águas salobras sob jurisdição nacional, reporta como limiar abaixo do
7 qual há menor probabilidade de efeitos adversos a biota, concentrações de $34\mu\text{g g}^{-1}$ de
8 Cu e $150\mu\text{g g}^{-1}$ de Zn. Já a Resolução CONAMA 420/2009, que dispõe valores
9 orientadores menos restritivos quanto a qualidade de solo, apresenta valores de
10 prevenção $60\mu\text{g g}^{-1}$ de Cu e $300\mu\text{g g}^{-1}$ de Zn em solos. Isso demonstra que os resultados
11 aqui obtidos encontram-se abaixo do limiar permitido nas resoluções vigentes.

12 A análise dos teores de Cu e Zn no músculo em base seca, resultou em uma
13 concentração média de $6,63 \pm 0,2\mu\text{g g}^{-1}$ de Cu e $19,76 \pm 0,2\mu\text{g g}^{-1}$ de Zn nos animais do
14 Viveiro 1 e, $7,6 \pm 0,51\mu\text{g g}^{-1}$ de Cu e $19,13 \pm 0,32\mu\text{g g}^{-1}$ de Zn nos animais do Viveiro 2
15 ($n=6$). Pinto et al. (2013) quantificaram os metais presentes no camarão-rosa
16 *Farfantepenaeus paulensis* proveniente da pesca extrativa no estuário da Lagoa dos
17 Patos, e até $12,9\mu\text{g g}^{-1}$ de Cu e $32,3\mu\text{g g}^{-1}$ de Zn foram detectados no tecido muscular.
18 Outro estudo avaliando as concentrações de Cu e Zn no tecido do camarão *L. vannamei*
19 proveniente de cultivos intensivos na China resultou em valores médios de $24,26\mu\text{g g}^{-1}$
20 de Cu e $171,56\mu\text{g g}^{-1}$ de Zn (Wu & Yang, 2011). As concentrações encontradas no
21 presente estudo estão bem abaixo do permitido pela legislação nacional e do encontrado
22 na média internacional. Alguns crustáceos decápodes são capazes de regular a
23 concentração corpórea de metais como Cu e Zn, disponibilizando o necessário para suas
24 necessidades bioquímicas e detoxificando o excedente (Rainbow, 1988; Rainbow et al.,
25 1999).

26 De maneira geral, os resultados obtidos neste estudo sugerem que o cultivo
27 semi-intensivo do camarão *L. vannamei* causou um impacto agudo e pontual em alguns
28 dos parâmetros de qualidade de água avaliados, uma vez que as alterações não
29 excederam a capacidade suporte do ambiente e o sistema assimilou em um curto espaço
30 de tempo (1 a 5 dias) o efluente liberado. Não foi observado impacto do lançamento dos
31 efluentes sobre o fitoplâncton, protozooplâncton, mesozooplâncton, macrozoobentos, ou
32 mesmo sobre a concentração de COT, NT, Cu e Zn no sedimento (frações láveis e mais
33 fortemente adsorvida), assim como do Cu e Zn no tecido dos camarões. As diferenças
34 observadas nos parâmetros abióticos avaliados foram decorrentes de oscilações naturais

que ocorrem em sistemas estuarinos. Dessa forma, pode-se concluir que o ciclo de produção de camarão *L. vannamei* nas condições de cultivo que está sendo aplicada demonstra ótimas condições de manejo, por não gerar de forma significativa efluentes com alta carga orgânica ou metais pesados para o ambiente natural.

5

1 **REFERÊNCIAS BIBLIOGRÁFICAS**

2

- 3 ABREU PC, M BERGESCH, LA PROENÇA & C ODEBRECHT. 2010. Short- and
4 long- term chlorophyll a variability in the shallow microtidal Patos Lagoon
5 estuary, Southern Brazil. *Estuar. Coast.*, 33: 554-569.
- 6 ANH, PT, C KROEZE, SR BUSH & ARTHUR PJM. 2010. Water pollution by
7 intensive brackish shrimp farming in southeast Vietnam: Causes and options
8 for control. *Agr. Water Manag.*, 97: 872–882.
- 9 ALONSO-RODRÍGUES, R & F PÁEZ-OSUNA. 2003. Nutrients, phytoplankton and
10 harmful algal blooms in shrimp ponds : a review with special reference to the
11 situation in the Gulf of California. *Aquaculture* 219: 317–336.
- 12 ANSARI, ZA. BS INGOLE & AH PARULEKAR. 1986. Effect of high organic
13 enrichment of benthic polychaete population in an estuary. *Mar. Pollut. Bull.*
14 17: 361- 365.
- 15 BEMVENUTI, CE. 1987. Predation effects on a benthic community in estuarine soft
16 sediments. *Atlântica* 9: 33– 63.
- 17 BEMVENUTI, CE. 1997. Benthic Invertebrates. In: Seeliger, U.; Odebrecht, C. &
18 Castello, J.P. (ed.) Subtropical convergence environments. The coast and sea in
19 the southwestern Atlantic. Springer-Verlag. 43-46 p.
- 20 BEMVENUTI, CE. 1998. In: SEELIGER, U.; ODEBRECHT, C.; CASTELLO, J.P.
21 (Eds.). Os Ecossistemas Costeiro e Marinho do Extremo Sul do Brasil. Rio
22 Grande, Editora Ecoscientia. 46-51 p.
- 23 BOYD, CE & CS TUCKER. 1998. Pond Aquaculture Water Quality Management.
24 Kluwer Academic Publishers, Boston, MA. 700 pp.
- 25 BOYD, CE, CW WOOD, PL CHANEY & JF QUEIROZ. 2010. Role of aquaculture
26 pond sediments in sequestration of annual global carbon emissions. *Environ.*
27 *Pollut.*, 158: 2537–2540.
- 28 BRASIL. Resolução CONAMA – Conselho Nacional do Meio Ambiente. Resolução nº
29 420/2009. Dispõe sobre critérios e valores orientadores de qualidade do solo
30 quanto à presença de substâncias químicas e estabelece diretrizes para o
31 gerenciamento ambiental de áreas contaminadas por essas substâncias em
32 decorrência de atividades antrópicas. Diário Oficial da República Federativa do
33 Brasil, Brasília, DF 31 de dezembro de 2009.

- 1 BRASIL. Resolução CONAMA – Conselho Nacional do Meio Ambiente. Resolução nº
2 454/2012. Estabelece as diretrizes gerais e os procedimentos referenciais para o
3 gerenciamento do material a ser dragado em águas sob jurisdição nacional.
4 Brasília, DF, 01 nov. 2012.
- 5 BRIGGS, MRP & SJ FUNGE-SMITH. 1994. A nutrient budget of some intensive
6 marine shrimp ponds in Thailand. *Aquac. Fish. Manag.*, 25:789–811.
- 7 BURFORD, M. 1997. Phytoplankton dynamics in shrimp ponds. *Aquac. Res.*, 28: 351–
8 360.
- 9 BURFORD, MA & PM GLIBERT. 1999. Short-term nitrogen uptake and regeneration
10 in early and late growth phase shrimp ponds. *Aquac. Res.*, 30: 215–227.
- 11 BURFORD, MA & KC WILLIAMS. 2001. The fate of nitrogenous waste from shrimp
12 feed. *Aquaculture* 198: 79-93.
- 13 CARDOZO, AP, VO BRITTO & C ODEBRECHT. 2011. Temporal variability of
14 plankton and nutrients in shrimp culture ponds vs. adjacent estuarine water.
15 *Pan Am. J. Aq. Sciences* 6: 28–43.
- 16 CARDOZO, AP & C ODEBRECHT. 2012. Effects of shrimp pond water on
17 phytoplankton: importance of salinity and trophic status of the receiving
18 environment. *Aquac. Res.*, 1–11.
- 19 DAUVIN, JC. 2007. Paradox of estuarine water quality: benthic indicators and indices,
20 consensus or debate for the future. *Mar. Pol. Bull.*, 55: 271-281.
- 21 DECAMP, O, J CODY, L CONQUEST, G DELANOY, & AGJ TACON. 2003. Effect
22 of salinity on natural community and production of *Litopenaeus vannamei*
23 (Boone) within experimental zero-water exchange culture systems. *Aquac.*
24 *Res.*, 34:345-355.
- 25 FOISSNER W. 1988. Taxonomic and nomenclatural revision of Sladeczek's list of
26 ciliates (Protozoa: Ciliophora) as indicators of water quality. *Hydrobiologia*
27 166: 1-64.
- 28 FUKAMI, K, A WATANABE, S FUJITA, K YAMAOKA & T NISHIJIMA. 1999.
29 Predation on naked protozoan microzooplankton by fish larvae. *Mar. Ecol-*
30 *Prog Ser.*, 185: 285-291.
- 31 HARGREAVES, JA. 1998. Nitrogen biogeochemistry of aquaculture ponds.
32 *Aquaculture* 168: 181–212.

- 1 HOLLAND, AF, AT SHAUGHNESSY & MH HIEGEL. 1987. Long-term variation in
2 mesohaline Chesapeake Bay macrobenthos: Spatial and temporal patterns.
3 Estuaries 10: 227–245.
- 4 HUTCHINGS, P. 1998. Biodiversity and functioning of polychaetes in benthic
5 sediments. Biodivers. Conserv., 7: 1133–1145.
- 6 HIRST, AG & AJ BUNKER. 2003. Growth of marine planktonic copepods: global rates
7 and patterns in relation to chlorophyll a, temperature, and body weight.
8 Limnol. Oceanogr., 48: 1988–2010.
- 9 JACKSON, C, N PRESTON & PJ THOMPSON. 2004. Intake and discharge nutrient
10 loads at three intensive shrimp farms. Aquac. Res., 35: 1053–1061.
- 11 KANTIN, R & MGZ BAUMGARTEN. 1982 Observações hidrográficas no estuário da
12 Lagoa dos Patos: distribuição e flutuações dos sais nutrientes. Atlântica 5: 76–
13 92.
- 14 KJERFVE, B. 1986 Comparative oceanography of coastal lagoons. In Estuarine
15 Variability (Wolfe, D. A., ed.). Academic Press, New York, pp. 63-81.
- 16 LEMONNIER, H & S FANINOZ. 2006. Effects of water exchange rate on effluent and
17 sediment characteristics and on partial nitrogen budget in semi-intensive
18 shrimp ponds in New Caledonia. Aquac. Res., 37: 938–948.
- 19 LIN, YF, SR JING, DY LEE, YF CHANG, YM CHEN & KC SHIH. 2005.
20 Performance of a constructed wetland treating intensive shrimp aquaculture
21 wastewater under high hydraulic loading rate. Environ. Pollut. 134: 411–421.
- 22 LORENZEN, S, M PREIN & C VALENTIN. 1987. Mass aggregations of the free-
23 living marine nematode *Pontonema vulgare* Oncholaimidae in organically
24 polluted fjords. Mar. Ecol-Prog. Ser., 37: 27-34.
- 25 MCLAREN, IA & CJ CORKETT. 1981. Temperature-dependent growth and
26 production by a marine copepod. Can. J. Fish. Aquat. Sci., 38: 77–83.
- 27 MÖLLER, OO, JA LORRENZZENTI, JL STECH & MM MATA. 1996. The Patos
28 Lagoon summertime circulation and dynamics. Cont. Shelf Res., 16: 335-351.
- 29 MÖLLER, O & E FERNANDES. 2010. In: SEELIGER, U & C ODEBRECHT (eds.) O
30 estuário da Lagoa dos Patos: Um século de transformações. FURG, Rio
31 Grande, 17-30 p.
- 32 MUXAGATA, E, WJA AMARAL & CN BARBOSA. 2012. *Acartia tonsa* production
33 in the Patos Lagoon estuary, Brazil. ICES J. Mar. Sci., 69: 475-482.

- 1 OLSEN, LM, M HOLMER, Y OLSEN. 2008. Perspectives of nutrient emission from
2 fish aquaculture in coastal waters Literature review with evaluated state of
3 knowledge FHF project no. 542014. The Fishery and Aquaculture Industry
4 Research Fund.
- 5 PÁEZ-OSUNA, F, SR GUERRERO-GALVÁN, AC RUIZ-FERNANDEZ & R
6 ESPINOZA-ÂNGULO. 1997. Fluxes and mass balances of nutrients in a semi-
7 intensive shrimp farm in northwestern Mexico. Mar. Pollut. Bull., 34: 290–297.
- 8 PEARSON, TH & R ROSENBERG. 1978. Macrobenthic succession in relation to
9 organic enrichment and pollution of the marine environment. Oceanogr. Mar.
10 Biol. A. Rev., 16: 229–311.
- 11 PELLETIER, MC, DE CAMPBELL, KT HO, RM BURGESS, CT AUDETTE & NE
12 DETENBECK. 2011. Can sediment total organic carbon and grain size be used
13 to diagnose organic enrichment in estuaries? Environ. Toxicol. Chem., 30:
14 538–547.
- 15 PINTO, AMTP, IM HIRDES & PJ SANCHES-FILHO. 2013. Determinação de metais
16 pesados nos camarões (*Farfantepenaeus paulensis*) consumidos na cidade de
17 Pelotas-RS. Ecotoxicol. Environ. Contam., 8: 129-134.
- 18 RAINBOW, PS. 1988. The significance of trace metal concentrations in decapods.
19 Symp. Zool. Soc. Lond., 59: 291-313.
- 20 RAINBOW, PS, C AMIARD-TRIQUET, JC AMIARD, BD SMITH, SL BEST, Y
21 NASSIRI & WJ LANGSTON. Trace metal uptake rates in crustaceans
22 (amphipods and crabs) from coastal sites in NW Europe differentially enriched
23 with trace metals. Mar. Ecol-Prog. Ser., 183: 189-203.
- 24 RITVO, G, JB DIXON, AL LAWRENCE, TM SAMOCHA, WH NEILL & ME
25 SPEED. 1998. Accumulation of Chemical Elements in Texas Shrimp Pond
26 Soils. J. World Aquacult. Soc., 29: 422–431.
- 27 ROSA, LC & CE BEMVENUTI. 2006. Temporal variability of the estuarine
28 macrofauna of the Patos Lagoon, Brazil. Revista de Biología Marina y
29 Oceanografía 41: 1–9.
- 30 RUSSELL, M, CD ROBINSON, P WALSHAM, L WEBSTER & CF MOFFAT. 2011.
31 Persistent organic pollutants and trace metals in sediments close to Scottish
32 marine fish farms. Aquaculture 319: 262-271.
- 33 SANARES, RC, SA KATASE, AW FAST & KE CARPENTER. 1986. Water quality
34 dynamics in brackishwater shrimp ponds with artificial aeration and

- 1 circulation. In: Maclean, J.L., Dizon, L.B., Hosillos, L.V. Eds. The First Asian
2 Fisheries Forum. Asian Fisheries Society, Manila, Philippines, pp. 83–86.
- 3 SANTOS, IR, P BAISCH & GNTP LIMA. 2003. Metais pesados em sedimentos
4 superficiais da Lagoa Mirim, fronteira Brasil-Uruguai. Geochimica brasiliensis
5 17: 037-047.
- 6 SCHAAANNING, MT. 1994. Distribution of sediment properties in coastal areas
7 adjacent to fish farms and environmental evaluation of five locations
8 surveyed in October 1993. Norwegian Institute for Water Research (NIVA),
9 Report No. O-93205, O-93062. Oslo, Norway. 29 pp.
- 10 SHERR E & B SHERR. 1988. Role of microbes in pelagic food webs: a revised
11 concept. Limnol Oceanogr 33: 1225-1227.
- 12 SMITH, PT. 1996. Physical and chemical characteristics of sediments from prawn
13 farms and mangrove habitats on the Clarence River, Australia. Aquaculture
14 146: 47–83.
- 15 SUTHERLAND, TF, SA PETERSEN, CD LEVINGS & AJ MARTIN. 2007.
16 Distinguishing between natural and aquaculture-derived sediment
17 concentrations of heavy metals in the Broughton Archipelago, British
18 Columbia. Mar. Pollut. Bull., 54: 1451–1460.
- 19 TACON, AGJ & IP FORSTER. 2003. Aquafeeds and the environment: policy
20 implications. Aquaculture 226: 181–189.
- 21 VINATEA, L, J MALPARTIDA & ER ANDREATTA. 2006. Caracterização do ph,
22 carbono orgânico e potencial redox de solos de viveiros de cultivo semi-
23 intensivo do camarão marinho *Litopenaeus vannamei*. B. Inst. Pesca, São
24 Paulo, 32: 25-30.
- 25 WU, XY & YF YANG. 2011. Heavy metal (Pb, Co, Cd, Cr, Cu, Fe, Mn and Zn)
26 concentrations in harvest-size white shrimp *Litopenaeus vannamei* tissues from
27 aquaculture and wild source. J. Food Compos. Anal., 24: 62–65.
- 28 YANG, EJ, JH HYUN, D KIM, J PARK, SH KANG, HC SHIN & S LEE. 2012.
29 Mesoscale distribution of protozooplankton communities and their herbivory in
30 the western Scotia Sea of the Southern Ocean during the austral spring. J. Exp.
31 Mar. Biol. Ecol., 428: 5–15.
- 32

1 **ANEXO**

2



3
4 Canal de abastecimento dos viveiros.

5



6
7 Viveiro de cultivo no momento do esvaziamento prévio a despesca.
8



1
2 Momento da despesca.
3



4
5 Saída do efluente da bacia de sedimentação para o canal de lançamento.
6



1
2 Bacia de sedimentação seca.
3



4
5 Chegada do efluente no estuário e coleta de macrozoobentos ao fundo.
6



1
2 Encontro do canal de lançamento dos efluentes com o estuário.
3



4
5 Material de apoio utilizado durante as coletas.
6

1



2

Coleta de dados de oxigênio dissolvido e temperatura com auxílio de um oxímetro.

3

4



5

6

Momento da coleta do zooplâncton com auxílio de rede cilindro-cônica.



1
2 Coleta de macrozoobentos com auxílio de corer de PVC.
3



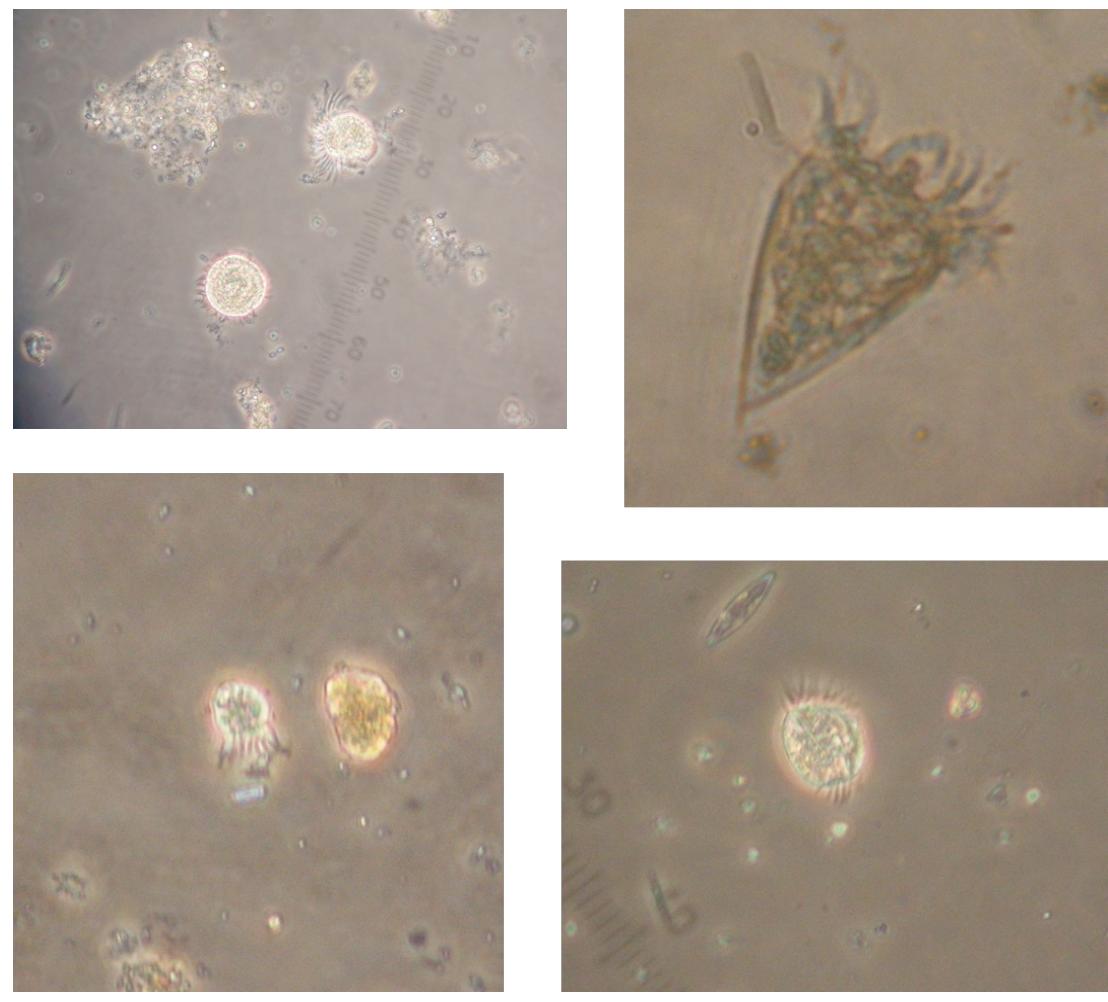
4
5 Peneiramento do macrozoobentos em tela com abertura de 500µm.



1
2 Amostra de fitoplâncton e protozooplâncton armazenada em frasco âmbar.
3

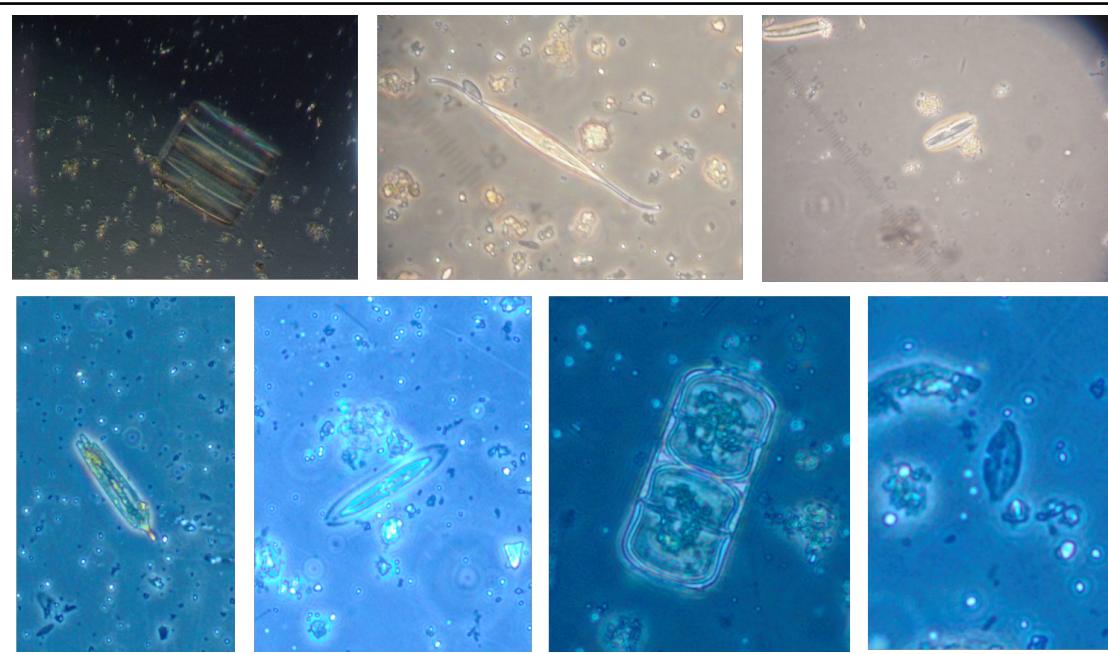


4
5 Material utilizado para identificação e captura de imagens de organismos
6 fitoplanctônicos e protozooplantônicos. Detalhe para o microscópio invertido.



1
2

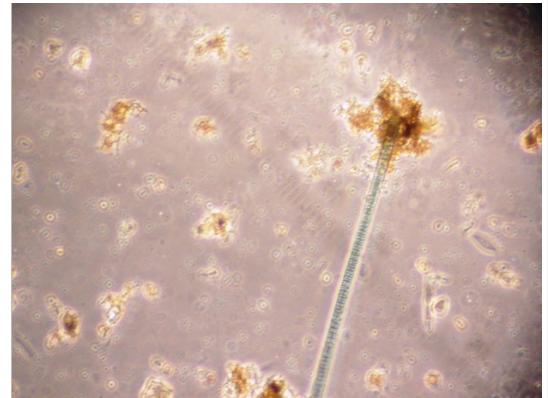
3 Espécimes de ciliados encontrados.
4



5
6

7 Espécimes de diatomáceas encontradas.
8

1



2

3

4

Espécimes de cianobactérias encontradas.

5



6

7

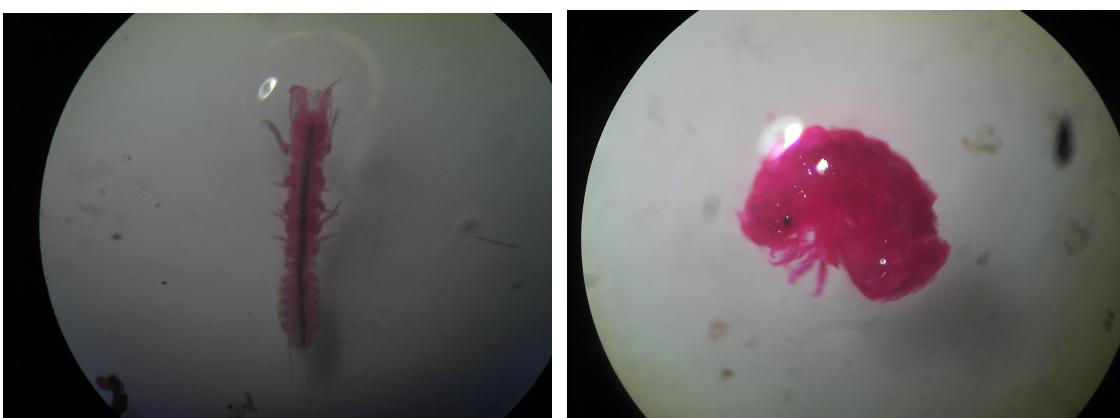
8

Amostras de macrozoobentos fixados em solução formaldeído 4% coradas com rosa de bengala, previamente a identificação.

9



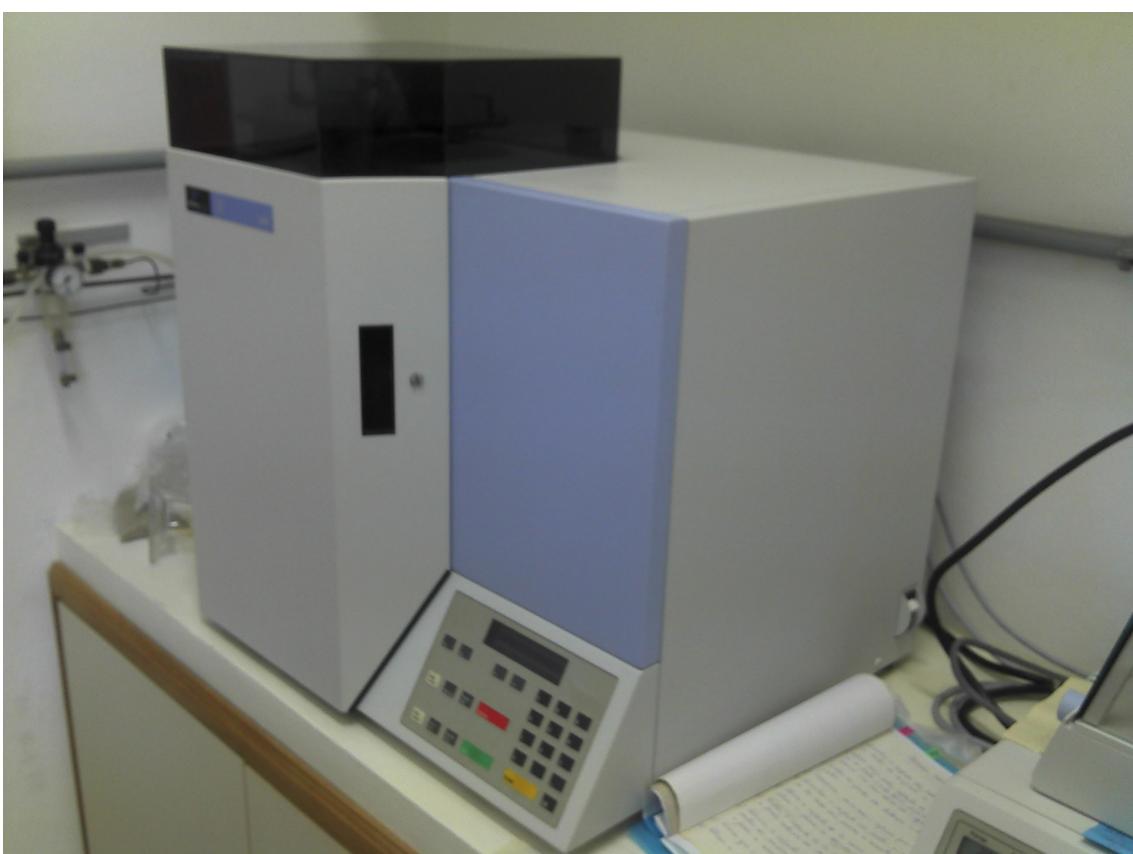
1
2 Lupa utilizada na identificação do macrozoobentos.
3



4
5 Exemplares de macrozoobentos identificados – *Kalliapseudes schubartii* a direita e
6 *Sphaeromopsis mourei* a esquerda.
7



1
2 Balança utilizada para pesagem de sedimento durante análise de C e N.
3



4
5 Analisador elementar CHNS/O Série 2400 da Perkin Elmer utilizado para determinação
6 do teor de C e N no sedimento.